



Department of
Environmental
Conservation

ROCHESTER EMBAYMENT AREA OF CONCERN LOSS OF FISH AND WILDLIFE HABITAT BENEFICIAL USE IMPAIRMENT REMOVAL REPORT

JUNE 2023

Kathy Hochul, Governor | Basil Seggos, Commissioner

Prepared by:

New York State Department of Environmental Conservation

and

Monroe County Department of Public Health

This Beneficial Use Impairment (BUI) Removal Report was prepared by the New York State Department of Environmental Conservation (NYSDEC) and the Monroe County Department of Public Health (MCDPH) and was substantially funded by the United States Environmental Protection Agency (USEPA) through the Great Lakes Restoration Initiative (GLRI). The NYSDEC and MCDPH have engaged stakeholders and the public, including the Remedial Advisory Committee, through the BUI removal process. For more information, please contact the Remedial Action Plan Coordinator at MCDPH or the AOC Coordinator at NYSDEC.

Rochester Embayment Area of Concern
Loss of Fish and Wildlife Habitat
Beneficial Use Impairment (BUI) Removal Report

June 2023

Prepared by:

New York State Department of Environmental Conservation

And

Monroe County Department of Public Health

This Beneficial Use Impairment (BUI) Removal Report was prepared by the New York State Department of Environmental Conservation (NYSDEC) and the Monroe County Department of Public Health (MCDPH) and was substantially funded by the United States Environmental Protection Agency (USEPA) through the Great Lakes Restoration Initiative (GLRI). The NYSDEC and MCDPH have engaged stakeholders and the public, including the Remedial Advisory Committee (RAC), through the BUI removal process. For more information, please contact the Remedial Action Plan Coordinator at MCDPH or the AOC Coordinator at NYSDEC.

Table of Contents

List of Abbreviations	5
1. Introduction and Report Purpose	7
2. Background and BUI Removal Criteria.....	8
2.1 BUI Removal Criteria.....	8
2.2 BUI Removal Considerations	9
2.3 Strategy and Rationale	9
3. BUI Indicator Status Resolution	10
3.1 Criterion 1: There is no net loss of acreage and quality of federal or state-designated wetlands, using 1996 as the baseline year for comparisons.....	10
3.1.2 Project Descriptions.....	13
3.1.3 Other Supporting Habitat Restoration Projects:	16
3.1.4 Monitoring of Restoration Projects.....	17
3.2 Criterion 2: There is no net loss of the existing 50-foot-wide buffer strip of trees and shrubs on both sides of NYSDEC classified streams and the Genesee River up to the Lower Falls (using 1999 as the baseline year for comparisons).	18
3.3 Criterion 3: Suspended sediment concentrations in the Genesee River remain less than 30 mg/L for at least 80% of a year, and exceed 200 mg/L for no more than five events with a combined duration of not greater than 20 days, as determined by a five-year average.....	19
3.3.1 Background.....	19
3.3.2 Water Quality Monitoring Program.....	19
3.3.3 Other Efforts Supporting Water Quality Improvements.....	24
3.4 Criterion 4: Hexagenia, or another appropriate indicator, is present in the Embayment and in suitable habitats in the Genesee River up to the Lower Falls; members of the stonefly, mayfly, and caddisfly families are present in streams.	25
3.4.1 Background.....	25
3.4.2 1999 – 2002 USGS Assessment	26
3.4.3 2013 NYSDEC/USGS Assessment	26
3.4.4. NYSDEC Rotating Integrated Basin Studies (RIBS) Monitoring.....	27
3.5 Criterion 5: Amphibian diversity and abundance in the study area (including the Genesee River up to the Lower Falls if monitoring can be performed safely) are comparable to expected standards for the type of habitat.....	31
3.5.1 Background.....	31
3.5.2 Amphibian diversity and abundance.....	31
3.6 Criterion 6: Lake sturgeon of different life stages inhabit the Genesee River up to the Lower Falls and the Embayment, OR physical and biological habitat are suitable for sturgeon.	35

3.6.1 Background.....	35
3.6.2 New York State Lake Sturgeon Recovery Plan.....	35
3.6.3 USGS Sturgeon Population Assessments	35
3.7 Criterion 7: Mink inhabit and reproduce within areas contiguous to the Genesee River and streams within the defined area, OR physical and biological habitat are suitable for mink.....	36
3.7.1 Background.....	36
3.7.2 Mink Studies (2003 – 2007)	36
3.7.3 Follow-up Mink Studies (2013 – 2015).....	38
3.7.4 Overall Assessment of Mink Studies.....	39
3.8 Criteria Conclusions.....	39
4. Additional Activities Supporting BUI Removal.....	40
4.1 2018-2022 Lake Ontario Lakewide Action and Management Plan.....	40
4.2 Water Education Collaborative (WEC)/H2O Hero Campaign.....	40
4.3 Genesee RiverWatch Initiative	41
4.4 Genesee River Nine Element (9E) Plan.....	41
4.5 DEC Habitat Management Plans (HMP).....	42
5. Public Outreach.....	42
6. Conclusions.....	42
6.1 Removal Statement.....	42
6.2 BUI Removal Steps.....	43
6.3 Post-Removal Responsibilities	43
7. References.....	45
Appendices.....	48
Appendix A: List of Remedial Advisory Committee Members & Letter of Support	49
Appendix B: Habitat Restoration Projects.....	50
Appendix C: Braddock Bay Restoration Fact Sheet.....	51
Appendix D: 2013 – 2018 TSS Data	52
Appendix E: 2003-2007 Mink Studies	53
Appendix F: 2013-2015 Mink Studies.....	54

List of Abbreviations

AMP	Adaptive Management Plan
AOC	Area of Concern
BAP	Biological Assessment Profile
BCC	Bioaccumulative Chemical Contaminant
BMP	Best Management Practice
BUI	Beneficial Use Impairment
CEI	Center for Environmental Information
CMU	Central Michigan University
CSMI	Cooperative Science and Monitoring Initiative
DU	Ducks Unlimited
ECCC	Environment and Climate Change Canada
GLCWMP	Great Lakes Coastal Wetland Monitoring Program
GLPF	Great Lakes Protection Fund
GLRI	Great Lakes Restoration Initiative
GLWQA	Great Lakes Water Quality Agreement
HMP	Habitat Management Plan
HSI	Habitat Suitability Index
IBI	Indices of Biotic Integrity
IJC	International Joint Commission
LAMP	Lakewide Action and Management Plan
LC100	Lethal Concentration-100% (the concentration of a substance that results in 100% mortality of a test species)
LOAEL	Lowest Observed Adverse Effect Level
MCDOH	Monroe County Department of Health
MCDPH	Monroe County Department of Public Health
MMP	Marsh Monitoring Program

NOAEL	No Observed Adverse Effect Level
NRCS	Natural Resources Conservation Service
NRD	Natural Resource Damage
NWI	National Wetlands Inventory
NYFO	New York Field Office
NYSDAM	New York State Department of Agriculture and Markets
NYSDEC	New York State Department of Environmental Conservation
PAH	Polycyclic Aromatic Hydrocarbon
PCB	Polychlorinated Biphenyl
RAC	Remedial Advisory Committee
RAP	Remedial Action Plan
RCRA	Resource Conservation and Recovery Act
RIBS	Rotating Integrated Basin Studies
SWCD	Soil and Water Conservation District
SUNY	State University of New York
TEQ	Toxic Equivalency
TRL	Technical Review Lead
TSS	Total Suspended Solids
USACE	United States Army Corps of Engineers
USEPA	United States Environmental Protection Agency
USFWS	United States Fish and Wildlife Service
USGS	United States Geological Survey
USPC	United States Policy Committee
WEC	Water Education Collaborative
WMA	Wildlife Management Area
9E	Nine Element Plan

1. Introduction and Report Purpose

In the Great Lakes Basin, the International Joint Commission (IJC) has identified 43 Areas of Concern (AOCs) under Annex 1 of the Great Lakes Water Quality Agreement (GLWQA) where pollution from past industrial activities and waste disposal practices has caused significant ecological degradation. Up to fourteen beneficial use impairments (BUIs), or indicators of poor water quality, are used to evaluate the condition of an AOC.

The Rochester Embayment AOC encompasses the lower portion of the Genesee River from the mouth up to the Lower Falls in Rochester, NY and the portion of Lake Ontario within a straight line drawn from Bogus Point to Nine Mile Point (**Figure 1**). This area was originally listed as an AOC due to the known or suspected presence of multiple BUIs, including Loss of Fish and Wildlife Habitat, which, is considered impaired “when fish and wildlife management goals have not been met as a result of loss of fish and wildlife habitat due to a perturbation in the physical, chemical, or biological integrity of the Boundary Waters, including wetlands” (IJC, 1991).



Figure 1: Map of the Rochester Embayment Area of Concern

Following an evaluation of data gathered to address this impairment, the New York State Department of Environmental Conservation (NYSDEC) and the Monroe County Department of Public Health (MCDPH) have determined that the Loss of Fish and Wildlife Habitat BUI can be removed in accordance with established AOC program guidance. The local Remedial Advisory Committee (RAC) supports the removal of the BUI (**Appendix A**). The purpose of this BUI

removal report is to present the rationale and supporting data to remove the Loss of Fish and Wildlife Habitat BUI from the Rochester Embayment AOC. In those instances where the criteria have not been fully met, the data presented herein demonstrate that the source of the root problems associated with the BUI lie outside of the AOC, and therefore outside the scope of the Remedial Action Plan (RAP) that was developed to address the causes of the impairment. The removal criteria established to remove the BUI designation have been met to the maximum extent practicable under the RAP.

2. Background and BUI Removal Criteria

Per Annex 1 of the 1987 amendment to the GLWQA, all AOCs were mandated to develop a RAP in three stages. Under this framework, the Stage I RAP identifies specific BUIs and their causes, the Stage II RAP outlines the restoration work needed, and the Stage III RAP documents completion of these restoration activities and recommends the delisting of the AOC. Currently, the RAP for the Rochester Embayment AOC consists of the Rochester Embayment Remedial Action Plan Stage I (NYSDEC/Monroe County Department of Planning and Development, 1993) and Rochester Embayment Remedial Action Plan Stage II [NYSDEC and Monroe County Department of Health (MCDOH), 1997]. Subsequent addenda to the Stage II RAP have also been completed.

The Loss of Fish and Wildlife Habitat BUI was originally listed as impaired in the Stage I and Stage II RAPs due to:

- Loss or degradation of suitable wetland areas for fish and wildlife;
- Degradation of riparian vegetation important for stabilization and habitat;
- Turbid water quality conditions;
- Decreasing macroinvertebrate species indicative of healthy aquatic conditions;
- Lacking amphibian diversity and abundance;
- Loss of juvenile and adult Lake Sturgeon in the Genesee River; and,
- Decreasing mink presence and reproduction in areas of the Genesee River.

2.1 BUI Removal Criteria

The ecological indicators used to assess the Loss of Fish and Wildlife Habitat BUI, and the criteria established to remove the BUI, were selected by the RAC Habitat Oversight Committee. BUI removal criteria were developed to define endpoints for each of the conditions above that led to the BUI designation and were first documented in the Stage II RAP Addendum (NYSDEC/MCDOH, 2002) with subsequent minor edits made and presented in the Rochester Embayment Area of Concern Beneficial Use Impairment Delisting Criteria Report (Ecology and Environment, 2009). The final removal criteria are as follows:

- There is no net loss of acreage and quality of federal or state-designated wetlands, using 1996 as the baseline year for comparisons; and
- There is no net loss of the existing 50-foot-wide buffer strip of trees and shrubs on both sides of NYSDEC classified streams and the Genesee River up to the Lower Falls (using 1999 as the baseline year for comparisons); and

- Suspended sediment concentrations in the Genesee River remain less than 30 mg/L for at least 80% of a year, and exceed 200 mg/L for no more than five events with a combined duration of not greater than 20 days, as determined by a five-year average; and
- *Hexagenia*, or another appropriate indicator, is present in the Embayment and in suitable habitats in the Genesee River up to the Lower Falls; members of the stonefly, mayfly, and caddisfly families are present in streams; and
- Amphibian diversity and abundance in the study area (including the Genesee River up to the Lower Falls if monitoring can be performed safely) are comparable to expected standards for the type of habitat; and
- Lake sturgeon of different life stages inhabit the Genesee River up to the Lower Falls and the Embayment, OR physical and biological habitat are suitable for sturgeon; and
- Mink inhabit and reproduce within areas contiguous to the Genesee River and streams within the defined area, OR physical and biological habitat are suitable for mink.

2.2 BUI Removal Considerations

When evaluating whether to proceed with the removal of the Loss of Fish and Wildlife Habitat BUI, which included the review of technical reports, presentations, and other supporting documents, the following questions were considered:

- Are the methods and results cited in the material technically and scientifically sound?
- Does the information cited regarding restoration of the impaired beneficial use sufficiently address the removal criteria?
- Do the RAC and public concur that the removal criteria have been met?
- In the case where BUI removal criteria have not been explicitly or fully met, are any alternative removal scenarios applicable in accordance with established programs and guidance?

2.3 Strategy and Rationale

The United States Environmental Protection Agency (USEPA) guidance document *Restoring United States Great Lakes Areas of Concern: Delisting Principles and Guidelines*, accepted by the United States Policy Committee (USPC, 2001) states that removal of a beneficial use impairment can occur under any of these scenarios:

- A delisting target has been met through remedial actions which confirms that the beneficial use has been restored.
- It can be demonstrated that the beneficial use impairment is due to natural rather than human causes.
- It can be demonstrated that the impairment is not limited to the local geographic extent but rather is typical of lakewide, region-wide, or area-wide conditions (under this situation, the beneficial use may not have been originally needed to be recognized as impaired).
- The impairment is caused by sources outside the AOC. The impairment is not restored but the impairment classification can be removed or changed to “impaired-not due to local sources.” Responsibility for addressing “out of AOC” sources is given to another party.

A description of how each of the seven components of the removal criteria for the Loss of Fish and Wildlife Habitat BUI have been met in relation to the four scenarios above is provided in **Section 3**.

3. BUI Indicator Status Resolution

As presented in **Table 1** and subsequently described in this section, the available data demonstrate that the removal criteria established for the Loss of Fish and Wildlife Habitat BUI have either been met to the maximum extent practicable under the RAP, are indicative of lakewide/regional conditions not unique to the Rochester Embayment AOC or are due to sources outside of the AOC.

Table 1: Basis of removal for each BUI criterion.

Criterion (<i>abbreviated</i>)	Basis for Removal
1: Loss of wetland acreage and quality	Criterion met to the extent feasible under AOC Program
2: Riparian buffer (vegetation)	Criterion met to the extent feasible under AOC Program
3: Suspended sediment	Natural causes/Sources outside AOC
4: <i>Hexagenia</i> or other indicator	Criterion met
5: Amphibian diversity and abundance	Criterion met
6: Lake Sturgeon	Criterion met
7: Mink presence/reproduction/habitat	Criterion met

3.1 Criterion 1: There is no net loss of acreage and quality of federal or state-designated wetlands, using 1996 as the baseline year for comparisons.

In 2011 Ecology & Environment Inc was commissioned by USEPA with support from the RAC to create an Interim Rochester Embayment AOC Strategic Plan for BUI delisting (Interim Plan, E&E 2011). This document became the guidance used to assess the current condition of each criterion and identified data gaps along with suggested methods to fill them. With regards to the first criterion of “no net loss in quality or quantity of wetlands, using 1996 as a baseline,” the E&E 2011 Interim Plan recognized that if there was a loss of wetland acreage in the AOC since the 1996 baseline, it would be difficult to add wetland back considering most of the loss was due to development. Therefore, a recommendation was made in the 2011 Interim Plan to request assistance from the U.S. Fish and Wildlife Service (USFWS) New York Field Office (NYFO) in identifying wetlands that could be enhanced in quality. NYFO’s assessments were focused on aerial imagery of emergent wetlands captured during flights conducted in October 1951 (USFWS,

2014). NYFO was not able to obtain reports or data from a wetland survey purportedly conducted in 1996, as referenced in the BUI removal criterion. Considering this, the RAC was convened in January 2013 and agreed to use the year 1951 as a replacement for the 1996 reference year for assessment purposes. NYFO used the guidance in the E&E 2011 Interim Plan to determine a project area around the AOC boundary (USEPA 2014), and wetlands within that boundary would be assessed.

NYFO's assessments determined there has been an overall net loss of wetland acreage within the project area and in coastal Lake Ontario emergent wetlands since 1951 (USFWS 2014). The greatest net losses in acreage were in Buck Pond (-121 acres), Braddock Bay (-67 acres), and the Genesee River (-46 acres). The greatest losses as a percent of 1951 acreage were seen in Slater Creek (-43%), Genesee River (-35%), Long Pond (-27%), and Braddock Bay (-23%). There were apparent gains in emergent wetland in Salmon Creek (+40%), West Creek (+44%), and Irondequoit Bay tributaries (+7%), but these gains were in dense invasive cattail mass. Losses of wetland acreage were due principally to fill and hydrological changes resulting from transportation and residential development, erosion, stream channelization, lake water level management, and shifts in vegetation types at the water-emergent vegetation boundary.

In 2014, recognizing that there were limitations on the extent to which wetland habitat within and adjacent to the AOC could be restored, a priority list of 10 carefully selected habitat projects were developed by USFWS NYFO in collaboration with the RAC, NYSDEC, United States Army Corps of Engineers (USACE), and other partners. This was done in lieu of setting a numeric acreage goal for restoration. The projects were identified based on habitat assessments completed by USFWS and USACE, and targeted areas that represented the most degraded habitat areas within and adjacent to the AOC in terms of losses in quality and quantity of emergent wetlands, including some of the areas listed above. Sites were prioritized and vetted through the RAC and NYSDEC, and it was determined that completion of these projects would satisfy the BUI removal criterion. The list of projects was revised in 2016, with two projects in the lower Genesee River being removed due to uncertain implementation dates resulting from sediment investigations that were ongoing in the river related to the Kodak/Eastman Business Park site, under NYSDEC's Resource Conservation and Recovery Act (RCRA) program. Despite being removed from the list, USFWS subsequently implemented the two projects in 2021 after remedial work in the affected area of the river had been completed, using funding from the Natural Resource Damage (NRD) settlement with Kodak. These two RCRA habitat projects are described below in Section 3.1.3 "Other Supporting Habitat Restoration Projects."

The final list of eight projects that would be implemented to address the BUI removal criterion are listed below. Documentation related to the development and finalization of the habitat project list is provided in **Appendix B**.

1. USACE – Braddock Bay (Alternative 7c)
2. USFWS – Braddock Bay
3. USFWS – Salmon Creek
4. USFWS – Confluence of West and Salmon Creek
5. USFWS – Long Pond West
6. USFWS – Buck Pond East

7. Ducks Unlimited – Buck Pond Phase I
8. Ducks Unlimited – Buck Pond Phase II

The project locations (**Figure 2**) are contained inside the Braddock Bay Wildlife Management Area (WMA), a New York State owned property managed by NYSDEC for wildlife conservation and wildlife-associated recreation (hunting, trapping, wildlife viewing, and photography). The Braddock Bay WMA is a shallow water bay-marsh complex existing in five units along the Lake Ontario shoreline, ranging from two to six miles west of Rochester.



Figure 2: Habitat restoration project areas

All of these areas except Braddock Bay itself, which is directly connected to the lake, are connected to the lake by intermittent channels. The channels plug and reopen as lake currents and wave action change the character of the gravel and sand barrier bars. Due to altered adjacent shorelines and changes in sediment dynamics in Lake Ontario, some of these openings have become permanently clogged and inaccessible to some wildlife or unusable as suitable spawning/nesting habitat.

This bay-marsh area was selected as significant AOC habitat because it provides excellent waterfowl nesting, resting, and feeding habitats, along with protective and spawning habitat for fish species, including Northern pike. Puddle ducks, particularly mallards, blue-winged teal, and

wood ducks are common nesters. During the spring and fall migrations, all waterfowl common to the Atlantic flyway have been observed utilizing the area including scarce species such as brant, Barrow's goldeneye, and the Harlequin duck. The primary wildlife objective for this WMA is to provide a diversity of habitats to benefit the full range of migratory bird and fish species that depend on this area. This is an important migration stop-over area for many species of songbirds and raptors, as well as waterfowl. Grasslands are managed to provide nesting and wintering areas for several species of songbirds and raptors as well as habitat for grassland gamebirds and small mammals.

The eight projects implemented in the WMA, described further below, were designed to restore function and biodiversity and improve the overall quality of these areas in terms of fish and wildlife habitat.

3.1.2 Project Descriptions

Braddock Bay (Includes Projects 1 & 2 Above)

The namesake of the Braddock Bay WMA, Braddock Bay was historically a barrier wetland, but nearby development and alterations in the sediment flow of Lake Ontario had resulted in the loss of the protective barrier. Over time the emergent wetland has been shrinking and becoming a monoculture of narrow leaf cattail (*Typha angustifolia*) (Wilcox et. al, 2008). A series of studies and design evaluations were performed by the USACE to determine possible solutions for enhancing and protecting this wetland. These options were presented to the RAC, and design Alternative 7c was selected (USACE 2014). USACE in collaboration with USFWS worked to excavate almost 7 acres of potholes and 12,000 feet of channels to break up the dense cattail (Figure 3), with the goal of improving connectivity and biodiversity of wetlands. A 3-acre emergent wetland was also added to a section of the open water using sand from on-site, along with a rock barrier for wetland protection (Figure 4). Additional information of the Braddock Bay restoration project, which was completed in 2016-2017, can be found in **Appendix C** and at this [website](#).

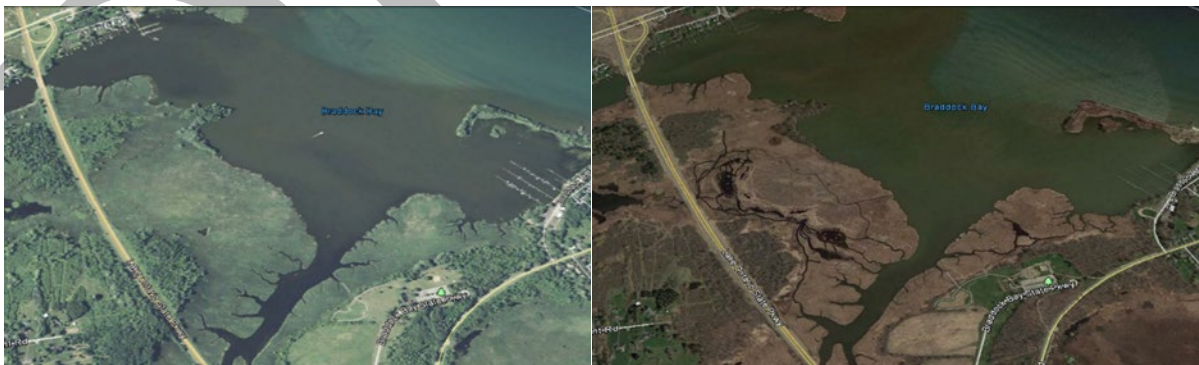


Figure 3: Braddock Bay before and after



Figure 4: Braddock Bay barrier beach

Salmon Creek (Project 3 Above)

Twenty acres of emergent marsh and sedge meadow habitat were restored along the edge of Salmon Creek, a tributary to Braddock Bay. And, 1,800 feet of channels were created/enhanced, providing hydrological connectivity to the restored marsh and Salmon Creek (**Figure 5**). This work was completed in 2016. Water is managed through the construction of a low berm and water control structure with fish passage. The system is managed by NYSDEC to provide sedge meadow habitat for spawning fish in the spring, and forging resources for migrating waterbirds. Ducks Unlimited (DU) developed a long-term management plan to control the water level in the newly restored marsh in cooperation with NYSDEC to maximize the value of the system for fish and wildlife. The restoration of the meadow and emergent marsh will also assist with reducing non-point source pollutants entering Salmon Creek.

Confluence of West and Salmon Creeks (Project 4 Above)

This is an area located within the Braddock Bay WMA and includes adjacent wetlands along Salmon Creek, as well as vernal pools (**Figure 5**). Almost six acres of potholes and 2500 feet of channel were opened to enhance connectivity, and six acres of habitat mounds, elevated areas designed to mimic transitional sedge meadow habitat, were planted with native plants to increase diversity, along with cattail control. This was completed in 2016.



Figure 5: Salmon Creek and Confluence of West and Salmon Creek before and after

Long Pond West (Project 5 Above)

This area is one of five wetland bays that are part of the Braddock Bay WMA. This project, completed in 2016, opened 5.7 acres of potholes, 1,000 linear feet of channels, and created 6 acres of wet meadow habitat mounds thus increasing biodiversity and connectivity (**Figure 6**).

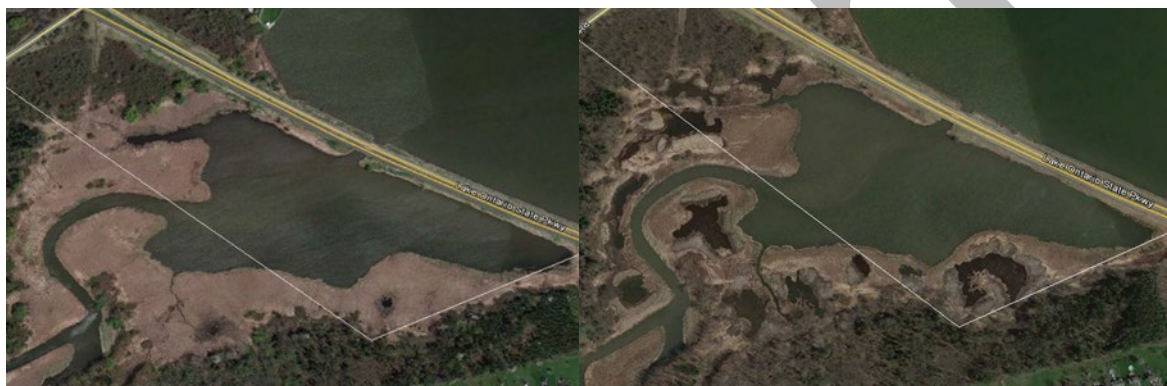


Figure 6: Long Pond West before and after

Buck Pond (Includes Projects 6, 7, & 8 Above)

Another of the five wetland bays that are part of the Braddock Bay WMA, Buck Pond is approximately 700 acres and is directly connected to the Rochester Embayment AOC. As part of Buck Pond Phase I, completed in 2014, 200 acres of coastal marsh was enhanced by DU, with the addition of 6,000 linear feet of channel and 2 acres of potholes in the southern section of the marsh. Buck Pond Phase II, completed in 2015, included an additional 4,000 linear feet of channel, 3 acres of potholes, and 10 acres of sedge meadow restoration on the western side of the pond. Additional work was completed by USFWS in 2016 in an area east of the DU work. This project opened 3.9 acres of potholes, 3,000 linear feet of channels, and created 4 acres of wet meadow habitat mounds (**Figure 7**).



Figure 7: Buck Pond before and after

As previously described, the implementation of the eight projects above restored habitat to the extent feasible and were deemed sufficient to meet the intent of the BUI removal criterion, despite not meeting the specific metrics of the criterion relating to “no net loss.” As such, this BUI removal criterion has been met to the degree possible under the AOC program and taking into consideration current and anticipated future land use practices within the AOC.

3.1.3 Other Supporting Habitat Restoration Projects:

While the RAC determined that the 8 projects above were all that were necessary to meet the BUI removal criterion, additional completed and ongoing habitat restoration efforts will also contribute to the overall improvement of habitat within and adjacent to the AOC. Some of these are described below.

Buttonwood Creek

Located within the Braddock Bay WMA, Buttonwood Creek is a tributary to Braddock Bay and is less than 0.1 miles from the Rochester Embayment AOC. This shallow creek had become clogged with invasive cattail, reducing connectivity to Braddock Bay and Lake Ontario. Approximately 6,000 feet of meandering channel were excavated to establish connectivity between the main stem of Buttonwood Creek, restored sedge meadow, and spawning pool. Four acres of spawning pools and approximately 10 acres of sedge meadow were restored and connected by channels within the cattail mat (**Figure 8**). DU worked with NYSDEC to develop a management plan to ensure the resource is managed to remain a valuable and diverse habitat.



Figure 8: Buttonwood Creek area before and after

Resource Conservation and Recovery Act

Some recommended restoration projects that were included on the original list of 10 projects to be implemented were constructed after remedial sediment work in the lower Genesee River was completed. Wetland areas in the remediation footprint (Wetland C North and Wetland C South) consisted of a monoculture of cattails that left little room for vegetation biodiversity. The cattail monoculture and contaminated sediments were removed and replaced with clean sediments and plantings of 38 different native species including emergent, floating aquatic and submerged aquatic vegetation. The plantings included 12,000 individual wetland plugs of 20 different native species and a seed mix with an additional 18 native species. In addition, localized areas of adjacent wetlands were lowered to create varying water depths that were planted with a mix of emergent

and submergent habitat types to encourage fish and amphibian spawning and reptile breeding. This work was completed in 2021 in the lower section of the Genesee River, in the vicinity of the turning basin, within and adjacent to Wetland C (**Figure 9**, also refer this [fact sheet](#)).



Figure 9: Completed Wetland C North site

3.1.4 Monitoring of Restoration Projects

As mentioned above, management plans have been developed for some of the habitat restoration work that has been completed, and some areas will be managed as part of NYSDEC's ongoing activities for the Braddock Bay WMA. Additionally, in 2015, NYSDEC and the State University of New York (SUNY) at Brockport partnered to design a monitoring and adaptive management plan (AMP), in collaboration with USACE, USFWS, USEPA, and the Town of Greece. The purpose of this monitoring effort was to collect data that can be used to assess if the Braddock Bay barrier beach and pothole/channel restoration project has been successful in achieving its objectives and to support local resource agencies and stakeholders in adaptive management. The Braddock Bay barrier wall was designed to be a source and sink for lake sediments, with the goal of design being that it would maintain itself as a sandy beach, instead of just a protective rock wall. This is a novel method of offshore wetland protection, and the monitoring plan was designed to evaluate efficacy and allow the project to evolve if restoration objectives were not being met. The data collected will be compared against baseline performance criteria identified in the AMP including:

- Emergent and submerged vegetation surveys
- Fish, waterbird, and amphibian surveys,
- Wetland erosion studies
- Bathymetric surveys
- Water quality
- Sediment transfer
- Barrier beach structural monitoring

The report data (Monitoring and Adaptive Management Reports 2016-2021, USACE/Wilcox/Schultz) have shown a positive response to the restoration activities. Target species such as northern pike have been observed in restoration areas, along with migratory birds

that have not been counted in the area for several years prior to the restoration activities. The final report is expected by September 2023 and will contain suggestions for future management and monitoring in the project area.

3.2 Criterion 2: There is no net loss of the existing 50-foot-wide buffer strip of trees and shrubs on both sides of NYSDEC classified streams and the Genesee River up to the Lower Falls (using 1999 as the baseline year for comparisons).

When the RAC first selected this as a removal criterion the AOC was defined as “approximately 35 sq miles between Nine Mile Point in the town of Webster and Bogus Point in the Town of Parma, approximately a six-mile reach of the lower Genesee River from the lower falls to the river mouth, and the entire Rochester Embayment watershed.” After the development of the Stage I and II RAPs, the AOC footprint was reduced to include only the embayment and the lower six miles of the Genesee River as depicted by the AOC boundary shown on **Figure 1**. This reduced area excludes the NYSDEC classified streams that drain into the Rochester Embayment.

Recognizing that it is unlikely that developed areas could be restored to habitat (NYSDEC December 2011) and given that the AOC straddles several townships with different levels of stream and floodplain protection ordinances, the RAC determined that the NYS Environmental Conservation Law Article 15, Title 5, Section 15-0501 (Protection of Water) would provide sufficient protections from future shoreline development along the AOC portion of the river and classified streams. Article 15 states: *A Protection Of Waters Permit is required for disturbing the bed or banks of a stream with a classification of AA, A or B, or with a classification of C with a standard of (T) or (TS) (disturbance may be either temporary or permanent in nature). "Banks" means that land area immediately adjacent to and which slopes toward the bed of a watercourse and which is necessary to maintain the integrity of the watercourse. A bank will not be considered to extend more than 50 feet horizontally from the mean high-water line.* Additional information can be found [here](#).

The RAC also determined that the habitat projects used to address BUI removal Criterion 1 (described in **Section 3.1**) would also help satisfy the portion of Criterion 2 pertaining to the classified streams, since portions of West, Salmon, and Buttonwood Creeks were all included in the habitat restoration projects. Additionally, the riparian area of the lower Genesee River (within 100' of Lake Ontario and river gorge, except in manufacturing/industrial zones) is classified by the City of Rochester as a [Critical Environmental Area](#). This designation is taken into consideration during the permitting process of activities within the designated area.

Together, the Critical Environmental Area designation of the lower Genesee River and the NYS protection of water program provide protections within a 50-foot-wide buffer strip along classified streams and the lower Genesee River, satisfying the BUI removal criterion to the extent possible.

3.3 Criterion 3: Suspended sediment concentrations in the Genesee River remain less than 30 mg/L for at least 80% of a year, and exceed 200 mg/L for no more than five events with a combined duration of not greater than 20 days, as determined by a five-year average.

3.3.1 Background

Widespread deforestation, conversion of land for agricultural uses, construction of dams, stormwater management practices, and other land use modifications within the Genesee River watershed have significantly altered the flow of sediment naturally introduced through the system and greatly increased the capacity of exposed soil and glacial debris to be transported to nearby waterways. Various agencies and organizations have collaborated over the years to assess changing water quality conditions within the Genesee River and identify specific areas in need of best management practices within the watershed to reduce erosion and the amount of sediment making its way into the river.

3.3.2 Water Quality Monitoring Program

In 2013, NYSDEC, in partnership with United State Geological Survey (USGS) and USEPA, initiated a five-year water quality monitoring program on the Genesee River and two tributaries to the river—Oatka Creek and Honeoye Creek. Funded by the federal Great Lakes Restoration Initiative (GLRI), this program was intended to address two important data needs: first, it would provide information on the extent to which the implementation of riparian/stormwater best management practices (BMPs) in the watershed were resulting in improved water quality conditions in the river and tributaries. This component is not directly related to the BUI removal criterion and is not discussed further in this report. The second component of the monitoring program was to provide a water quality data set that would be sufficient for assessing suspended sediment conditions within the AOC portion of the Genesee River relative to upstream areas, and the general degree to which the removal criterion had been met. It was acknowledged that the data may not provide all information necessary to fully assess each individual component of the removal criterion; specifically, the portion relating to “...for no more than five events with a combined duration of not greater than 20 days, as determined by a five-year average...” as this would have required daily monitoring for the full five-year period, which was not feasible.

NYSDEC staff collected samples from four locations along the Genesee River on an approximately monthly basis from May 2013 through April 2018. The samples were analyzed for a variety of water quality parameters, including total suspended solids (TSS). Additional samples were collected during high-flow periods, typically following heavy rain or snow melt events. The locations were selected to assess water quality in sections of the river with different physical characteristics and land use patterns (**Figure 10**).

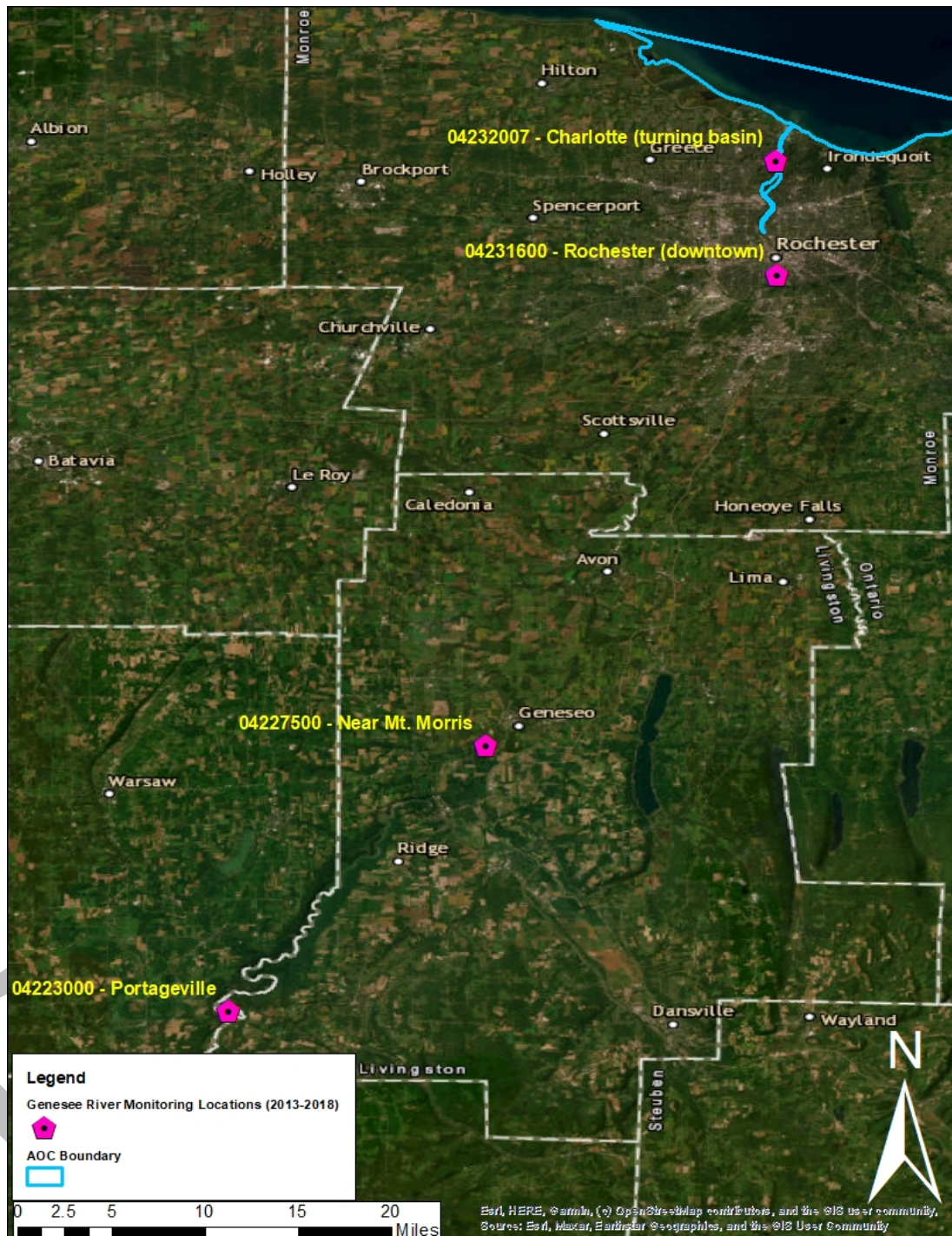


Figure 10: 2013–2018 TSS monitoring program locations

The monitoring location at the turning basin in the lower section of the river at Charlotte is within the AOC boundary. The next closest monitoring location was approximately seven river miles upstream within the downtown area of Rochester, outside the AOC boundary. Both locations are in predominantly urban/suburban sections of the river that are relatively deep, wide, and slow moving, with both undeveloped and heavily developed/hardened shoreline segments. The

remaining two locations, near Mt. Morris and at Portageville, are in shallower, narrower, and faster moving sections of the river, in predominately rural/agricultural areas.

The goal of the monitoring program was to collect monthly water quality samples for a full five years, with at least three additional event-based (high-flow) collections each year. For logistical and other reasons largely related to regional weather conditions (e.g., ice cover, highly localized rainfall, etc.), neither the monthly nor high-flow targets were specifically met each year at any of the monitoring locations. However, a very robust data set over a five-year period that included both base flow and high-flow conditions was obtained, and this serves as the basis for assessing the water quality component of the BUI removal criteria, as discussed below.

A summary of the TSS data from the monitoring program, as relevant to the BUI removal criterion, is reported in **Table 2**. The full data set is provided in **Appendix D**. The data in **Table 2** are presented for each 12-month (one-year) period, beginning with the onset of the program in May 2013. Cumulative/total results over the entire five-year period are also presented.

2013 – 2018 Genesee River TSS Monitoring					
		Charlotte (inside AOC)	Rochester (outside AOC)	Mt. Morris (outside AOC)	Portageville (outside AOC)
Year 1 (May 2013 – April 2014)	Number of Samples	12	12	13	14
	Min TSS (mg/L)	11	8.1	8.8	4.3
	Max TSS (mg/L)	482	618	1480	1450
	Average TSS (mg/L)	87	92	169	133
	Number samples < 30 mg/L	8	5	7	11
	% of Samples < 30mg/L	67%	42%	54%	79%
	Number of Samples > 200 mg/L	1	1	2	1
Year 2 (May 2014 – April 2015)	Number of Samples	12	10	12	12
	Min TSS (mg/L)	3	7.7	2.6	1.1
	Max TSS (mg/L)	262	772	2200	869
	Average TSS (mg/L)	74	168	425	183
	Number samples < 30 mg/L	8	5	6	7
	% of Samples < 30mg/L	67%	50%	50%	58%
	Number of Samples > 200 mg/L	2	3	5	3
Year 3 (May 2015 – April 2016)	Number of Samples	13	13	13	13
	Min TSS (mg/L)	7.3	8	13.4	3.7
	Max TSS (mg/L)	74.6	297	477	391
	Average TSS (mg/L)	27	48	106	63
	Number samples < 30 mg/L	8	9	3	11
	% of Samples < 30mg/L	62%	69%	23%	85%
	Number of Samples > 200 mg/L	0	1	2	2
Year 4 (May 2016 – April 2017)	Number of Samples	12	13	13	13
	Min TSS (mg/L)	2.4	10	13	3
	Max TSS (mg/L)	172	416	2570	1320
	Average TSS (mg/L)	35	55	258	180
	Number samples < 30 mg/L	10	8	8	9
	% of Samples < 30mg/L	83%	62%	62%	69%
	Number of Samples > 200 mg/L	0	1	2	2
Year 5 (May 2017 – April 2018)	Number of Samples	11	11	12	12
	Min TSS (mg/L)	8.5	10.2	24.8	3.8
	Max TSS (mg/L)	211	226	1010	3880
	Average TSS (mg/L)	57	74	194	382
	Number samples < 30 mg/L	6	5	3	7
	% of Samples < 30mg/L	55%	45%	25%	58%
	Number of Samples > 200 mg/L	1	1	4	2
Five-Year Totals	Number of Samples	60	59	63	64
	Min TSS (mg/L)	2.4	7.7	2.6	1.1
	Max TSS (mg/L)	482	772	2570	3880
	Average TSS (mg/L)	56	84	228	184
	Number samples < 30 mg/L	40	32	27	45
	% of Samples < 30mg/L	67%	54%	43%	70%
	Number of Samples > 200 mg/L	4	7	15	10

Table 2: Summary of Total Suspended Solids (TSS) in the Genesee River, 2013–2018 Monitoring Program

Graphical representations of the data are provided as **Figures 11a** and **11b**; these were separated into two figures to allow a better comparison of results between the monitoring locations within the upper and lower sections of the river. The lower river locations are the most relevant to the BUI evaluation.

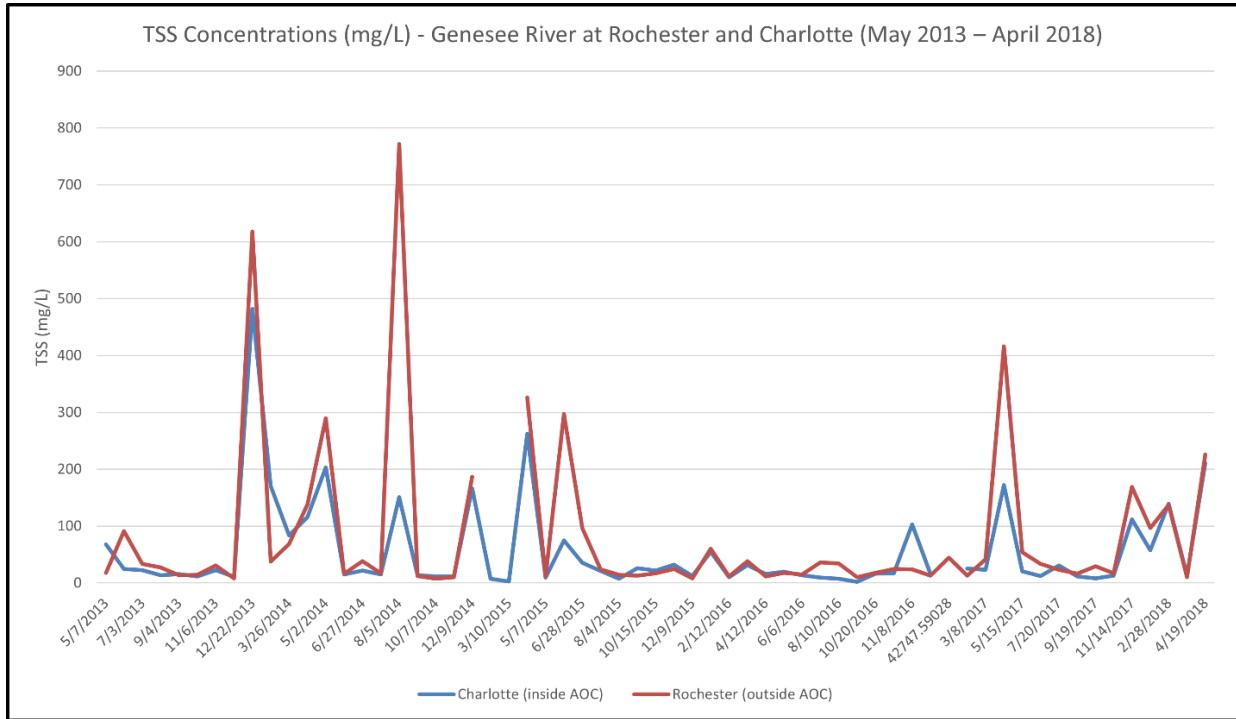


Figure 11a: Time series of total suspended solids (TSS) results from the lower Genesee River, 2013–2018

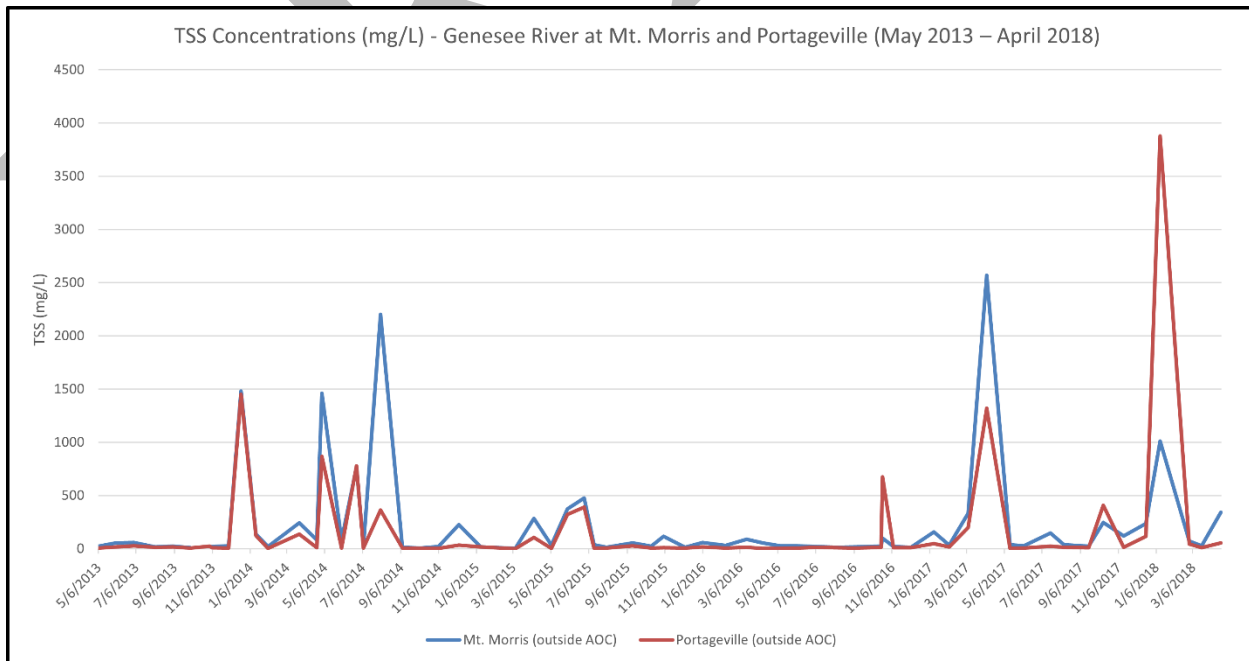


Figure 11b: Time series of total suspended solids (TSS) results from the upper Genesee River, 2013–2018

Overall, the results of the five-year monitoring program did not meet the first component of the BUI removal criterion, “Suspended sediment concentrations in the Genesee River remain less than 30 mg/L for at least 80% of a year.” Within the AOC, the results at Charlotte met this target in only one of five years. The other three locations are outside of the AOC and are not subject to a direct evaluation against the removal criteria, but the results were similar to those at Charlotte, with only one of the three locations (Portageville, the furthest upstream) exhibiting TSS concentrations less than 30 mg/L for 80% of a year, and this also only occurred in one of five years. Of particular importance within the data set are the results at the Rochester monitoring location relative to those at Charlotte. These are the only two locations within the deep, wide, and relatively slow-moving section of the river in an urban/suburban setting. The location at Rochester is the best indicator of TSS concentrations that are coming into the AOC from upstream sources. Over the five-year monitoring period, TSS concentrations at Rochester were less than 30 mg/L 54% of the time, compared to 67% at Charlotte. While not meeting the target of 80% each year, TSS concentrations within the AOC were less than 30 mg/L more often than at the nearest location upstream of the AOC.

The second component of the BUI removal criterion, “Suspended sediment concentrations in the Genesee River exceed 200 mg/L...for no more than five events with a combined duration of not greater than 20 days, as determined by a five-year average,” is more difficult to directly assess. Part of this criterion (i.e., “five events with a combined duration of not greater than 20 days”) was intended to reflect annual conditions, averaged over a five-year period. However, as previously stated, obtaining the data set to allow for this would generally require continuous (daily) monitoring of TSS for five years, which was not feasible. When the five-year monitoring program was planned and implemented this was understood, but it was determined that the resulting data set would be sufficient for assessing TSS conditions within the AOC relative to upstream areas and determining whether sources within the AOC were resulting in the elevated TSS concentrations causing the BUI.

Figure 11a shows that TSS concentrations at Charlotte and Rochester exhibited similar concentration patterns over the five-year monitoring period, with the Rochester location exhibiting somewhat higher concentrations during the high-flow events (represented by higher peaks on the figure). As shown on **Table 2**, only 4 of the 60 samples collected at the AOC monitoring location at Charlotte over the five-year period exhibited TSS concentrations greater than 200 mg/L. At the Rochester location, upstream of the AOC, 7 of 59 samples were greater than 200 mg/L. Additionally, the upstream location at Rochester exhibited maximum and average TSS concentrations greater than those at Charlotte in all five years of the monitoring program. The maximum and average concentrations at Charlotte were also less than those at the upper watershed locations (Mt. Morris and Portageville) in all five years.

Taken altogether, the results of the study as presented above are a clear indication of sediment loading within the Genesee River watershed, mostly originating outside of the Rochester Embayment AOC. The data do not suggest that there are significant sources of sediment loading within the AOC portion of the lower Genesee River. If significant AOC sources did exist, elevated TSS concentrations would have been expected to be a more frequent occurrence at Charlotte, relative to the Rochester location.

A number of factors can contribute to high sediment load from sources upstream of the AOC that result in persistent water clarity issues. These include agricultural land use practices; nonpoint sediment sources, such as highly erodible glacial till streambanks; and runoff during significant precipitation events. In addition to anthropogenic and climate-driven forces, the Genesee Valley's unique origins and geology are also important factors to consider. As previously mentioned, the upper watershed is predominantly rural/agricultural, and highly erodible banks exist in much of this area. Relative to the downstream section of the river, elevation changes in the upper watershed are more significant, and higher river velocity further exacerbates bank erosion, especially during high-flow events. In the lower section of the river, elevation change is less dramatic and land use gradually transitions to an urbanized setting with increased impervious surfaces and development. Not surprisingly, TSS concentrations in the lower part of the river at Rochester and Charlotte were consistently lower than those at Mt. Morris and Portageville.

3.3.3 Other Efforts Supporting Water Quality Improvements

Sediments within the federal navigation channel portion of the Genesee River are routinely dredged by the USACE. Because of the potential for water quality impacts during dredging activities, including elevated TSS concentrations, the dredging permits issued by NYSDEC include a stipulation that, under no circumstances, are the dredging operations to be conducted in such a manner that water and/or suspended sediments, be allowed to be discharged from the vessel by “overflow dredging”—the process of allowing excess water that accumulates within the dredge barge to overflow as it is filled.

In September 2015, NYSDEC approved the Genesee River Basin Nine Key Element Watershed Plan for Phosphorus and Sediment, with the goal of reducing nutrient and sediment loading to the Genesee River. Nine Element (9E) Plans are locally developed watershed-scale management plans designed to address known water quality issues. The 9E Plan identifies major sub-basins within the Genesee River watershed and prioritizes nutrient and sediment reduction efforts within these sub-basins. Based on data collected through a suite of scientific studies, load estimates from point and nonpoint sources are listed, identifying land use and human activities which contribute the greatest negative impacts. From here, the 9E Plan has identified specific management measures and associated load reduction estimates favored to improve water quality throughout the watershed when implemented. Management measures such as grassed waterways, stream bank stabilization, cover crops, buffer strips, and other green infrastructure projects are effective practices identified for reducing phosphorus and sediment transport within and specific to the Genesee River watershed. Additionally, the Genesee River watershed has been designated as an Agricultural Priority Watershed under the federal GLRI Action Plan III. The development of the 9E Plan, along with the federal priority designation, serve to provide greater access to

State and Federal Great Lakes funding opportunities that will continue to support projects to reduce watershed sediment and nutrient loads.

3.4 Criterion 4: *Hexagenia*, or another appropriate indicator, is present in the Embayment and in suitable habitats in the Genesee River up to the Lower Falls; members of the stonefly, mayfly, and caddisfly families are present in streams.

3.4.1 Background

This criterion was developed by the RAC as an endpoint addressing a specific condition that led to the BUI designation, “Invertebrate habitat is impaired as indicated by absence of indicator species” (MCDOH/NYSDEC, 2011). However, as was the case for Criterion 2 described above in Section 3.2, Criterion 4 was developed prior to the original AOC footprint being reduced to include only the embayment and the lower six miles of the Genesee River, as depicted on **Figure 1**. Additionally, “another appropriate indicator” in the first part of the criterion was never well-defined. These two factors presented a challenge when assessing whether the BUI removal criteria had been met. While data from multiple studies are presented, no single data set specifically meets the removal criterion as written, even when the additional two factors noted above are taken into consideration. Collectively, however, the various data sets provide sufficient evidence that conditions within the current AOC boundary do meet the intent of the removal criterion, when assessed relative to the various BUI removal scenarios described in Section 2.3. To provide some additional context for the data discussion in Sections 3.4.2 through 3.4.4, a brief background on some of the assessment methodology is presented below.

Benthic macroinvertebrates serve as indicator species of stream water quality and biotic health and are studied to assess aquatic habitat due to their relatively long lives, diversity, widespread occurrence, and sampling ease. EPT richness is a common index used to evaluate benthic macroinvertebrate communities, measuring the number of different species represented in an ecological community out of three orders of macroinvertebrate species: *Ephemeroptera* (mayfly), *Plecoptera* (stonefly), and *Trichoptera* (caddisfly). The EPT richness is considered a good measure of the overall health of freshwater streams because these three orders are sensitive to several aquatic stressors. *Hexagenia*, order *Ephemeroptera*, is one of the original IJC environmental condition indicators (Shear et al 2003) and has been used extensively in water quality monitoring programs throughout the Great Lakes (Environment Canada, U.S EPA 2009. New York State DEC uses EPT richness as part of a multimetric index of biological integrity called the Biological Assessment Profile (BAP) to assess macroinvertebrate community health. BAP scores are calculated using numerous individual metrics, including: EPT richness, species richness, nutrient biotic index, species diversity, Hilsenoff’s biotic index, species diversity percent model affinity and non-Chironomidae and Oligochaeta richness. A composite BAP score is calculated from the various individual metrics, and ranges from 0 to 10 with 5 representing the biological threshold. A score below 5 indicates moderately to severely impacted conditions and suggests biological impairment while a score of 5 or above is indicative of slightly to non-impaired conditions (Smith et al., 2012).

Although not explicitly stated in the removal criterion, EPT richness effectively fulfills the intent of the “another appropriate indicator” portion of the criterion since, as described above, EPT are

sensitive to aquatic stressors. BAP scores, which incorporate EPT richness long with other metrics of benthic community health, provide an additional means of assessing the “another appropriate indicator” component.

3.4.2 1999 – 2002 USGS Assessment

Between 1999 and 2002 the USGS led a pre-stocking assessment of the Genesee River to evaluate the quality and availability of habitat for all life stages of Lake Sturgeon. As part of this study the researchers conducted a habitat evaluation and assessment of the benthic community as a potential food source. Results showed that the benthic invertebrate community included members of the Chironomidae, Oligochaeta, Megaloptera, Mollusca Families and members of the orders of *Trichoptera*, and *Ephemeroptera*. The highest densities of these organisms occurred within the first 2.5 river miles. Further upstream and still in the AOC between river miles 5.1 and 5.6 the burrowing mayfly (*Hexagenia* sp.) was found in low numbers (Dittman 2006). These results indicate that *Hexagenia* and other indicator species were present in suitable habitats in the Genesee River up to the Lower Falls. It is recognized that this assessment occurred over 20 years ago and did not include the embayment portion of the AOC and, as such, it does not provide sufficient current information regarding the presence of *Hexagenia* throughout the AOC. However, it is important to note the study findings as part of the overall discussion of the BUI removal criterion.

3.4.3 2013 NYSDEC/USGS Assessment

In 2013, for the purpose of evaluating the status of the benthic macroinvertebrate community within the Rochester Embayment AOC, sediment samples were collected by NYSDEC/USGS from 17 sites located inside and outside the AOC, with the outside sites serving as reference locations. Seven locations were on the Genesee River (4 AOC, 3 reference), eight were on Lake Ontario (4 AOC, 4 reference), and two were within Braddock Bay (1 AOC, 1 reference). The locations within the Genesee River portion of the AOC were generally selected to represent worst-case conditions, where previous sediment chemistry results indicated elevated levels of metals contamination. Non-AOC reference sites were located within the Braddock Bay WMA, in the Genesee River upstream of the AOC boundary and along the Lake Ontario nearshore area adjacent to the AOC. Benthic community assessments and sediment toxicity analyses were conducted on the sediment samples.

A comprehensive summary of this assessment and a map showing the monitored locations can be found in the [Rochester Embayment Area of Concern Remedial Action Plan Beneficial Use Impairment Removal Report - Degradation of Benthos](#) (NYSDEC 2017). Even with the selection of river locations that were intended to represent worst-case conditions, the biological assessment of macroinvertebrate communities resulted in BAP values within the AOC areas that were either of similar or better condition than the reference areas, and the evaluation of differences in the composition and abundance of benthic macroinvertebrates indicated similar community structure between AOC and reference groups (Duffy et al 2017). Additionally, the toxicity of sediments (*Chironomus dilutes* growth and survival) from sites sampled in 2013 in the Rochester Embayment AOC were found to be no different than those in sediments from comparable reference sites across the region (Duffy et al 2017).

It should be noted that no EPT taxa (including *Hexagenia*) were found in any sample from the AOC or non-AOC sites during this assessment. For the embayment (Lake Ontario) portion of the AOC, the absence of *Hexagenia* is consistent with earlier findings that suitable habitat in the embayment has not been reported in the past and there is no historical evidence that *Hexagenia* was ever present in large numbers in the embayment (Ecology and Environment June 2011). Further evidence supporting that *Hexagenia* is nearly non-existent in the Lake Ontario basin is the fact that in the USGS long term benthic monitoring program, which has been sampling the benthic community annually since 1992 through 2019 (except for the years of 1996 through 1998) at 45 sites at varying depths around Lake Ontario, only a total of five mayfly nymphs (species not identified) have been captured (Weidel personal communication October 14, 2022, February 7, 2023).

3.4.4. NYSDEC Rotating Integrated Basin Studies (RIBS) Monitoring

NYSDEC monitors water quality in watersheds throughout New York State on a rotating basis through the Rotating Integrated Basin Studies (RIBS) program. A key component of these studies is the assessment of the macroinvertebrate communities. Monitoring locations include two sites along the lower Genesee River within the AOC, and several other sites on smaller streams outside the AOC boundary that feed into the embayment (lake) portion of the AOC (**Figure 12, Tables 4 and 5**). Monitoring the benthic community at these sites is conducted through the collection of macroinvertebrates using one of two different methods, dependent on water depth. The first method is a kick sample, generally used in wadable streams. It involves disturbing the bottom sediments by foot and catching the dislodged organisms in an aquatic net. The second method is multiplate sampling and uses artificial substrate samplers suspended in the water column over a defined period before retrieval and analysis. For both methods, the organisms collected are assessed via the BAP methodology previously described. The most recent BAP scores for the monitored locations range from severely impacted to non-impacted, as reported on **Table 4**. It should be noted that not all locations are monitored on the same rotational cycle.



Figure 12: Rotating Integrated Basin Studies (RIBS) locations

Six of the nine creeks that feed into the embayment portion of the AOC had BAP scores that placed them in the moderately impacted category, two scored as slightly impacted and one scored as severely impacted (**Table 4**). The unnamed tributary to the Genesee River also scored as severely impacted. Importantly, the two sites in the Genesee River within the AOC scored as non and slightly impacted and, while *Hexagenia* have not been detected at these locations, several members of the mayfly and caddisfly families have been found (**Table 5**). Several members of the stonefly, mayfly, and caddisfly families were also found in the streams tributary to the Rochester Embayment (**Table 5**). Statewide data from the RIBS program, including data reported in **Tables 4 and 5**, can be found [here](#) (this website includes a searchable map that shows each of the locations below).

RIBS Site & Description	Year	Method	Number of EPT Species (Richness)	BAP Score	Impact Category
Salmon creek 03-SAMC-5.8 5m below RT 259 bridge	2004	Kick	9	6.01	Slight
	2015	Kick	6	5.71	Slight
Buttonwood Creek 03-BUTN-7.0 50 m above Rt. 259/18 bridge	2015	Kick	4	3.59	Moderately
Black Creek					

03-BLCN-0.6 State Rt. 261	2015	Kick	4	4.92	Moderately
Northrup Creek 03-NRUP-0.8 North Greece rd. bridge; 50m upstream	2015	Kick	7	4.73	Moderately
Round Pond Creek 03-RPON-1.8 Island Cottage Rd. bridge; 20 m downstream	2004	Kick	4	5.13	Slight
Slater Creek 03-STLR-0.1 10 m upstream of Edgemere Dr, bridge	2004	Kick	1	2.31	Severely
Slater Creek 03-SLTR-1.5 15 m downstream of Dewey Ave bridge	2008	Kick	0	2.93	Moderately
	2015	Kick	3	2.91	Moderately
Fleming Creek 03-FLEM-0.8 40 yds above Lotta Rd. bridge.	2015	Kick	4	4.38	Moderately
Unnamed tributary to Genesee River 04-GENS_T3-0.3 Van Voorhis Ave.	2009	Kick	2	2.33	Severely
<i>Genesee River</i> <i>100 m below Rt. 104 bridge,</i> <i>04-GENS-4.7</i> <i>Starboard side</i>	2004	Multiplate	7	8.5	Non
	2009	Multiplate	5	6.7	Slightly
	2014	Multiplate	7	7.5	Non
	2019	Multiplate	9	8.5	Non
<i>Genesee River</i> <i>04-GENS-2.6</i> <i>Genesee Docks at Boxart St.</i>	2004	Multiplate	5	7.9	Non
	2009	Multiplate	9	5.7	Slightly
	2014	Multiplate	4	6.5	Slightly
	2019	Multiplate	5	6.9	Slightly
Shipbuilders Creek End of Forest Lawn Dr.	2015	Kick	5	4.38	Moderately

Table 4: RIBS monitoring locations, sample dates, number EPT species (richness), BAP scores and related impact categories (locations in bold italics are within the AOC boundary)

RIBS Site & Description	Sample year	Ephemeroptera Mayflies	Plecoptera Stoneflies	Trichoptera Caddisflies
Salmon Creek 03-SAMC-5.8 5m below RT 259 bridge	2015	<i>Baetis flavistriga</i>	<i>Paragnetina media</i>	<i>Hydroptila sp.</i> <i>Cheumatopsyche sp.</i> <i>Ceratopsyche sparna</i> <i>Ceratopsyche bronta</i>
Buttonwood Creek 03-BUTN-7.0 50 m above Rt. 259/18 bridge	2015	<i>Baetis sp.</i>		<i>Helicopsyche borealis</i> <i>Hydropsyche betteni</i> <i>Cheumatopsyche sp.</i>
Black Creek 03-BLCN-0.6 State Rt. 261	2015	<i>Baetis intercalaris</i> <i>Baetis flavistriga</i>		<i>Cheumatopsyche sp.</i> <i>Hydropsyche betteni</i>
Northrup Creek 03-NRUP-0.8 North Greece rd. bridge; 50m upstream	2015	<i>Baetis intercalaris</i> <i>Baetis flavistriga</i> <i>Undetermined Heptageniidae</i>		<i>Ceratopsyche bronta</i> <i>Cheumatopsyche sp.</i> <i>Ceratopsyche sparna</i> <i>Psychomyia flavida</i>
Round Pond Creek 03-RPON-1.8	2004	<i>Baetis flavistriga</i>		<i>Cheumatopsyche sp.</i> <i>Hydropsyche betteni</i> <i>Hydroptila sp.</i>

Island Cottage Rd. bridge; 20 m downstream				
Slater Creek 03-STLR-0.1 10 m upstream of Edgemere Dr, bridge	2004	<i>Baetis sp.</i>		
Slater Creek 03-SLTR-1.5 15 m downstream of Dewey Ave bridge	2015	<i>Baetis sp.</i>		<i>Cheumatopsyche sp.</i> <i>Ceratopsyche bronta</i>
Fleming Creek 03-FLEM-0.8 40 yds above Lotta Rd. bridge	2015	<i>Baetis intercalaris</i> <i>Baetis flavistriga</i>		<i>Hydroptila sp.</i> <i>Ceratopsyche sparna</i>
Unnamed Tributary to Genesee River 04-GENS_T3-0.3 Van Voorhis Ave.	2009			<i>Hydropsyche betteni</i>
<i>Genesee River 04-GENS-2.6 Genesee Docks at Boxart St.</i>	2019	<i>Maccaffertium sp.</i> <i>Stenacron interpunctatum</i> <i>Stenacron sp.</i>		<i>Cheumatopsyche sp.</i> <i>Cyrnellus fraternus</i> <i>Hydropsyche orris</i> <i>Hydroptila sp.</i>
<i>Genesee River 04-GENS-4.7 Starboard side</i>	2019	<i>Caenis sp.</i> <i>Baetis intercalaris</i> <i>Stenonema femoratum</i> <i>Stenacron interpunctatum</i>		<i>Cheumatopsyche sp.</i> <i>Cyrnellus fraternus</i> <i>Hydropsyche orris</i> <i>Hydroptila sp.</i> <i>Maccaffertium pulchellum</i> <i>Maccaffertium sp.</i> <i>Maccaffertium teminatum</i> <i>Oecetis sp.</i> <i>Stenacron sp.</i> <i>Stenacron interpunctatum</i> <i>Tricorythodes sp.</i>
Shipbuilders Creek 03-SHIP_T2-0.4 No site description	2015	<i>Baetis tricaudatus</i>		<i>Cheumatopsyche sp.</i> <i>Hydropsyche betteni</i>
Shipbuilders Creek 03-SHIP_0.1 End of Forest Lawn Dr.	2015	<i>Baetis tricaudatus</i>		<i>Cheumatopsyche sp.</i> <i>Ceratopsyche sparna</i> <i>Hydropsyche betteni</i> <i>Hydroptila sp.</i>

Table 5: RIBS monitoring locations, and most recent EPT species collected at those sites (locations in bold italics are within the AOC boundary)

In summary, the monitoring programs and assessments described above demonstrate that:

- Hexagenia was present in suitable habitats of the lower Genesee River between 1999 and 2002, although no known subsequent data exists that would confirm their continued presence in this section of the river. There is no historical evidence that *Hexagenia* was ever present in large numbers in the embayment portion of the AOC.
- Based on a comprehensive assessment in 2013, benthic communities and the toxicity of sediments from sites in the AOC are no different than those from comparable reference sites across the region.

- BAP scores for two sites in the Genesee River within the AOC boundary and monitored via NYSDEC’s RIBS program indicate non or slightly impacted conditions and, while *Hexagenia* have not been detected at these locations, several members of the mayfly and caddisfly families have been found. RIBS monitoring data also show that stonefly, mayfly, and caddisfly families are present in streams tributary to the Rochester Embayment (although, as previously noted, these are not within the AOC boundary as currently defined).

Collectively, these data sets demonstrate that the benthic community within the AOC meet the intent of the removal criterion; that is, while the presence of *Hexagenia* has not been confirmed in recent years, other mayflies and caddisflies have been detected within the river section of the AOC and other indicators of benthic conditions (EPT richness, BAP scores, sediment toxicity) indicate non or slightly impacted conditions throughout the AOC and are similar to regional reference sites.

3.5 Criterion 5: Amphibian diversity and abundance in the study area (including the Genesee River up to the Lower Falls if monitoring can be performed safely) are comparable to expected standards for the type of habitat.

3.5.1 Background

Wetlands provide crucial habitat for different life stages of multiple fish and wildlife species including anurans (frogs or toads), which are sensitive to habitat degradation and environmental stressors (ECCC, USEPA, 2019). They are a key component of wetland fauna and function serving as both predator and prey (Hecnar 2004). As such, anurans are a sub-indicator of ecosystem health widely used by scientists to assess the state of the Great Lakes. For these reasons this BUI removal criterion was developed by the RAC as an endpoint addressing a specific condition that led to the BUI designation, “Amphibian habitat impaired by diversity and density.” The monitoring programs below, which provide data that are being used to assess whether the BUI removal criterion has been met, use anurans as the indicator of amphibian diversity and abundance.

3.5.2 Amphibian diversity and abundance

In 1995, in partnership with USEPA and ECCC, Birds Canada launched the Great Lakes Marsh Monitoring Program (MMP). The goal of the program is to contribute to conservation planning and management and increase public awareness of the value and importance of wetlands. The program relies on volunteers to collect data on frogs, birds and their habitats (data collection protocols can be found [here](#)). This data is used to assess frog and bird populations at scales that range from individual wetlands to the entire Great Lakes basin. One of the primary objectives of the MMP was to contribute to the assessment and long-term monitoring of Great Lake AOCs in terms of anuran and marsh bird relative occurrence, relative abundance, and community structure. In 2005 USEPA funded a special two-year wetland monitoring project to supplement the MMP. The primary objective of the project was to increase MMP coverage in priority marsh habitats within AOC boundaries as well as marshes within the larger source watershed. Using a scoring system called the Indices of Biotic Integrity (IBI) which incorporated multiple metrics including, but not limited to, anuran species richness and abundance values, researchers calculated an IBI score ranging from 0 to 100 for each wetland. Wetlands with higher IBI scores indicate better

biological conditions than wetlands with low scores. Based on these IBI scores researchers determined that among sites within/adjacent to the Rochester Embayment AOC boundary, five sites (Braddock Bay-Buttonwood Creek, Buck Pond, Braddock Bay-West Creek, Cranberry Pond, and Flats Marsh, **Figure 13**) had no impairment to amphibian diversity (Archer et al, 2006), however; four sites (Round Pond, Turning Point Park, East Manitou and Braddock Bay-Salmon Creek, **Figure 13**) were impaired in their ability to support amphibian species. (Archer et al, 2006) (Note: for convenience, and because the adjacent sites are important to the overall habitat restoration that has occurred as previously described in this report, the nine MMP sites are subsequently referred to in the text simply as “AOC sites” despite not all of them being within the AOC boundary.)

Researchers concluded that the majority of the Rochester Embayment AOC sites were in good condition with respect to habitat quality as determined by amphibian community composition and that the sites feature sufficiently suitable marsh habitat coverage to support a high diversity of amphibian species (Archer et al, 2006). In fact, a total of ten different anuran species including all five indicator species (Bull Frog (*Lithobates catesbeinus*), Chorus Frog (*Pseudacris*), Mink Frog (*Lithobates septentrionalis*, Northern Leopard Frog (*Lithobates pipiens*), and Spring Peeper (*Pseudacris crucifer*), as identified by the MMP, were found within the AOC sites ranking the area above average for those Great Lakes basin non-AOC MMP survey locations (Archer et al, 2006).

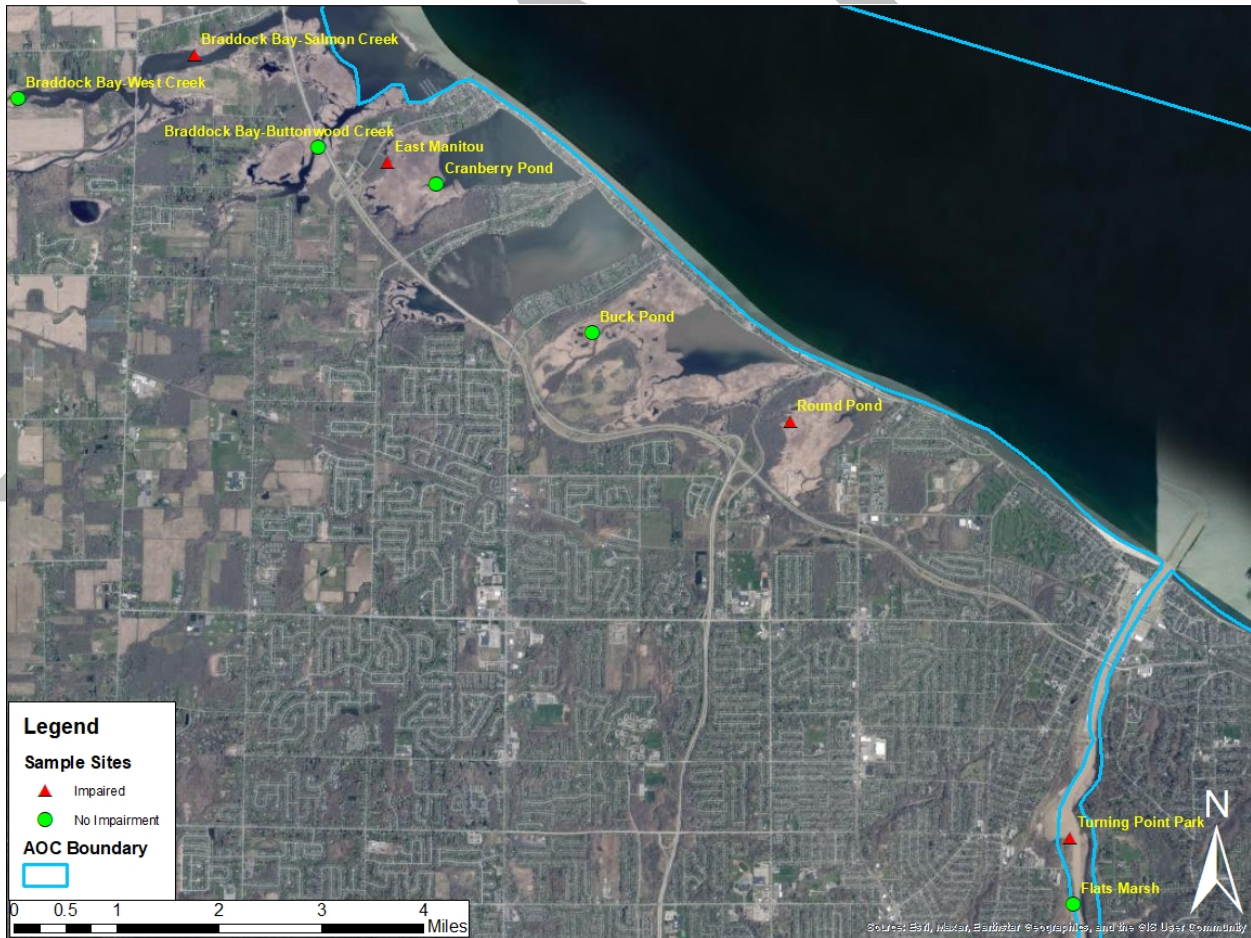


Figure 13: MMP wetland locations

Building off the MMP, in the early 2000s Central Michigan University (CMU) recognized there was not a routine monitoring program in place that could determine the status and trends of Great Lakes coastal wetlands at a basin wide scale. In 2010 CMU launched the Great Lakes Coastal Wetlands Monitoring Program (GLCWMP) to assess status and trends of these wetlands by evaluating multiple parameters including anuran populations. Information regarding data collection protocols and other detailed information can be found [here](#). In 2017 the GLCWMP became a completely developed monitoring system in which data are collected by wetland experts from 15 universities and government agencies from both the U.S. and Canada. Wetlands across the Great Lakes Basin were selected for this project based on four criteria: 1) Must be larger than four hectares; 2) Must have or appear to have a Great Lake connection navigable by fish; 3) Must have herbaceous vegetation; and 4) Must be influenced by lake water levels. In total 1,014 sites were chosen. Approximately 200 sites are randomly surveyed each year on a five-year rotation. Sites termed “benchmark sites” are also surveyed, these are sites of special interest for restoration or protection which may be sampled more than once in the five-year rotation and may not have been on the original survey list (Uzarski, 2021). The Braddock Bay restoration area described in **Section 3.1.2** is a benchmark site.

To date, two five-year sampling rounds have been completed (round one: 2011-2015; round two: 2016-2020) with a total of 1,221 sites surveyed in round one and 978 in round two. Fewer wetlands were sampled in the second round due to the global pandemic. Currently the GLCWMP is in the third year of its third five-year sampling rotation. Over the first two sampling rounds a total of 185 sites were surveyed around Lake Ontario, 11 of which were in or adjacent to the Rochester Embayment AOC (**Figure 14**). Some of these locations were in the same general areas as MMP monitoring sites. (*Note: For the same reasons cited above for the MMP, these 11 GLCWMP sites are subsequently referred to in the text simply as “AOC sites” despite not all of them being within the AOC boundary.*)

Results from the first two survey rounds indicate the mean number of anuran species across the non-AOC sites is 4.9 and that the mean number of anuran species for the AOC sites is 5.1. Although the low number of AOC sites relative to non-AOC sites around Lake Ontario does not allow for a statistical comparison, the mean number of species for the non-AOC and AOC sites appears to be comparable. The benchmark Braddock Bay restoration site was monitored annually in the second round of surveys from 2016-2020. Survey results have shown that the number of anuran species at this site has increased from four to six since 2013 (Wilcox et al 2018).

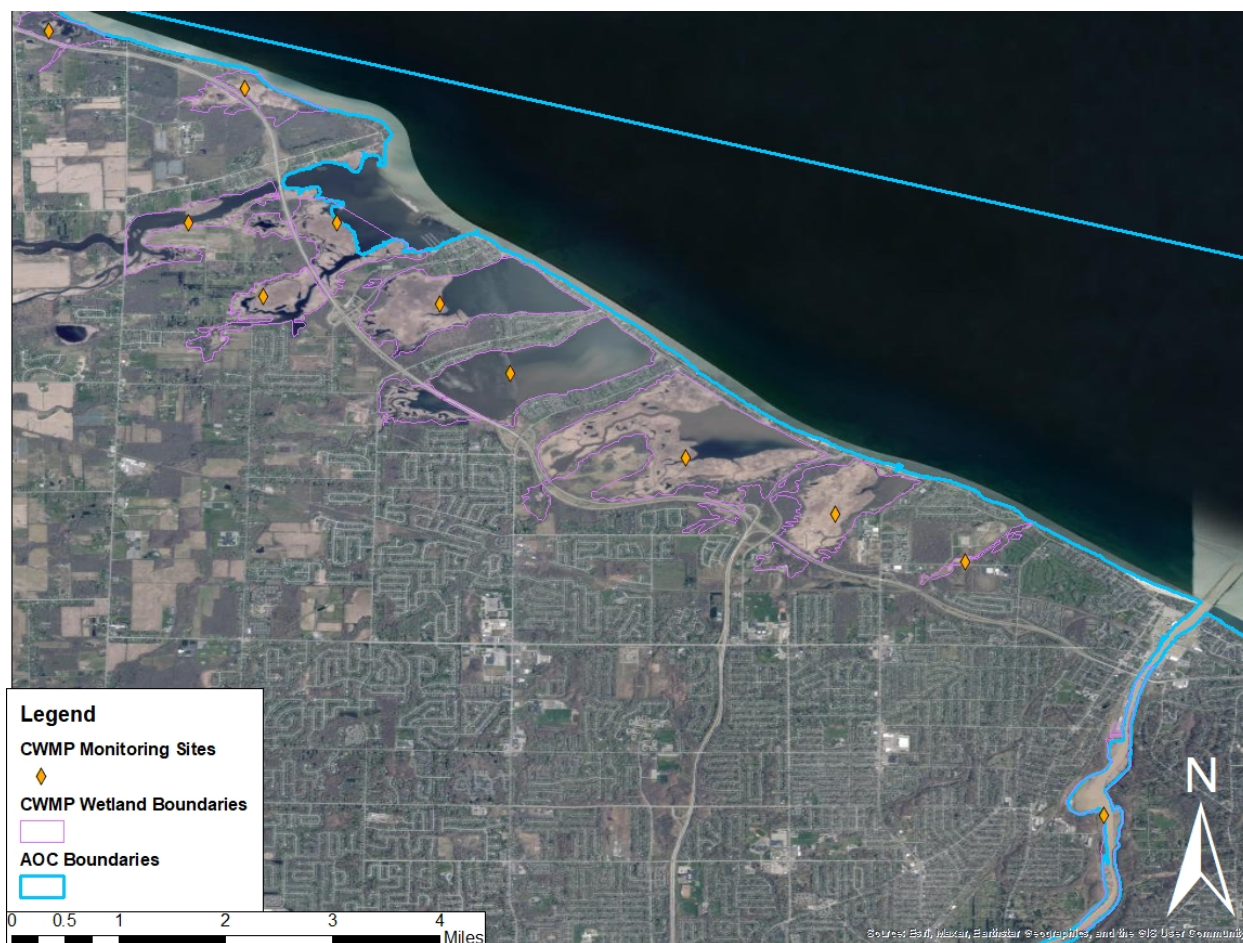


Figure 14: Great Lakes Coastal Wetland Monitoring Program sampling locations within/adjacent to the AOC

Results of the 2005-2006 MMP indicated that, of the nine AOC wetland sites surveyed, five had no impairment in amphibian diversity and four were impaired in their ability to support amphibian species. MMP researchers concluded that the majority of these sites were in good condition with respect to habitat quality as determined by amphibian community composition and that the sites feature sufficiently suitable marsh habitat coverage to support a high diversity of amphibian species (Archer et al, 2006). It should be noted that two of the four impaired sites (Turning Point Park and Braddock Bay) have had habitat restoration projects completed in them (see **Section 3.1.2** for project descriptions and locations).

Furthermore, the results from the two five-year survey rounds (2011-2015 and 2016-2020) conducted by GLCWMP indicate, although not statistically comparable due to small number of sampling sites, that mean number of anuran species within AOC wetland sites appear to be comparable to wetland sites around Lake Ontario. Together, the results of the MMP and GLCWMP studies indicate that habitat is suitable to support a high diversity of amphibian species, total number of species has increased in at least one of the restored wetlands in the AOC, and that the number of species per monitored wetland sites within the AOC appear to be comparable to wetlands around Lake Ontario at 5.1 and 4.9 respectively.

3.6 Criterion 6: Lake sturgeon of different life stages inhabit the Genesee River up to the Lower Falls and the Embayment, OR physical and biological habitat are suitable for sturgeon.

3.6.1 Background

Ecologically and culturally revered, Lake Sturgeon (*Acipenser fulvescens*) is the oldest and largest native fish species in the Great Lakes watershed. Historically in Lake Ontario, the lower Genesee River provided ample habitat for spawning and egg-incubation for Lake Sturgeon. However, settlement and industrial development severely impacted habitat conditions and by the 1930's lake sturgeon were extirpated from the area. In 1983 the sturgeon was listed as a Threatened Species in New York State. The severely impacted habitat conditions led to the BUI designation that the "fishery habitat was impaired by unsuitability for Lake Sturgeon". The RAC Habitat Oversight Committee developed this criterion as an endpoint addressing the extirpation causal conditions.

3.6.2 New York State Lake Sturgeon Recovery Plan

In 1994 NYSDEC developed its first statewide Lake Sturgeon Recovery Plan. The plan was revised in 2000, 2005, and most recently in 2018. The [current plan](#) divides the historic range of sturgeon into seven management units, one of which – the Western Lake Ontario Management Unit – includes the Rochester Embayment AOC, including the lower Genesee River. The goal of the restoration plan is to establish or maintain sufficient self-sustaining populations of lake sturgeon. For a management unit to be considered recovered the unit must have at least 750 sexually mature sturgeon in its population and there must be evidence of ongoing recruitment, demonstrated by three-year classes of wild production within a five-year period. Research to measure the goals of the Lake Sturgeon Recovery Plan inform managers on the status of recovery effort relevant to the delisting criterion.

3.6.3 USGS Sturgeon Population Assessments

During the years of 1999 through 2002 USGS conducted a study titled "Assessment of Habitat Use by Experimentally Stocked Juvenile Lake Sturgeon" of the lower Genesee River within the AOC. The purpose of this study was to assess the area for any remnant population and to evaluate the quality and availability of habitat for all life stages of lake sturgeon over multiple years. The study was designed to analyze the feasibility of restoration of lake sturgeon to an area where the species was once abundant (Dittman et al 2006). Comprehensive benthic fauna and habitat sampling was conducted to provide data to the Lake Sturgeon habitat suitability model, results rated foraging habitat for juvenile and adult sturgeon as good and the overall habitat of the Rochester Embayment AOC as suitable (Dittman et al 2006). Following the investigation, in 2003-2004, 1,900 hatchery raised juvenile sturgeon were stocked in the lower Genesee River. Stocking has continued annually since 2013. As of 2016 the overall population of the lower Genesee River was estimated to be 1,288 ranging in age from 1 to 14 from 6 different cohorts (NYSDEC, 2018). Ongoing monitoring indicate that these fish often move between the lower Genesee River and the Rochester Embayment (Dittman et al 2022). Eighteen years after the initial stocking, in 2021, four ripe males and one mature female were captured ½ kilometer from potential spawning habitat (Dittman et al 2022). This female was one of the 1,900 originally stocked in 2003-2004. It was

1,550 mm in length, weighed 31.6 kg and likely produced ~ 330,000 eggs (Dittman et al 2022). Results of this long-term research demonstrate that sturgeon of different life stages from juvenile to adult inhabit the lower Genesee River and that, as of 2021, spawning is likely taking place. Based on this evidence Criterion 6 has been satisfied.

3.7 Criterion 7: Mink inhabit and reproduce within areas contiguous to the Genesee River and streams within the defined area, OR physical and biological habitat are suitable for mink.

3.7.1 Background

Decreasing mink presence and reproduction in areas of the Genesee River was noted in the Stage I and II RAP documents as one of the indicators of the Loss of Fish and Wildlife Habitat BUI, and for which BUI removal criteria were developed. Since the BUI designation, several mink studies have been conducted within and around the AOC to assess the status of mink presence and reproduction. A summary of these is provided below.

3.7.2 Mink Studies (2003 – 2007)

Researchers at SUNY Brockport conducted a series of studies investigating mink within the Rochester Embayment AOC, focusing on the Braddock Bay Wildlife WMA in the western portion of the AOC. Inland locations and areas along the Lake Ontario shoreline outside the AOC boundary were also included, to serve as reference sites. This work built upon ongoing mink studies at the time by SUNY Brockport researchers and was intended to assess the status of this BUI as well as the Degradation of Fish and Wildlife Populations and Bird or Animal Deformities or Reproduction Problems BUIs. The additional studies were made possible through funding support provided in 2004 from the Great Lakes Protection Fund (GLPF). (*Note: while citations for each of the individual studies described below are provided, the final reports for these studies are included in their entirety as appendices to a larger comprehensive final report prepared by SUNY Brockport as part of the GLPF grant, included as **Appendix E** to this BUI removal report.*)

The first study (Wellman and Haynes, March 2006), conducted in 2003 – 2004, involved the use of a proprietary “MustelaVision” system to attempt to document the actual presence of mink breeding within the AOC. MustelaVision was a remote-controlled video capture system developed for this project that researchers set up in wetland areas and along shorelines at locations where mink were likely to be observed. Researchers selected sites both within the Rochester Embayment AOC and at reference sites outside of the AOC to compare the relative abundance of mink. During the study period, mink families were recorded multiple times at AOC locations by the MustelaVision system, in one instance four young and one adult mink were recorded. The researchers note in their report that mink are normally solitary except for mating pairs and mothers with kits and, since some of the mink captured on video were clearly young, the researchers assumed that these were kits together with their mother. The MustelaVision video provided direct evidence that mink were not only present but successfully reproducing within the AOC.

The second study (Wellman and Haynes, June 2006) centered on the development of a food web model for mink within the Rochester Embayment AOC, to establish levels of mink exposure to bioaccumulative chemical contaminants (BCCs), which could then be used to assess potential

impacts to populations or reproduction. This study involved the trapping and collection of mink for direct tissue analysis of nitrogen and carbon isotope concentrations. The fact that mink were successfully trapped and collected provided additional direct evidence that mink do inhabit areas within the AOC. The relative concentrations of nitrogen and carbon isotopes in the mink tissue samples were used to evaluate the diets of mink both within the AOC and at reference locations, in terms of trophic levels and terrestrial and aquatic food sources. From this information, researchers were able to construct a food web/bioaccumulation model for mink in the AOC that could be used to predict body burdens of BCCs in mink in relation to their diets, based on the concentrations of BCCs in ambient water in Lake Ontario or the AOC. These predicted contaminant body burdens could then be used to assess potential adverse effects to mink populations or reproduction.

This model was subsequently tested by comparing modeled/predicted BCC concentrations in mink tissue against measured tissue concentrations. The mink tissue analyses, and model testing are further described in subsequent third (Pagano and Haynes, March 2007) and fourth (Wellman and Haynes, March 2007) studies, respectively, both of which are summarized below.

The third study (Pagano and Haynes, March 2007) involved the BCC analysis of tissue samples collected from lakeshore and inland populations of mink both within and outside of the AOC. From the resulting analytical data, the researchers determined that there were no significant differences in BCC concentrations in tissue from mink captured inside and outside of the AOC, although mean concentrations were almost always higher in those within the AOC. While tissue contaminant levels in mink are not a direct consideration when assessing attainment of the BUI removal criteria, the results of this study are noted here because the tissue contaminant data were subsequently used as part of testing the food web model developed in the second study above. This model testing is described further below (fourth study).

The fourth study (Wellman and Haynes, March 2007) included a literature review of reports on BCCs in mink tissue corresponding with adverse effects on populations or reproduction. Through a review of existing literature, researchers compiled a list of adverse endpoints (e.g., reproductive failure, kit mortality, litter size reduction, etc.) and linked them to multiple effect levels for various BCCs. Some of the effect levels considered were the lowest-observed-adverse-effect-level (LOAEL), the no-observed-adverse-effect-level (NOAEL), and the lethal concentration for 100 percent of the population (LC100). The LOAEL represents the lowest concentration of a chemical for which there are observed adverse effects to an exposed group, whereas the NOAEL represents the highest dose of a chemical for which there are no observed adverse effects. The LC100 represents the concentration for a chemical for which exposure results in mortality for the entirety of an exposed group. This endpoint is often scaled to different mortality rates; comparatively, a LC10 value would represent a concentration that would result in 10 percent mortality of an exposed group. A table of these ecological endpoints is included in the study report (Wellman and Haynes, 2007).

This fourth study also included testing the model developed in the second study discussed above (Wellman and Haynes, June 2006) that was being used to assess the adverse effects on populations or reproduction, by comparing the modeled/predicted BCC concentrations in mink tissue based on Lake Ontario ambient water concentrations against the actual tissue concentrations from mink

tissue analyses performed in the third study. Comparisons were limited to mink captured along the lake shoreline, since these would presumably have a diet of organisms exposed to the lake water. It was found that the modeled estimates of the levels of mink exposure to BCCs worked well for some, but not all, chemical contaminants being assessed.

A final report was prepared by the SUNY Brockport researchers (Haynes, Wellman, and Pagano, August 2007) tying together all the information from the various studies above. This report is included as **Appendix E** of this BUI removal report and as previously noted, contains the reports from the individual studies above as appendices. The conclusions from the final August 2007 report were that the series of studies documented the presence of mink populations and mink reproduction in the AOC and that analytical, modeling and literature review results all suggest that mink reproduction is unlikely to be impaired in the AOC. The final study report also included a recommendation that additional monitoring of mink be conducted; this additional work is discussed in Section 3.7.3.

3.7.3 Follow-up Mink Studies (2013 – 2015)

Several years after the conclusion of the studies described above, SUNY Brockport researchers conducted a follow-up project to reassess many of the same ecological indicators that had previously been studied, including the presence of reproducing mink populations within the AOC. This was based on the recommendations made as part of the previous studies, and to confirm the status of multiple BUIs, including Loss of Fish and Wildlife Habitat. This follow-up project, made possible through funding support from the GLRI, was similarly comprehensive in scope. The study results below are discussed in more detail in the final report for the project (Haynes and Wellman, December 2015), included as **Appendix F** of this BUI removal report.

To re-assess the status of mink within the AOC, researchers placed non-lethal “black trakka” traps at 20 locations throughout the AOC where mink are likely to be found. These traps were designed to capture mink tracks and document direct evidence of mink presence within the AOC. Additionally, researchers used the USFWS Habitat Suitability Index (HSI) model for mink to estimate habitat suitability at 41 sites within the AOC. The USFWS HSI model uses three criteria (percent of surface water, percent vegetation cover within 30m of the shoreline, and percent shoreline cover within 1m of surface water) to assign a habitat score to a given site, on a scale of 0-1, with a score of 1 representing optimum habitat. The 41 sites were selected using existing aerial imagery to identify potential areas of suitable habitat, and subsequently assessed on foot and via boat by the researchers, who recorded detailed habitat observations. An experienced mink trapper accompanied the researchers during these assessments and provided an “experience-based” HSI for mink. Also using a 0-1 scale, this index was intended to provide a measure of the likelihood of trapping a mink.

As a result of the follow-up study, definitive evidence of mink was found at 10 of the 20 “black trakka” trap locations. Additionally, field staff observed one live mink swimming across the Genesee River. The USFWS HSI model calculated an average value of 0.85 out of 1 at the sites within 100m of the shoreline of the Genesee River portion of the AOC, suggesting that habitat appears to be highly suitable for mink in these areas. Comparatively, a professional trapper hired by the researchers rated the average habitat suitability for mink at 0.43 throughout the study area,

due primarily to steep, rocky slopes along the lower Genesee River and evidence of human activity along the shoreline. As described above, this latter index was not a direct assessment of habitat, rather a measure of the likelihood of trapping a mink. Therefore, a direct comparison of the results from the two HSIs cannot be made. However, the results from both collectively suggest that suitable habitat for mink exists, and there is a moderate chance that mink could be captured, in the 41 locations assessed.

The SUNY Brockport research team also re-evaluated the risk of various BCCs that were noted in RAP documents to be linked to potential reproductive failure in mink within the AOC by collecting samples of common prey organisms from the Genesee River portion of the AOC and analyzing them for chemical contaminants. Exposure of mink to BCCs was estimated through modeling several possible mink diets, which showed that:

- Using the “highest exposure” mink diet found in published literature (92% from aquatic sources) and using mean concentrations of contaminants found in potential mink prey in the Genesee River portion of the AOC, the maximum dietary exposure of mink would be 81% of the LOAEL for total mercury, 23% of the LOAEL for total PCBs, and 69% of the LOAEL for toxic equivalency (TEQ, a measure of dioxin and dioxin-like compounds). These results represent the “worst case” diet scenario. For this study, TEQ included contributions from PCBs, dioxins/furans, and polycyclic aromatic hydrocarbons (PAHs).
- Using the average of six mink diets reported in published literature (65% from aquatic sources) comparable to what mink would eat in the Genesee River portion of the AOC and using mean concentrations of BUI-related contaminants found in potential mink prey in the study area, the dietary exposure of mink would be 48% of the LOAEL for total mercury, 13% of the LOAEL for total PCBs, and 40% of the LOAEL for total TEQ. This is the “likely” diet scenario.

These findings demonstrate that contaminant levels in mink prey within the Genesee River portion of the AOC are not likely resulting in reproductive problems in mink.

3.7.4 Overall Assessment of Mink Studies

The collective findings of the 2003-2007 and 2013-2015 studies summarized in this section show that mink inhabit and reproduce within AOC areas contiguous to the Genesee River and that the physical and biological habitat are suitable for mink, thus demonstrating that the BUI removal criterion has been met.

3.8 Criteria Conclusions

It cannot be overstated the extensive amount of time, funding, and support invested to improve environmental conditions, including fish and wildlife habitat, within the Rochester Embayment AOC since the Stage I/II RAP documents were prepared. While we recognize the limitations of what can be achieved in terms of returning developed areas within the AOC and surrounding watershed back to more natural habitat that would better support fish and wildlife, the restoration work and studies/assessments conducted in recent years have demonstrated that significant

progress has been made addressing the causes of this BUI that were identified in the RAP and that the criteria established to allow for removal of the BUI designation have been met to the extent possible.

Maintenance and monitoring of improved conditions will continue to be challenged and tested by societal pressures (land use, economic development, etc.) and increasingly unpredictable climate patterns and natural forces within the watershed. Additional actions and programs will be implemented, where warranted and as feasible, both within and outside of the AOC, to continue the improvements made under the AOC program. Some of these are described in the next section of this report.

4. Additional Activities Supporting BUI Removal

4.1 2018-2022 Lake Ontario Lakewide Action and Management Plan

The 2018-2022 Lake Ontario Lakewide Action and Management Plan (LAMP) is a binational ecosystem-based action plan to restore and protect the water quality of Lake Ontario and its connecting river systems, the Niagara and St. Lawrence Rivers. This was the first Lake Ontario LAMP under the 2012 amendment of the GLWQA. Subsequent updates are anticipated every five years (the 2023-2027 LAMP is under development as of June 2023). The LAMP is developed by member agencies of the Lake Ontario Partnership which is a collaborative team of natural resource managers led by the governments of the U.S. and Canada, in cooperation and consultation with State and Provincial Governments, Tribal Governments, and watershed management agencies committed to restoring and protecting Lake Ontario, the Niagara River and the St. Lawrence River. In preparing the LAMP, the Lake Ontario Partnership also seeks input from scientists, First Nations, Métis, stakeholders, non-governmental organizations, and the general public.

The purpose of the LAMP is: 1) to summarize the current state of Lake Ontario according to the nine General Objectives of the GLWQA and point out key threats; 2) to outline actions that will be taken to address the threats and contribute to the restoration and protection of water quality in Lake Ontario; and 3) to engage all groups and individuals in the Lake Ontario basin to take action in protecting the water quality in Lake Ontario. LAMP guided project implementation and agency support of the overall goals and objectives of the LAMP are expected to contribute to the continued restoration of beneficial uses within the Rochester Embayment AOC.

4.2 Water Education Collaborative (WEC)/H2O Hero Campaign

To help mitigate the problem of urban stormwater runoff carrying nutrients and pollutants to local waterways, the WEC was established in 2001 based on a recommendation originally made in the Stage II Rochester Embayment RAP. The WEC is comprised of experts, public interest parties, and local stakeholders. The primary goal of the WEC is to raise local stakeholder awareness of environmental issues and opportunities to mitigate them through public engagement and education initiatives. To achieve this goal, the WEC, with the support of Causewave Community Partners (formerly the Advertising Council of Rochester) and the Stormwater Coalition of Monroe County, launched the H2O Hero campaign in 2007 with the vision that individuals can have a positive

impact on local water conditions through awareness and modest changes in certain everyday activities.

Through the H2O Hero campaign, the WEC offers interactive educational resources on the main sources of residential pollution and shows residents how they can reduce their pollution contribution and exhibit more environmentally responsible behavior through the proper use and storage of household and yard care products. By implementing the WEC and the H2O Hero campaign, the population within the larger Genesee River watershed can better understand how they impact local water quality as well as how they can actively participate in efforts to improve and protect their water resources through the reduction of nutrient and pollutant loading to waterways.

4.3 Genesee RiverWatch Initiative

In 2014, the Center for Environmental Information (CEI) commenced the Genesee RiverWatch initiative and since, has passionately and successfully engaged communities within the watershed to provide awareness of critical issues and promote an appreciation for the river and its tributaries. Each year, Genesee RiverWatch facilitates educational and recreational experiences to enhance public knowledge and increase commitment to its future health. In addition to educating and informing, the organization collaborates with state and local partners to seek ways to improve water quality and overall environmental conditions. Through collaborative project design and implementation, Genesee RiverWatch is committed to restoring degraded riparian areas, reducing phosphorus and sediment load, and continuously supports ongoing monitoring efforts.

Within the context of the Rochester Embayment AOC, ongoing collaboration and support of this organization represents a commitment to stewardship and education for the future. The Rochester Embayment will continue to benefit from programs, projects, and initiatives led by the Genesee RiverWatch community, long after delisting.

4.4 Genesee River Nine Element (9E) Plan

In September 2015, NYSDEC approved the Genesee River Basin Nine Key Element Watershed Plan for Phosphorus and Sediment, with the goal of reducing nutrient and sediment loading to the Genesee River. Nine Element Plans are locally developed watershed-scale management plans designed to address known water quality issues. The 9E Plan identifies major sub-basins within the Genesee River watershed, and prioritized nutrient and sediment reduction efforts within these sub-basins. Based on data collected through a suite of scientific studies, load estimates from point and non-point sources are listed, identifying land use and human activities which contribute the greatest negative impacts. From here, the 9E Plan has identified specific management measures and associated load reduction estimates favored to improve water quality throughout the watershed when implemented. Management measures such as grassed waterways, stream bank stabilization, cover crops, buffer strips, and other green infrastructure projects are effective practices identified for reducing phosphorus and sediment transport within and specific to the Genesee River watershed. Additionally, the Genesee River watershed has been designated as an *Agricultural Priority Watershed* under the federal GLRI Action Plan III. The development of the 9E Plan, along with the federal priority designation, serve to provide greater access to State and Federal Great

Lakes funding opportunities that will continue to support projects to reduce watershed sediment and nutrient loads.

4.5 DEC Habitat Management Plans (HMP)

As part of DEC's regular stewardship of State lands, the habitat of the Braddock Bay WMA will continue to be maintained under the guidance of the site's HMP. Site-specific habitat management plans are created for WMAs and other properties overseen by DEC's Bureau of Wildlife. The goals of the plans are to guide habitat management decisions, which benefit wildlife and support wildlife-related recreation. The plans guide management for a ten-year time period. After which time, the plans will be assessed and modified if needed. These plans incorporate management recommendations from:

- Unit Management Plans
- existing WMA habitat management guidelines
- NY Natural Heritage Program's WMA Biodiversity Inventory Reports
- Bird Conservation Area guidelines
- other documents available for individual WMAs

5. Public Outreach

On _____, 2023, NYSDEC and MCDPH held a virtual outreach event to present the rationale for removing the Loss of Fish and Wildlife Habitat BUI to the public. The outreach event consisted of a brief formal presentation of the BUI removal criteria and how each portion of it has been addressed, followed by a question-and-answer session where local stakeholders were provided an opportunity to ask questions directly to NYSDEC and MCDPH staff regarding this BUI removal. Approximately ___ people attended the event, including ___ representatives of federal, state, and local government agencies, ___ from the general public (including members of the AOC RAC), and ___ unidentified participants.

Beginning on _____, 2023, the draft BUI removal report was posted on the MCDPH website for a 30-day period during which the public was encouraged to review the report and to provide any comments or questions via email to MCDPH. ___ comments or questions were formally submitted by the public during this 30-day period (...add additional information as necessary based on public input received).

6. Conclusions

6.1 Removal Statement

The *Loss of Fish and Wildlife Habitat* BUI was originally listed as impaired due to multiple documented ecological issues described in the Stage I RAP, including:

- Loss or degradation of suitable wetland areas for fish and wildlife,
- Degradation of riparian vegetation important for stabilization and habitat,
- Turbid water quality conditions,
- Decreasing macroinvertebrate species indicative of healthy aquatic conditions,

- Lacking amphibian diversity and abundance,
- Loss of juvenile and adult Lake Sturgeon in the Genesee River,
- Decreasing mink presence and reproduction.

Over the past three decades, a significant amount of work has been completed to address the loss of habitat and the specific causes of the BUI noted above. MCDPH and NYSDEC have determined that the established removal criteria for the Loss of Fish and Wildlife Habitat BUI have been met to the extent possible under the AOC Program. Additionally, data presented herein illustrate that some causes underlying the BUI are the result of impacts from the surrounding watershed outside of the AOC, and/or are considered lakewide or regional concerns not unique to the Rochester Embayment AOC. Therefore, under the BUI removal scenarios presented earlier in this report, the *Loss of Fish and Wildlife Habitat BUI* can be removed. The Rochester Embayment RAC fully supports the removal of this BUI (**Appendix A**).

6.2 BUI Removal Steps

	<i>Completed</i>	<i>Date</i>	<i>Step Taken</i>
1.	√	8/1993	BUI first documented as “Impaired” in the Stage I RAP.
2.	√	2002	BUI removal criteria developed and included in Stage II RAP Addendum
3.	√	5/2012	BUI removal criteria revised with RAC consensus.
4.	√	12/2018	RAP advisory committee agreed to proceed forward with BUI removal.
5.	√	12/2022	Initial draft BUI removal report sent to USEPA for review by Technical Review Lead (TRL).
6.			NYSDEC/MCDPH hold public outreach event and 30-day period for public to review draft BUI removal report.
7.			Input from the public during the 30-day period reviewed and incorporated as appropriate into the BUI removal report. NYSDEC completes final modifications to the BUI removal document and provides to USEPA for final review.
8.			NYSDEC submits final report to USEPA.

6.3 Post-Removal Responsibilities

New York State Department of Environmental Conservation

The Braddock Bay Monitoring and Adaptive Management Reports 2016-2021 have shown a positive response to the restoration activities. There is interest in continuing the monitoring to assess the progress of restoration at the Braddock Bay WMA, but future funding and project management would need to be determined. As part of DEC’s regular stewardship of State Land

the habitat of the Braddock Bay WMA will continue to be maintained under the guidance of the site's HMP, as previously noted.

NYSDEC will continue to monitor water quality and the benthic macroinvertebrate community in the lower Genesee River and the surrounding watershed through the RIBS program. NYSDEC will also continue to work with other partners, including federal agencies, to ensure appropriate monitoring of the Genesee River is conducted, where necessary and feasible, as part of binational lakewide management activities such as the Lake Ontario Cooperative Science and Monitoring Initiative (CSMI).

United States Environmental Protection Agency

USEPA will continue to provide funding for RAP/RAC coordination and technical assistance to support the delisting of the Rochester Embayment AOC. USEPA and other federal agencies will also continue to recognize the Genesee River as an Agricultural Priority Watershed within the Great Lakes Basin. After delisting, habitat restoration efforts within the Genesee River watershed and the Rochester Embayment will continue to the extent feasible under the Lake Ontario LAMP, which is a binational plan led in the U.S. by USEPA.

Monroe County Department of Public Health

With USEPA/GLRI funding, MCDPH currently provides a Coordinator for the Rochester Embayment AOC RAP, facilitation with RAC efforts, and technical assistance for AOC documentation and project design. With anticipated continued funding support from USEPA, MCDPH may continue in these roles through the delisting of the Rochester Embayment AOC.

Remedial Advisory Committee

The RAC will continue to forward the objectives of the RAP by evaluating, supporting, and documenting the restoration of the Rochester Embayment AOC, until the long-term goal of delisting the AOC can be achieved.

7. References

Archer, R.A., Timmermans, S.T.A., Robinson, C.L. (December 2006) Monitoring and Assessing Marsh Habitats in Greatlakes Area of Concern Final Project Report.

Dittman, D.E., Chalupnicki, M., Randall, P., Wyatt, J., Zollweg-Horan, E., Sanderson, M., Morton, K., Poster presentation for the May 2022 Great Lakes Area of Concern Conference.

Dittman, D.E., Zollweg, E.C. 2006. Final Report, Assessment of Habitat Use by Experimentally Stocked Juvenile Lake Sturgeon

Duffy, B.T., George, S.D., Baldigo, B.P., Smith, A.J. (February 2017). Assessing condition of macroinvertebrate communities and bed sediment toxicity in the Rochester Embayment Area of Concern, New York, USA. *Journal of Great Lakes Research* Volume 43, Issue 5, October 2017, p. 890-898.

Ecology and Environment, Inc. March 2009. Rochester Embayment Area of Concern Beneficial Use Impairment Delisting Criteria Report.

Ecology and Environment, Inc. June 2011. Interim Rochester Embayment Area of Concern (AOC) Strategic Plan for Beneficial Use Impairment (BUI) Delisting

Environment and Climate Change Canada., U.S. Environmental Protection Agency. State of the Great Lakes -2009 Technical Report.

Environment and Climate Change Canada., U.S. Environmental Protection Agency. State of the Great Lakes -2019 Technical Report.

Haynes, J.M., Wellman, S.T. December 2015. Final Report: BUI Delisting Studies in the Rochester Embayment AOC, 2013-2014.

Haynes, J.M., Wellman, S.T., and Pagano, J.J. August 2007. RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, Trophic Relationships, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species. Final Report to the New York State Great Lakes Protection Fund.

Hecnar, S.J., Great Lakes Wetlands as Amphibian Habitats: A Review. *Aquatic Ecosystem Health and Management* 1 April 2004; 7(2): 289-303.

International Joint Commission. 1991. Restoring Beneficial Uses in Areas of Concern.

International Joint Commission (IJC). 1991. Proposed listing/delisting guidelines for Great Lakes Areas of Concern. Focus on International Joint Commission Activities. Volume 14, Issue 1, Insert. 1991.

New York State Department of Environmental Conservation (NYSDEC) and Monroe County Department of Planning and Development. August, 1993. Rochester Embayment Remedial Action Plan Stage I.

NYSDEC and Monroe County Department of Health (MCDOH). September, 1997. Rochester Embayment Remedial Action Plan Stage II.

NYSDEC and Monroe County Department of Health (MCDOH). 2002. Rochester Embayment Remedial Action Plan 2002 Addendum.

NYSDEC December 2011, Addendum to Stage 1 and 2 Remedial Action Plans Rochester Embayment Area of Concern.

NYSDEC April 2017, Rochester Embayment Area of Concern Remedial Action Plan Beneficial Use Impairment Removal Report - Degradation of Benthos.

NYSDEC January 31, 2018. Lake Sturgeon Recovery Plan 2018-2024.

Pagano, J.J, Haynes, J.T. March 2007. Levels of Bioaccumulative Chemicals of Concern in Mink In and Out of the Rochester Embayment Area of Concern and Along and Inland from the Shore of Lake Ontario.

Schultz, Rachel, et al. 2021. February Braddock Bay Restoration, 2020 Monitoring and Adaptive Management Report

Schultz, Rachel, et al. 2022. February Braddock Bay Restoration, 2021 Monitoring and Adaptive Management Report

Shear, H., Stadler-salt, N., Bertram, P., & Horvatin, P. (2003). The development and implementation of indicators of ecosystem health in the great lakes basin. *Environmental Monitoring and Assessment*, 88(1-3), 119-52. doi:<https://doi.org/10.1023/A:1025504704879>.

Smith, A.J., Heitzman, D.L., Lojpersberger, J.L., Duffy, B.T., Novak, M.A., 2012. Standard Operating Procedure: Biological Monitoring of Surface Waters in New York State. NYSDEC, Albany, NY.

U.S. Army Corps of Engineers, Buffalo District. September 2014. Braddock Bay Restoration Feasibility Study.

U.S. Army Corps of Engineers, Buffalo District. March 2017 Braddock Bay Restoration, 2016 Monitoring and Adaptive Management Report.

U.S. Army Corps of Engineers, Buffalo District, March 2018 Braddock Bay Restoration, 2017 Monitoring and Adaptive Management Report.

USEPA. 2014. —Rochester Embayment Area of Concern Web Site. Available online at: <http://www.epa.gov/glnpo/aoc/rochester.html>. Retrieved 01-27-14.

U.S. Fish and Wildlife Service, New York Field Office. March 2014. Wetland Assessment in the Rochester Embayment Area of Concern in Support of the Loss of Fish and Wildlife Habitat BUI Removal Evaluation.

United States Policy Committee (USPC). 2001. Restoring United States Areas of Concern: Delisting Principles and Guidelines. Adopted by the US Policy Committee on December 6, 2001.

Uzarski, D.G. April 2021 Continuation of the Great Lakes Coastal Wetland Monitoring Program..

Wellman, S.T., Haynes, J.T. March 2006. Are there Differences in the Relative Abundance of Lakeshore and Inland Mink Populations In and Out of the Rochester Embayment of Lake Ontario Area of Concern? Monitoring Mink Populations Using Video Traps.

Wellman, S.T., Haynes, J.T. June 2006. Age, Size, and Stable Isotope Data of Mink Populations, and a Predictive Model of Bioaccumulation of Chemicals of Concern in the Rochester Embayment of Lake Ontario.

Wellman, S.T., Haynes, J.T. March 2007. Bioaccumulative Chemicals of Concern in Mink: Adverse Effects Levels and Results of a Predictive Model for the Rochester Embayment of Lake Ontario.

Wilcox, Douglas A., et al. 2008. "Cattail Invasion of Sedge/grass Meadows in Lake Ontario: Photointerpretation Analysis of Sixteen wetlands over five decades." *Journal of Great Lakes Research* 34.2 (2008): 301-323.

Wilcox, Douglas A., et al. March 2018. Braddock Bay Restoration, 2017 Monitoring and Adaptive Management Report.

Wilcox, Douglas A., et al. March 2019. Braddock Bay Restoration, 2018 Monitoring and Adaptive Management Report.

Wilcox, Douglas A., et al. January 2020. Braddock Bay Restoration, 2019 Monitoring and Adaptive Management Report.

DRAFT

Appendix A: List of Remedial Advisory Committee Members & Letter of Support

DRAFT

Remedial Advisory Committee Members

Starr O'Neil
 Rochester Embayment Area of Concern - Remedial Action Plan Coordinator
 Monroe County Dept. of Public Health
 111 Westfall Road – Room 828
 Rochester, NY 14620
 585-753-5209
soneil@monroecounty.gov

Name	Organization	E-mail
Anne Spaulding	City of Rochester	Anne.Spaulding@CityofRochester.Gov
Charles Valeska	General Public	CHAZVAL46@YAHOO.COM
Charlie Knauf	General Public (MCDPH retiree)	anniebl@frontiernet.net
George Thomas	Genesee RiverWatch	gthomas@geneseeriverwatch.org
Jane Forbes	City of Rochester	forbesj@cityofrochester.gov
Jeff Wyatt	URMC	Jeff_Wyatt@URMC.Rochester.edu
June Summers	Genesee Valley Audubon Society	summers@frontiernet.net
Paul Flansburg	Great Lakes Comm., Sierra Club	pflansburg@hotmail.com
Paul Sawyko	Water Education Collaborative	psawyko@rmsc.org
Stevie Adams	The Nature Conservancy	sadams@tnc.org
Wayne D. Howard	Solara Concepts	whoward@solaraconcepts.com
Agency Staff		
Peter Rightmyer	MCDPH	prightmyer@monroecounty.gov
Sara Madison	MCDPH	saramadison@monroecounty.gov
Craig Pettinger	NYSDEC	Craig.Pettinger@dec.ny.gov
Andrea Pedrick	NYSDEC	andrea.pedrick@dec.ny.gov
James Lehnen	NYSDEC	James.lehnen@dec.ny.gov
Jennifer Dunn	NYSDEC	jennifer.dunn@dec.ny.gov

May 9, 2023

James Lehner

NYS Great Lakes Areas of Concern Coordinator
New York State Department of Environmental Conservation
700 Delaware Avenue
Buffalo, NY 14209

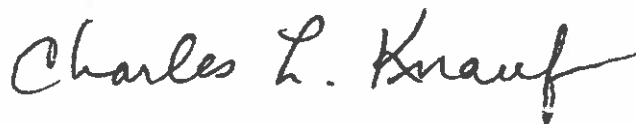
Dear Mr. Lehner:

This letter is to indicate the continued support of the Public Advisory Committee (PAC) for the removal of the *Loss of Fish and Wildlife Habitat* beneficial use impairment (BUI) for the Rochester Embayment Area of Concern (AOC).

During a meeting of the PAC on December 10, 2018, it was documented that the removal criteria for this BUI had been met and that we were ready to move forward with preparing the removal report, with NYSDEC taking the lead drafting role.

Recently, the members of the PAC were provided a draft version of the report for review. No subsequent concerns about the report or proceeding with the BUI removal process have been raised by the PAC members.

Sincerely,



Charles Knauf, Public Advisory Committee Chair

Cc: Starr O'Neil, MCDP

DRAFT

New York State Department of Environmental Conservation

Great Lakes Programs

270 Michigan Avenue, Buffalo, New York 14203-2915

Phone: (716) 851-7070 • Fax: (716) 851-7009

Website: www.dec.ny.gov



Joe Martens
Commissioner

June 25, 2014

Mr. Chris Korleski, Director
U.S. Environmental Protection Agency
Great Lakes National Program Office
77 W. Jackson Boulevard (G-17J)
Chicago, IL 60604-3511

**RE: Management Actions Required to Delist
Rochester Embayment Area of Concern (AOC)**

Dear Mr. Korleski:

Let me begin by expressing the New York State Department of Environmental Conservation's ("DEC") full commitment to the restoration of the lower Genesee River and Rochester Embayment and delisting of the Rochester Embayment Area of Concern (AOC), as well as acknowledging the outstanding efforts by the Monroe County Department of Health in coordinating the development and implementation of the Remedial Action Plan (RAP). The DEC truly hopes our partnership with Monroe County government as the AOC/RAP Coordinator will continue.

This letter is to advise you of the remaining management actions needed to satisfy delisting targets for removing the Beneficial Use Impairments (BUI) by a planned target date of late 2016, and ultimately full restoration of the Rochester Embayment AOC. DEC is defining "management actions" as key commitments to a federal or state program, major project or strategic set of projects that are intended to achieve significant restoration of water quality and water dependent resources, consistent with the Great Lakes Water Quality Agreement, Great Lakes Restoration Initiative or Great Lakes Legacy Act.

These management actions, identified by the DEC and Rochester Embayment Remedial Action Plan (RAP) Coordinator, are endorsed by the AOC's Remedial Advisory Committee and are considered to be generally supported by the local community.

The following list summarizes the management actions deemed necessary to satisfy the BUI restoration criteria/targets and ultimately delist the AOC:

- Recommend renewing the Great Lakes Restoration Initiative (GLRI) funding agreement enabling Monroe County government to continue serving as AOC/RAP Coordinator.
- Complete all ongoing and scheduled field assessments needed to satisfy existing BUI removal criteria (enclosure (a)) including needed follow-up actions based on results of assessments through GLRI funds;
- Design and implement a set of priority habitat restoration projects identified in the Rochester Embayment AOC Habitat Restoration Action Plan (enclosure (b)) through a combination of GLRI and other available funds, including an adaptive monitoring/management plan to evaluate ongoing restoration success or to adjust accordingly;
- Complete BUI removal activities for the following impairments: Fish Tumors BUI, Beach Closings BUI, Restrictions on Fish & Wildlife Consumption BUI, Restrictions on Dredging BUI, Degradation of Benthos BUI, Bird & Animal Deformities BUI, Degradation of Plankton Populations, Degradation of Fish and Wildlife Populations and Habitat BUIs, and Eutrophication & Aesthetics BUIs; and,
- Prepare an AOC Stage 3 delisting report and a post-delisting monitoring plan to include a one-time monitoring period implemented 5-10 years following delisting that is aligned with existing federal, state and local government program capabilities, and is designed to most cost-efficiently assure that the management actions achieved the desired conditions required to delist the AOC.

Based on the data and information currently available, the DEC believes that completion of these management actions, notwithstanding any unforeseen circumstances (i.e., unanticipated results from current and planned BUI criteria monitoring/data assessments, the future development and implementation of project-specific designs and cost estimates, availability of funding, etc.), will place the AOC on a trajectory for ecological recovery to sufficiently justify delisting from New York's and the U.S. EPA's Water Quality Management Programs within the next couple years. As demonstrated by NY's leadership with the Oswego River AOC, the DEC intends to work with EPA, involved federal agencies, the RAP Coordinator and Advisory Committee, and our local partners to carry-out the formal delisting process once the targets of BUI removal are attained.

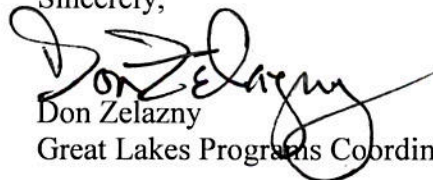
Furthermore, apart from these AOC management actions necessitated by the RAP, the lower Genesee River will be subject to investigation and potential remediation from a number of potential sources of contamination including RCRA corrective action requirements associated with Eastman Business Park (formerly known as Eastman Kodak Company's Kodak Park facility). It is anticipated that the DEC will perform a remedial investigation of the lower

Genesee River and subsequently determine remedial response actions that are needed to further restore and improve the riverine and riparian areas of the lower Genesee River.

The Genesee River and Rochester Embayment “community,” including a range of federal, state and local government agencies, business leaders and community action organizations, are working hard to help assure the ultimate restoration of the overall area is sustained through a series of ongoing or additional regulatory and related actions including (a) the Monroe County Pure Waters Project within the City of Rochester for reducing combined sewer overflows and promoting the benefits of green infrastructure, (b) implementing environmental remediation measures when warranted through the DEC’s State Superfund and Hazardous Waste Management programs to control pollution sources, (c) enforcing a City of Rochester ordinance establishing “Critical Environmental Area” designations which trigger a State Environmental Quality Review (SEQR) for development within the designated areas. This designation protects sensitive areas along the Genesee River gorge, helps to maintain upland habitat connectivity and enhances public access to the waterway, (e) integrating the Regional Economic Development & Community Sustainability Plans with Brownfield Opportunity Area and Environmental Justice programs, and (e) promoting a greater sense of river stewardship and appreciation through ongoing public education and river experiences spearheaded by local watershed groups like the Water Education Collaborative and the Center for Environmental Initiatives.

We look forward to continuing our joint efforts to restore and delist New York’s Areas of Concern. If you have any questions regarding this matter, feel free to contact me at (716) 851-7070 or email at dezela@gw.dec.state.ny.us; or contact Gerry Pratt, NYSDEC’s Statewide AOC Coordinator with detailed questions/comments at 518-402-8246; email ghpratt@gw.dec.state.ny.us.

Sincerely,


Don Zelazny
Great Lakes Programs Coordinator

Enclosure (a): Beneficial Use Impairment Assessments Scheduled for Completion
Enclosure (b): Rochester Embayment AOC Priority Habitat Restoration Action Plan projects

cc: Dave Cowgill, USEPA/GLNPO
Richard Balla, USEPA Region 2, Chief Watershed Management Branch
Charles Knauf, Monroe County Department of Health, Rochester Embayment RAP
Coordinator
James Tierney, NYSDEC, Assistant Commissioner of Water Programs

Enclosure (a) - - Beneficial Use Impairment Assessments Scheduled for Completion

1. A resumption of Genesee River Water Column Phenol testing is in process and completion is expected summer 2014. If phenol levels are still above the removal criteria, it's likely a source outside the AOC associated with coal gasification sites currently in the Superfund Program, and responsibility can be deferred to that program.
2. A study of mink prey tissue is in process by SUNY Brockport and completion is expected summer 2014. If prey tissue analysis demonstrates levels of contaminants do not adversely affect mink this sampling would meet delisting targets for the BUI. In the event that the BUI targets are not met, the impairment is most likely due to sediment and remedial actions would have to be considered.
3. An Invertebrate study by NYSDEC & USGS is in process and will be finalized fall 2014. If invertebrate analysis demonstrates little or no impact this sampling would likely meet sampling targets for the BUI. In the event that the BUI targets are not met, the impairment is most likely due to upstream or lake-wide water quality issues as such there are no additional management actions that can be taken by the RAP.
4. Implementation of Ontario Beach Algae Control Project is needed to mitigate algae not due to lake-wide conditions. Implementation expected Summer 2014. Impoundment of algae due to jetty structure is leading cause of persistent decomposing algae in the AOC and causes beach closure. Beach Best Management Practices (BMPs) implementation for Algae mitigation is to address beach closures and improve aesthetics. Completion of the Beach BMP is expected to meet BUI-criterion target. If subsequent monitoring of the Beach BMP proves effective, this project would meet delisting targets and the RAP Coordinator would recommend removal of the BUI.
5. Study of sediment sources in the river is in process by USGS and NYSDEC, its completion is expected spring 2015. If analysis demonstrates the AOC is not a significant source of sediment it would meet delisting targets. In the event that the BUI targets are not met, the impairment is most likely due to upstream water quality issues and there are no additional management actions that can be taken by the RAP.
6. A plankton study by NYSDEC & USGS is in process and will be finalized fall 2014. If analysis demonstrates little or no impact this sampling would meet specified targets for removing the BUI. In the event that the BUI targets are not met, the impairment is likely due to upstream or lake-wide water quality issues and responsibility will be deferred to another management program.

Enclosure (b) - - Rochester Embayment AOC Habitat Restoration Action Plan

Completion of the following list of projects will satisfy restoration criteria established within the Remedial Action Plan for the removal of the “Degradation of Fish and Wildlife Populations” and the “Loss of Fish and Wildlife Habitat” BUIs:

Sites USFWS Agreed to Implement	Sites which require funding for design and implementation	Sites Fully Funded and in Process
Genesee River –Turning Basin Genesee River- Turning Point Park Confluence of West and Salmon Creek Long Pond West Buck Pond East	Braddock Bay Alternate 7c	Buck Pond Phase I-DU Buck Pond Phase II-DU Braddock Bay- USFWS Salmon Creek-USFWS

Project selection was based on a USFWS study and a separate USACE study of habitat within the AOC. Sites selected for restoration represent the most degraded habitat areas within the AOC in terms of losses in quality and quantity of emergent wetlands, a primary criterion for removal of the Fish and Wildlife Habitat BUI. Emergent wetlands were the habitat of interest with respect to black tern and mink being chosen as indicators by the RAC. Sites recommended by USFWS and USACE were prioritized and vetted for consistency with the BUI criteria through the Remedial Advisory Committee and NYSDEC. The other criteria related to the habitat BUI are being fully satisfied through other evaluations and, pending final results of those assessments, no further management actions are presently anticipated to delist the habitat and fish & wildlife population BUIs for those additional criteria.

The DEC will work with the RAP Coordinator, Remedial Advisory Committee, U.S. Fish and Wildlife Service (USFWS), U.S. Army Corps of Engineers (USACE) and U.S. Environmental Protection Agency to develop and implement an adaptive monitoring and management strategy to be applied to all future habitat projects within the AOC. The DEC anticipates that scheduling of the additional habitat improvement projects by USFWS and USACE will entail a multi-year set of activities, subject to federal interagency agreements, with final feasibility study and design anticipated to occur in 2014-15, and contracting/construction to be completed by end of 2016, commensurate with the planned completion of other management activities. The DEC will also work with all involved agencies and organizations to promote constructive public involvement throughout the project design and adaptive monitoring/management strategy development processes.

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

Great Lakes Programs

270 Michigan Avenue, Buffalo, NY 14203-2915

P: (716) 851-7070 | F: (716) 851-7009

www.dec.ny.gov

March 29, 2016

Mr. Christopher Korleski
U.S. Environmental Protection Agency
Great Lakes National Program Office
77 West Jackson Blvd. (G-17J)
Chicago, IL 60604 Addressee

RE: Rochester Embayment Area of Concern
Modification to AOC Management Actions Letter of June 25, 2014

Chris
Dear Mr. Korleski:

This letter is to advise and seek concurrence from the United States Environmental Protection Agency (USEPA), Great Lakes National Program Office (GLNPO) of the need to modify the management actions previously adopted by the New York State Department of Environmental Conservation's (Department) for restoring the Rochester Embayment Area of Concern (AOC). The basis for this change is described below and is supported by the attachments to this letter.

In 2012 the United States Fish and Wildlife Service (USFWS), in collaboration with the Rochester Embayment AOC Remedial Action Committee (RAC), the Department and USEPA developed a priority list of habitat restoration projects to be completed as one management actions needed to remove the "Loss of Fish and Wildlife Habitat" Beneficial Use Impairment (BUI).

Two of the listed habitat restoration projects (referred to as the Turning Basin and Turning Point wetlands) are within the boundaries of an ongoing investigation of the Lower Genesee River related to the Kodak/Eastman Business Park (EBP) site under the Department's Resource Conservation and Recovery Act (RCRA) program. This investigation, referred to under RCRA as EBP Operable Unit 5 (OU-5) and herein simply as the RCRA project, is focused on the river section from the Lower Falls to Lake Ontario (Figures 1-2). The investigation objectives are to determine the nature and extent of contamination in the Lower Genesee River related to historic operations of EBP, and whether there are any continued loadings of contaminants entering the lower reach from such operations. In addition, the investigation will determine whether remedial action is warranted, and if so, identify and evaluate cleanup options. The RCRA project is funded through an environmental trust created during settlement of Kodak's bankruptcy and administered by the Department. Sampling in the Lower Genesee River began in 2015 and an investigation report is anticipated in 2016.



Department of
Environmental
Conservation

Previous sampling by EPA's Great Lakes Legacy Act program found no significant exceedances of legacy contaminants contributing to AOC BUIs.

Based on the Department's experience with other RCRA cleanup projects, the timetable needed to complete the evaluation (including any supplemental investigation) and implement the corrective measures (if warranted) is on the order of two to five years or more. Since the Turning Basin and Turning Point wetland restoration projects hinge on the outcome and schedule of the RCRA project and any necessary cleanup, it is unlikely that construction of these projects would begin for several years. Furthermore, the Department anticipates a subsequent two to five years of post-construction habitat monitoring are required before it can pursue the BUI removal. The Department is otherwise on track to delist the entire Rochester Embayment AOC by 2021, subject to favorable post-habitat construction monitoring results from other habitat restoration sites included in the RAP management actions.

Regarding the two wetland habitat sites being deleted from the current management action, the Remedial Advisory Committee (RAC) received a letter from the USFWS in February 2016 (included as Attachment 1), in which the USFWS committed to implementing the Turning Basin and Turning Point projects after the RCRA project and any corrective measures are completed. In the event that these projects are no longer feasible at that time, the USFWS will implement other appropriate projects within the AOC boundary of the Genesee River, as specified in the letter. The letter also explains that these habitat restoration projects will be funded by the Natural Resource Damage (NRD) settlement with Kodak.

Subsequently, at the March 16, 2016 RAC meeting in Rochester (attended by USEPA Region 2 and GLNPO), the committee discussed how to proceed with the management action and BUI removal, and ultimately AOC delisting, given the circumstances of the RCRA project. As documented in the meeting minutes (included as Attachment 2), the RAC ultimately recommended that the State request to modify the list of management actions to exclude the Turning Basin and Turning Point projects based upon the conditions described herein and summarized below:

- The RCRA project in the Lower Genesee River is expected to significantly delay construction of these habitat restoration projects, in turn causing significant delays to the BUI removal and AOC delisting schedule;
- The RAC is confident that the USFWS is dedicated to implementing these projects (or alternative habitat restoration projects in the AOC, if applicable) after any RCRA in-water corrective action;
- The two lower Genesee River habitat restoration projects will be funded through NRD funding. Therefore, it is no longer necessary to complete the projects using federal funding through the AOC program; and
- By modifying the list of management actions, the RAC remains on the fast track to delisting the AOC in 2021.

Accordingly, the modified list of habitat restoration projects that satisfies the management action to address the Loss of Fish and Wildlife Habitat BUI consists of the following:

1. USFWS – Confluence of West and Salmon Creek
2. USFWS – Long Pond West
3. USFWS – Buck Pond East
4. USACE – Braddock Bay Alternative 7c
5. DU – Buck Pond Phase I
6. DU – Buck Pond Phase II
7. USFWS – Braddock Bay
8. USFWS – Salmon Creek

Therefore, the Department requests GLNPO's concurrence with this modified management action and the revised set of habitat restoration actions. All other management actions and supporting actions identified in my June 25, 2014 letter remain the same.

If you need further information concerning this request, please contact me or Josh Haugh, New York's new Statewide AOC Coordinator at 518-402-8199 or joshua.haugh@dec.ny.gov.

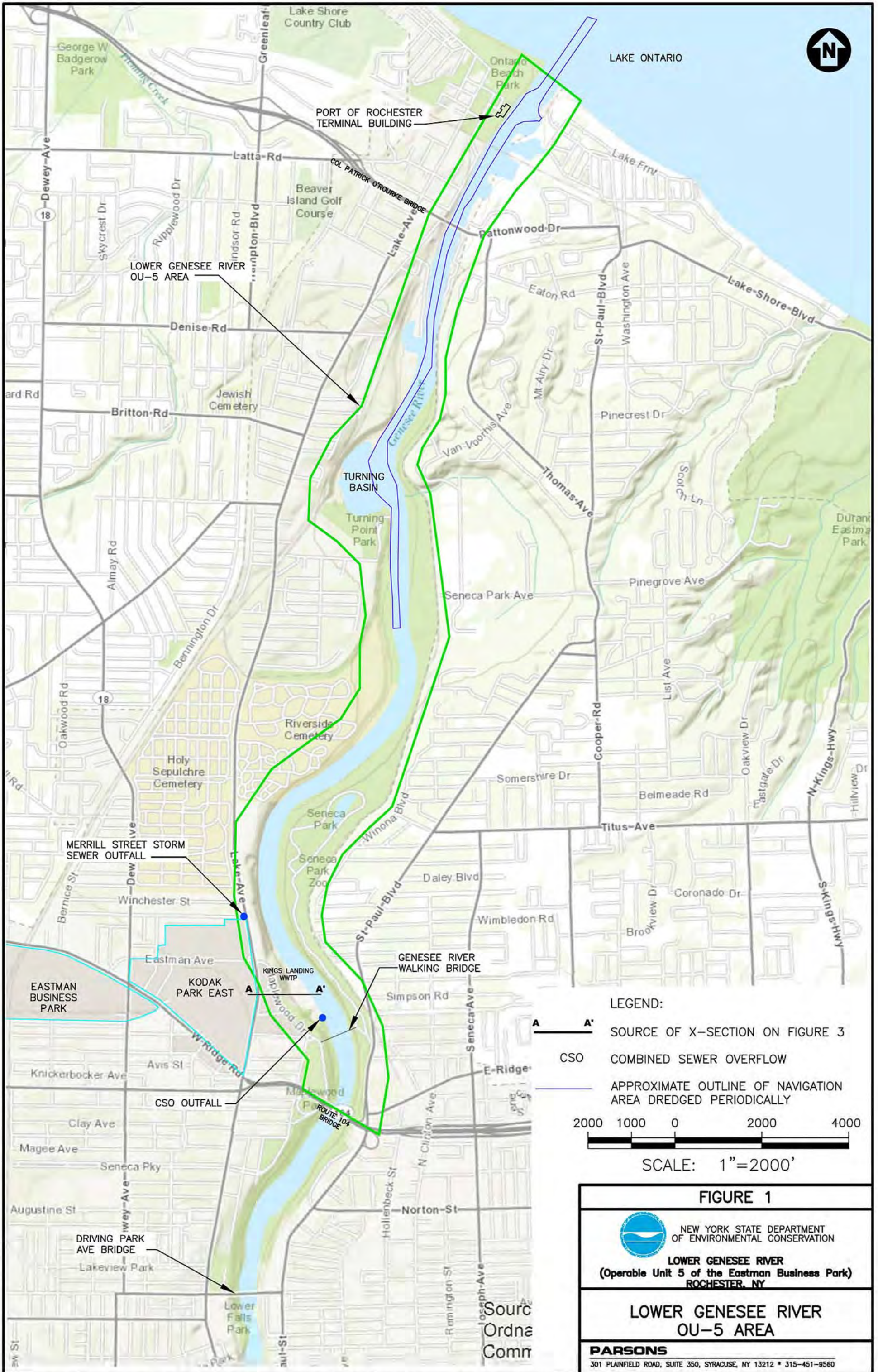
Sincerely,

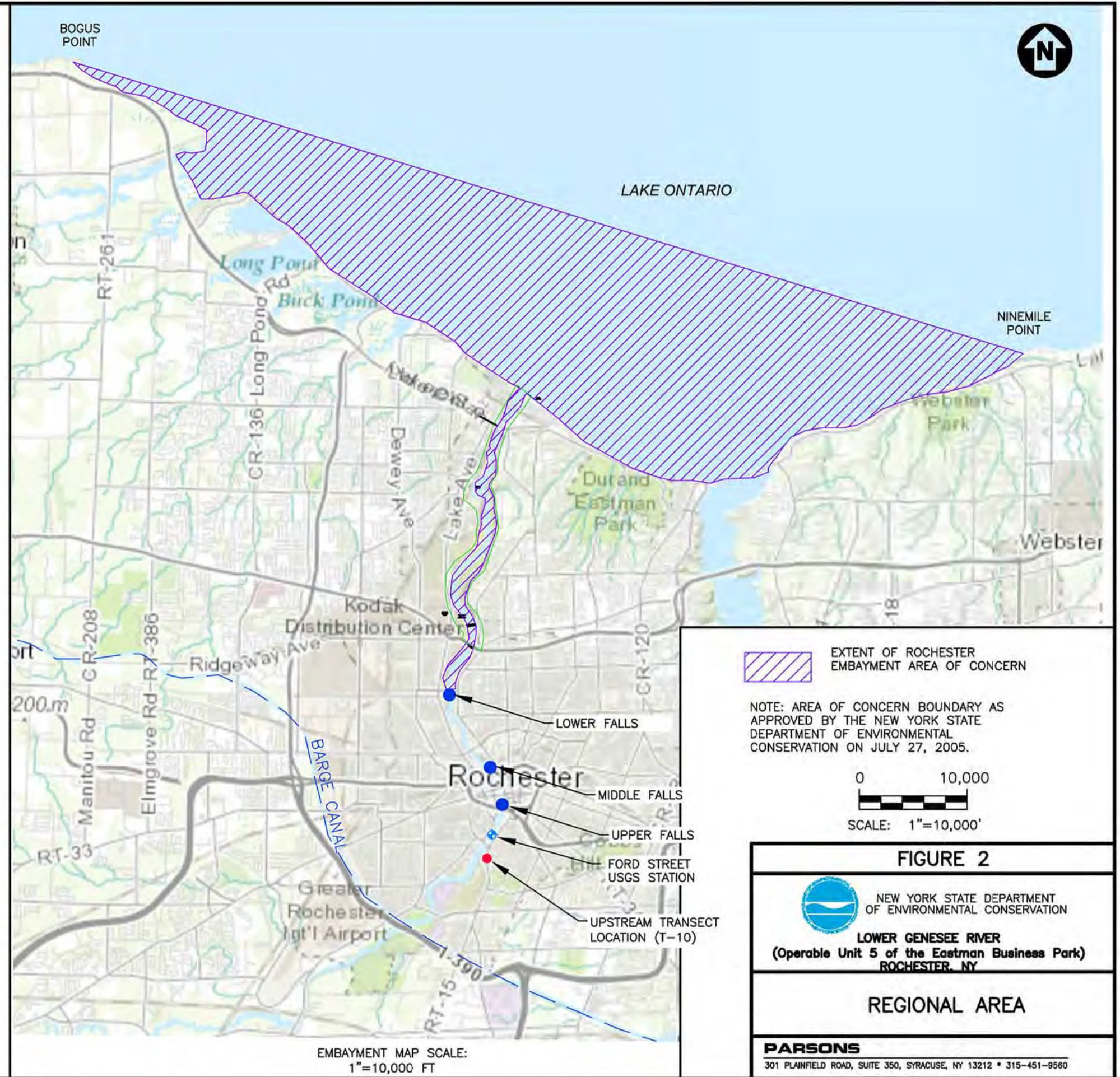
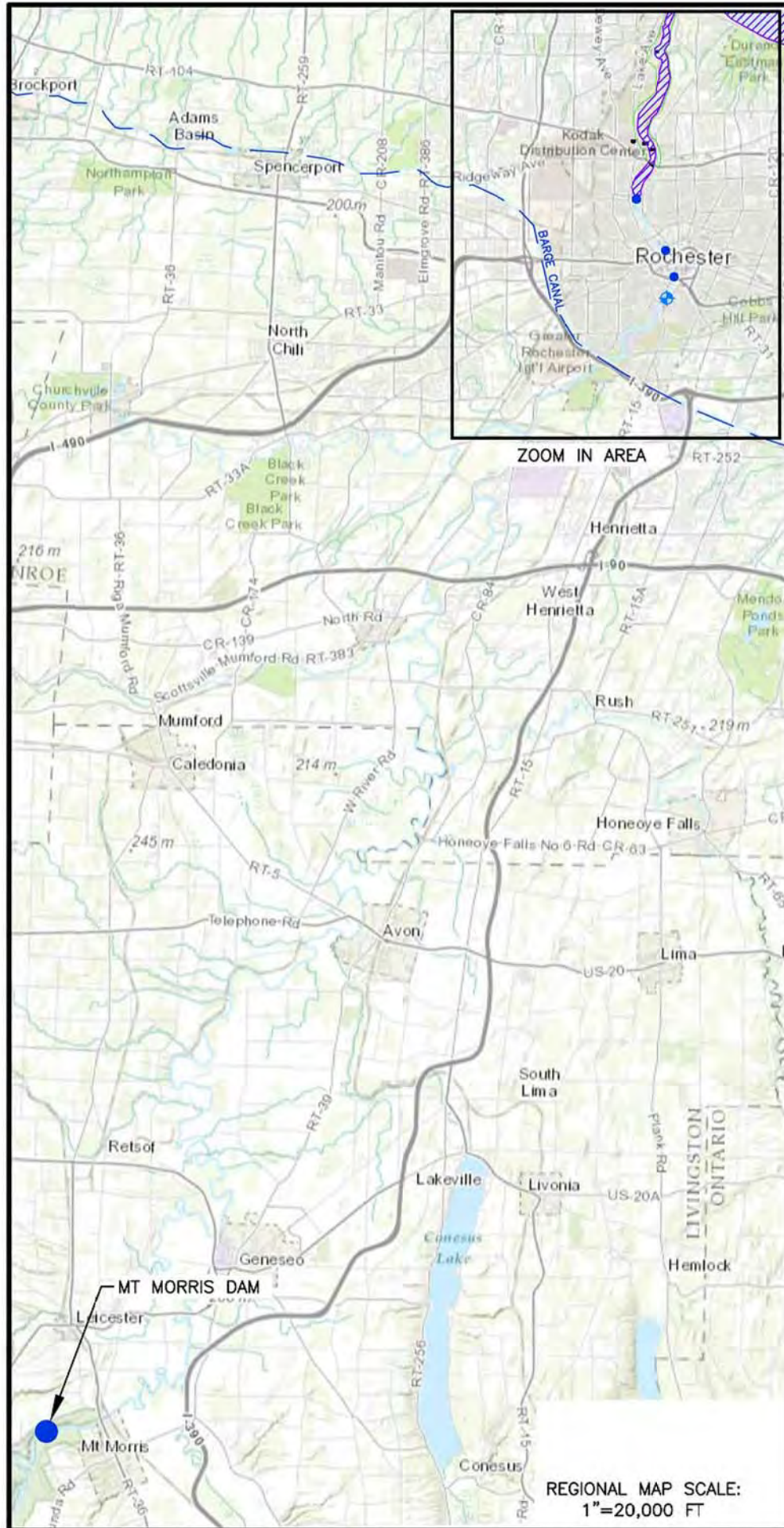


Donald Zelazny
Great Lakes Programs Coordinator

Ec: B. Jones, USEPA GLNPO
J. Haugh, NYSDEC
C. Knauf, RAC Chair
W. Silkworth, MCDOH
F. Luckey, USEPA Region 2

Encl (4)





Attachment 1



United States Department of the Interior



FISH AND WILDLIFE SERVICE

3817 Luker Road
Cortland, NY 13045

February 16, 2016

Dear Rochester Embayment Area of Concern Remedial Action Committee:

In 2014, the United States Department of the Interior (DOI), acting through the United States Fish and Wildlife Service (USFWS), and the State of New York, acting through the New York State Department of Environmental Conservation (NYSDEC), collectively the Trustees, resolved a natural resource damage claim against Kodak for the Genesee River located in the City of Rochester, Monroe County, New York. The Trustees' preferred restoration alternatives include a suite of restoration projects from restoration alternative categories that compensate for interim losses and satisfy the regulatory and site-specific criteria listed within the Draft Restoration Plan for the Genesee River and Genesee River Watershed, New York (Draft Restoration Plan). This letter is to inform the Rochester Embayment Area of Concern (AOC) Remedial Action Committee (RAC) of the Trustees' intention to fund wetland and/or stream enhancement and restoration within the AOC portion of the Genesee River. The preferred Genesee River enhancement projects are noted below, however, in the event that wetland enhancement cannot be conducted at the two locations, other wetland and/or stream enhancement or restoration options will be evaluated within the AOC boundary of the Genesee River. If the alternative Genesee River enhancement or restoration projects are found to meet the regulatory and site-specific criteria outline in the forthcoming Draft Restoration Plan, the projects will be funded.

Genesee River Turning Basin Wetland Enhancement and Turning Point Park Wetland Enhancement

In 2014, at the request of the United States Environmental Protection Agency (USEPA) Great Lakes National Program Office, under the Great Lakes Restoration Initiative, the USFWS New York Field Office (NYFO) completed an assessment of Rochester Embayment AOC wetlands to assess trends in wetland size and condition, and rank wetland habitats for protection and restoration (Gefell et al. 2014, USFWS 2014a). As part of the assessment project, the NYFO ranked relative wetland quality among 15 waterbodies (seven lotic and eight lentic) using a total of 26 metrics representing features of structural and vegetative habitat, water quality, and animal communities. The NYFO identified the environmental features contributing most to wetland habitat impairment across the project area.

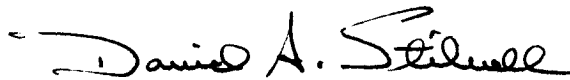
A Habitat Restoration Recommendation Project was a direct extension of the Rochester Embayment AOC wetland habitat assessment project (USFWS 2014b). This project utilized the output from the wetland assessment project and conducted further analyses of the assessment data to recommend solutions and prioritize areas most in need of restoration. Two of the ten

recommended habitat restoration projects for the AOC were within the Genesee River and were wetland enhancement projects, Turning Basin wetland enhancement and Turning Point Park wetland enhancement.

The two Genesee River wetland enhancement projects, Turning Basin wetland enhancement and Turning Point Park wetland enhancement, would involve excavation of channels and potholes within monotypic cattail marshes within the City of Rochester's Genesee River Turning Point Park, along the Genesee River. Enhancement actions may also include methods to restore natural habitat patchiness and topographic and vegetative complexity. Channels and potholes within emergent marshes would be sited to enhance areas that were historically open water. Enhancement of emergent marshes would occur in areas currently dominated by cattail, thereby reducing coverage of invasive species. Native herbaceous and mast-bearing shrubs would be planted to directly reduce coverage by invasive plants and encourage usage of wetlands by wildlife.

The Trustees are pleased to be working in conjunction with the Rochester Embayment AOC RAC, USEPA, and other Federal and non-Federal entities towards restoring habitat within and along the Genesee River. If you have any questions regarding this letter, please contact Amy Roe of my office at 607-753-9334.

Sincerely,



David A. Stilwell
Field Supervisor

References

Gefell, D., E. VanWyk, A. Secord, N. Vermeulen, E. Buckley, J. Ecret, A. Lowell, and A. Roe. 2014. Wetland Habitat Assessments at the Rochester Embayment Area of Concern on the South Shore of Lake Ontario. *Wetland Science and Practice*:31,4, 22-27.

USFWS. 2014a. Wetland Assessment in the Rochester Embayment Area of Concern in Support of the Loss of Fish and Wildlife Habitat BUI Removal Evaluation. Final Report. Submitted by: USFWS New York Field Office: Gefell, D., E. VanWyk, A. Secord, N. Vermeulen, E. Buckley, J. Ecret, A. Lowell, A. Roe, and C. Schwartz. Submitted to: USEPA Great Lakes National Program Office. Funded by: USEPA, Great Lakes Restoration Initiative. Available at: <http://www.fws.gov/northeast/nyfo/ec/glri.htm>.

USFWS 2014b. Final Status Report: Wetland Restoration Recommendations at the Rochester Embayment Area of Concern in Support of the Loss of Fish and Wildlife Habitat BUI Removal. Submitted by: USFWS New York Field Office: Gefell, D., E. VanWyk, G. Dodici, A. Secord, N. Vermeulen, C. Adams, and C. Schwartz. Submitted to: USEPA Great Lakes National Program Office. Funded by: USEPA, Great Lakes Restoration Initiative. Available at: <http://www.fws.gov/northeast/nyfo/ec/glri.htm>.

cc: Mark Barash, Office of the Solicitor, Newton, MA
Paul D'Amato, NYSDEC, Avon, NY
Patrick Foster and Sharon Brooks, Albany, NY

Letter sent to the Rochester AOC RAC and USEPA:

Wade Silkworth, Monroe County Department of Public Health, Rochester, NY
Fred Luckey, USEPA, New York, NY
Brenda Jones, USEPA, Chicago, IL

Wade Silkworth, P.E.
Senior Public Health Engineer
Monroe County Department of Public Health
111 Westfall Road, Room 938
Rochester, NY 14620
WadeSilkworth@monroecounty.gov

Frederick Luckey
Environmental Scientist
U.S. Environmental Protection Agency, Region 2
Clean Water Division
Watershed Management Branch
New York Watershed Management Section
290 Broadway, 24th Floor
New York, NY 10007-1866
luckey.frederick@epa.gov

Brenda R. Jones
U.S. Environmental Protection Agency
Great Lakes National Program Office
77 West Jackson Blvd. (G-17J)
Chicago, IL 60604
(312) 886-7188 phone
(312) 697-2069 fax
jones.brenda@epa.gov

Attachment 2



Department of Public Health

Monroe County, New York

Cheryl Dinolfo
County Executive

Jeremy T. Cushman, MD, MS, EMT-P, FACEP
Interim Health Commissioner

REMEDIAL ACTION COMMITTEE MEETING **ROCHESTER EMBAYMENT AREA OF CONCERN** **Meeting Minutes**

Present:

Jennifer Dunn, NYS DEC

Josh Haugh, NYS DEC

Wayne Howard, Solara Concepts

Brenda Jones, USEPA-GLNPO

Dorraine Kirkmire, City of Rochester

David Klein, The Nature Conservancy

Charlie Knauf; RAC Chair

Fred Luckey, USEPA-R2

Pete Rightmyer, MCDPH

Paul Sawyko, MC Stormwater Coalition

Wade Silkworth, MCDPH

June Summers, Gen. Valley Audubon Society

George Thomas, CEI

Charles Valeska, Self-proclaimed Master Gardener

Jeff Wyatt, URMC

Date: March 15, 2016

Time: 1:00 - 4:00 pm

Where: 111 Westfall Road, Rochester, NY 14620, ROOM 964

A. Brief Introductions: Josh Haugh introduced himself as the new Statewide AOC Coordinator, replacing Gerry Pratt at the DEC.

B. Genesee River Habitat Projects:

Discussion: Charlie discusses the RCRA activities on the Genesee River and how to satisfy the intended purpose of the habitat BUI's. Due to RCRA activities, the two habitat projects in the Genesee River will be put on hold until at least 2018, when it may be known how the RCRA cleanup will impact these project sites. This action, which is beyond the auspices of the AOC/RAP authority, impacts the original habitat management action and deadline commitments as described in NYSDEC's "Management Actions" letter to EPA's Great Lakes National Program Office. Choices are to: (1) modify the original 10 projects to 8, which will maintain the current timeline and *potentially* keep us in the "fast track" and allow us to obtain priority funding for monitoring; **or** (2) continue with the original 10 projects, which would delay the Rochester Embayment timeline for completion of management actions for at least another 2 years. General consensus with the group is to modify the management actions to 8 projects, with the understanding that the other two projects will eventually be completed. Funding for these two projects is secure and should not be affected by a new administration. Based on the recommendation of the RAC, a letter will be composed by the NYSDEC (Don Zelazny) proposing the modification to the habitat projects on the original management action list. The letter will discuss the modification to the management actions, justify the rationale for the change, and state that the RAC is confident that USFWS, based on their commitment letter, is dedicated to complete the two projects in the Genesee River.

Decision: Based on reasoning stated in the committee's discussion, The Remedial Action Committee recommends that the management actions related to the Loss of Fish and Wildlife Habitat BUI be reduced from the previously agreed upon 10 habitat restoration projects to 8 habitat restoration projects by removing the "Turning Basin" and "Turning Point Park" projects proposed in the Genesee River from the management action list.



Department of Public Health

Monroe County, New York

Cheryl Dinolfo
County Executive

Jeremy T. Cushman, MD, MS, EMT-P, FACEP
Interim Health Commissioner

D. Citizen Participation Plan

Wade has constructed a working document for public participation that is being updated continually. If any members of the RAC have events that they are part of and would like to report their outreach with the public, please let Wade know so he can update. Paul Sawyko discusses the purpose of the Water Education Collaborative and the need to coordinate and organize the outreach to eliminate redundancy. The WEC has resources that are available to the RAC to aid in outreach endeavors. Paul will reach out to Joan Kennedy (NYSDEC) and incorporate the history of the WEC into the plan. Brenda offers ideas such as Great Lakes Mud (Twitter, Facebook, etc....) to reach a broader group. Dorraine highlights that the Terminal Building at the Port of Rochester will be very active this summer and offers space to the Rochester AOC to engage the community.

F. Member reports/updates:

George Thomas: CEI is hosting their third annual Genesee River Basin Summit on May 25, 2016. Focus will be on sediment/erosion in the river. Dick Young and Joe Makarewicz will be the keynote speakers.

Jeff Wyatt: Seminar at CTSI at the University of Rochester in May in regards to sturgeon as a bioindicator of health. Dawn Dittman will be releasing sturgeon in the fall. Dates will be forthcoming.

Fred Luckey: Lake Ontario Lakewide Action Management Plan undertook a Great Lakes Science Initiative in conjunction with the NYSDEC and NYSDOT and have erected 20 signs along major highways (ie. thruway) in the state identifying entry into either the Lake Erie or Lake Ontario Basin.



DRAFT

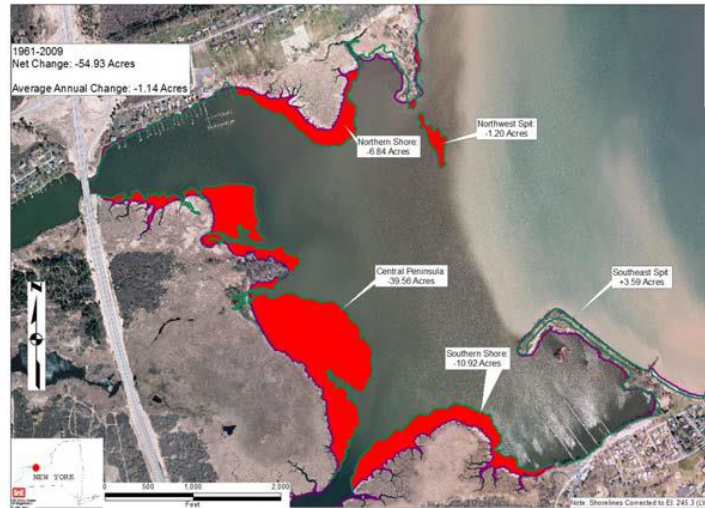


**US Army Corps
of Engineers®**

Braddock Bay Coastal Wetland Restoration, NY Economy Act

Project Location: Braddock Bay is located on the shore of Lake Ontario within the Great Lakes Rochester Embayment Area of Concern (AOC) in the town of Greece in Monroe County, NY.

Description of Problem: A historic barrier beach that was once present at Braddock Bay, no longer exists. This barrier beach was critical for maintaining the diverse ecological conditions within the bay. Over time, erosion of interior wetlands has occurred at a rate of 0.42 to 1.0 acre per year resulting in the loss of approximately 106 acres of wetland since 1902.



Proposed Project: The scope of this project includes the construction of a barrier beach: 1) a 1,700-foot long continuous rubblemound breakwater spine, as a core for the beach, with two 180-foot long rubblemound terminal groins attached; 2) a 3 -acre headland beach; and 3) two 150-foot long headland rubblemound breakwaters. The project will also include measures to restore diversity to the existing emergent marsh and create 4 acres of new emergent marsh. Restoration of the emergent marsh will be accomplished through invasive species treatment and excavation of channels and potholes to increase habitat diversity.



Partners and Collaboration: No cost share sponsor is required for the Design and Implementation of the project. Both the Town of Greece and New York State Department of Conservation support implementation.

Project Benefits: The project was identified by New York State as an essential management action for Rochester Embayment AOC delisting, particularly for delisting of the loss of fish and wildlife habitat Beneficial Use Impairment (BUI). The project benefits include: restoration of 185 acres of coastal wetlands protected and enhanced; protection of 0.6 miles of Lake Ontario shoreline, and contributing to the Lake Ontario Lakewide Management Plan (LaMP) goal to maintain/restore ecosystem to support self-reproducing diverse biological communities, and restoring habitat for the New York State endangered black tern that has not bred there since 1998.

Project Status: Phase 1 of the project, excavation of channels and potholes, was completed in March 2016. Phase 2, Construction of the barrier beach was completed in July 2018. Emergent wetland was completed September 2018. Adaptive management activities occurred in 2019, 2020, 2021 and are complete. Monitoring activities will continue through September 2022. A minor adaptive management design effort is underway for implementation in FY23 to clear channels that became closed following primary implantation. This effort is expected to cost \$200K and will be complete in April 2023.

Measure of Progress	Project Output
4.1.2 Miles of shoreline and riparian corridors protected and enhanced	0.3 miles
4.1.3 Acres coastal wetlands protected, restored and enhanced	340 acres



Estimated Project Costs	
Federal	\$10,700,000
Non-Federal	\$0
Total	\$10,700,000

Project Milestones	
Construction completion	Sep 2018
FY20 Monitoring & Adaptive Mgmt	Sep 2020
FY21 Monitoring & Adaptive Mgmt	Sep 2021
Adaptive Management of channels	April 2022

Point of Contact
Tim Noon; Project Manager 716-879-4261 Timothy.w.noon@usace.army.mil



DRAFT

USGS Location Identifier	Location Description	CHEMICAL_NAME	SAMPLE_DATE	Result	UNITS mg/l - Milligrams per liter	Qualifier J - Estimated value
04232007	Genesee River Charlotte	Total Suspended Solids	5/7/2013	68	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/4/2013	24.9	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	7/3/2013	22.7	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	8/6/2013	13.5	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	9/4/2013	15.8	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	10/2/2013	11.5	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	11/6/2013	22.7	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	12/3/2013	11	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	12/22/2013	482	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	1/16/2014	170	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	3/26/2014	83.6	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	4/24/2014	116	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	5/2/2014	203	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/2/2014	15.3	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/27/2014	21.7	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	7/7/2014	15.2	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	8/5/2014	151	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	9/9/2014	14	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	10/7/2014	11.9	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	11/6/2014	11.6	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	12/9/2014	166	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	1/13/2015	7.3	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	3/10/2015	3	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	4/9/2015	262	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	5/7/2015	9.3	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/2/2015	74.6	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/28/2015	35.5	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	7/15/2015	20.9	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	8/4/2015	7.3	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	9/15/2015	26.2	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	10/15/2015	21.9	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	11/4/2015	32.4	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	12/9/2015	12.2	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	1/6/2016	55.1	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	2/12/2016	10.2	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	3/17/2016	31.6	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	4/12/2016	15.9	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	5/10/2016	19.8	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/6/2016	13.4	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	7/7/2016	9.5	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	8/10/2016	7.3	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	9/7/2016	2.4	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	10/20/2016	16.1	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	10/21/2016	16.8	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	11/8/2016	103	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	12/6/2016	15.6	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	2/7/2017	25	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	3/8/2017	23	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	4/7/2017	172	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	5/15/2017	20.6	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	6/6/2017	12.1	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	7/20/2017	30.8	mg/l	J
04232007	Genesee River Charlotte	Total Suspended Solids	8/10/2017	11.9	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	9/19/2017	8.5	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	10/12/2017	13.1	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	11/14/2017	112	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	1/12/2018	57.7	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	2/28/2018	139	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	3/20/2018	13	mg/l	
04232007	Genesee River Charlotte	Total Suspended Solids	4/19/2018	211	mg/l	

USGS Location Identifier	Location Description	CHEMICAL_NAME	SAMPLE_DATE	Result	UNITS mg/l - Milligrams per liter	Qualifier J - Estimated value
04227500	Genesee River Mt Morris	Total Suspended Solids	5/6/2013	25.4	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/3/2013	56.2	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	7/2/2013	57.4	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	8/5/2013	15.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	9/3/2013	22.6	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	10/1/2013	8.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	11/5/2013	19.4	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	12/2/2013	27.2	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	12/22/2013	1480	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	1/15/2014	137	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	2/3/2014	20.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	3/26/2014	244	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	4/23/2014	83.9	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	5/1/2014	1460	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/2/2014	83.9	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/26/2014	767	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	7/7/2014	22.1	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	8/4/2014	2200	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	9/8/2014	13	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	10/6/2014	7.2	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	11/5/2014	20.2	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	12/8/2014	225	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	1/12/2015	18.5	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	3/9/2015	2.6	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	4/8/2015	283	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	5/6/2015	34	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/1/2015	374	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/28/2015	477	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	7/14/2015	37.7	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	8/3/2015	13.4	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	9/14/2015	55.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	10/14/2015	22.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	11/3/2015	115	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	12/8/2015	13.9	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	1/5/2016	58.3	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	2/11/2016	30.2	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	3/16/2016	88	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	4/11/2016	55.5	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	5/9/2016	27.9	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/6/2016	28.5	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	7/6/2016	19.3	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	8/9/2016	14.5	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	9/6/2016	18.6	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	10/19/2016	22.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	10/21/2016	98.8	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	11/8/2016	20	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	12/6/2016	13	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	1/12/2017	157	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	2/6/2017	34.9	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	3/8/2017	334	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	4/7/2017	2570	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	5/15/2017	37.4	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	6/6/2017	26.1	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	7/19/2017	149	mg/l	J
04227500	Genesee River Mt Morris	Total Suspended Solids	8/10/2017	36.9	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	9/19/2017	24.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	10/12/2017	246	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	11/14/2017	120	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	12/20/2017	237	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	1/12/2018	1010	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	2/28/2018	71.6	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	3/20/2018	26.8	mg/l	
04227500	Genesee River Mt Morris	Total Suspended Solids	4/20/2018	344	mg/l	

USGS Location Identifier	Location Description	CHEMICAL_NAME	SAMPLE_DATE	Result	UNITS mg/l - Milligrams per liter	Qualifier J - Estimated value
04223000	Genesee River Portageville	Total Suspended Solids	5/6/2013	5.1	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/3/2013	16.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	7/2/2013	27.9	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	8/5/2013	15.3	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	9/3/2013	16.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	10/1/2013	7.6	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	11/1/2013	22.9	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	11/5/2013	11.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	12/2/2013	6.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	12/22/2013	1450	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	1/15/2014	125	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	2/3/2014	4.3	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	3/26/2014	136	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	4/23/2014	9.9	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	5/1/2014	869	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/2/2014	6.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/26/2014	778	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	7/7/2014	7.1	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	8/4/2014	363	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	9/8/2014	5.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	10/6/2014	3.2	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	11/5/2014	4.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	12/8/2014	34	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	1/12/2015	15.5	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	3/9/2015	1.1	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	4/8/2015	107	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	5/6/2015	3.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/1/2015	322	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/28/2015	391	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	7/14/2015	6	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	8/3/2015	6.6	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	9/14/2015	27.9	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	10/14/2015	6.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	11/3/2015	9.9	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	12/8/2015	6.5	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	1/5/2016	16.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	2/11/2016	7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	3/16/2016	12.1	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	4/11/2016	3.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	5/9/2016	3	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/6/2016	8.3	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	7/6/2016	14.6	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	8/9/2016	10.6	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	9/6/2016	6.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	10/19/2016	13.9	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	10/21/2016	676	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	11/8/2016	9.3	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	12/6/2016	9.9	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	1/12/2017	48.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	2/6/2017	17.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	3/8/2017	200	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	4/7/2017	1320	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	5/15/2017	3.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	6/6/2017	7.4	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	7/19/2017	22.9	mg/l	J
04223000	Genesee River Portageville	Total Suspended Solids	8/10/2017	15.1	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	9/19/2017	11.3	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	10/12/2017	407	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	11/14/2017	15	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	12/20/2017	117	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	1/12/2018	3880	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	2/28/2018	43.8	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	3/20/2018	9.7	mg/l	
04223000	Genesee River Portageville	Total Suspended Solids	4/20/2018	55.2	mg/l	

USGS Location Identifier	Location Description	CHEMICAL_NAME	SAMPLE_DATE	Result	UNITS mg/l - Milligrams per liter	Qualifier J - Estimated value
04231600	Genesee River Rochester	Total Suspended Solids	5/7/2013	17.5	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/4/2013	91.4	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	7/3/2013	33.8	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	8/6/2013	27.2	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	9/4/2013	13.6	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	10/2/2013	14.2	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	11/6/2013	31.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	12/3/2013	8.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	12/22/2013	618	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	1/16/2014	37.9	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	3/26/2014	68.7	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	4/24/2014	138	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	5/2/2014	290	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/2/2014	16.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/27/2014	38.6	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	7/8/2014	17.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	8/5/2014	772	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	9/9/2014	12.3	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	10/7/2014	7.7	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	11/6/2014	10.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	12/9/2014	187	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	4/9/2015	326	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	5/7/2015	11.8	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/2/2015	297	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/28/2015	96.6	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	7/15/2015	24.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	8/4/2015	14.3	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	9/15/2015	13	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	10/15/2015	17.3	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	11/4/2015	24.8	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	12/9/2015	8	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	1/6/2016	60.5	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	2/12/2016	11.4	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	3/17/2016	38.6	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	4/12/2016	11.6	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	5/10/2016	17.8	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/6/2016	15.2	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	7/7/2016	36.2	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	8/10/2016	34.5	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	9/7/2016	10	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	10/20/2016	17.5	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	10/21/2016	24.7	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	11/8/2016	23.7	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	12/6/2016	13.2	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	1/12/2017	44.9	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	2/7/2017	12.8	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	3/8/2017	42	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	4/7/2017	416	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	5/15/2017	54.3	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	6/6/2017	33.7	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	7/20/2017	23.6	mg/l	J
04231600	Genesee River Rochester	Total Suspended Solids	8/10/2017	16.1	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	9/19/2017	29.2	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	10/12/2017	17.4	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	11/14/2017	169	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	1/12/2018	96.5	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	2/28/2018	137	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	3/20/2018	10.2	mg/l	
04231600	Genesee River Rochester	Total Suspended Solids	4/19/2018	226	mg/l	

DRAFT

**RAP Progress in the Rochester Embayment of Lake Ontario:
Population Monitoring, Trophic Relationships,
and Levels of Bioaccumulative Chemicals of Concern
in Mink, a Sentinel Species**

Final Report

to

The New York Great Lakes Protection Fund
NYS Dept. of Environmental Conservation
270 Michigan Avenue
Buffalo, NY 14203-2999

from

James M. Haynes and Sara T. Wellman
Department of Environmental Science and Biology
State University of New York at Brockport
Brockport, NY 14420-2973

and

James J. Pagano
Environmental Research Center
State University of New York at Oswego
Oswego, NY 13126

August 2007

EXECUTIVE SUMMARY

This project was designed to determine if two use impairments identified in the Remedial Action Plan (RAP) for the Rochester Embayment of Lake Ontario (RELO) can be delisted: 1) bird or animal deformities or reproductive problems, and 2) degradation of fish or wildlife populations. The eight research and one management questions addressed by this study, and the answers to those questions, follow.

1. *Are there differences in the relative abundance of lakeshore and inland mink populations in and out of the RELO AOC (Area of Concern)?* Video-trapping data tentatively suggested that there are no differences in the relative abundance of mink populations in and out of the AOC or between lakeshore and inland areas.
2. *Are mink reproducing in and out of the RELO AOC?* Video observations confirmed that mink are reproducing along the lakeshore in the AOC and at other locations studied. Mink less than one year old were physically trapped in all areas, implying reproduction in all areas.
3. *Can stable isotope analysis be used to evaluate mink diets, at lakeshore and inland areas in and out of the AOC, in terms of trophic levels and terrestrial and aquatic food sources?* Mink in the study areas fed on prey at an average trophic level of 2.5 (half way between first- and second-level predators). The percent aquatic diet could not be determined.
4. *Can stable isotopes be used to construct a food web/bioaccumulation model for mink in the RELO AOC to predict body burdens of BCCs (bioaccumulative chemicals of concern) in mink in relation to their diets?* Using trophic level determinations and estimated values from literature ranging from 50% to 90% aquatic diet, a food web bioaccumulation model, modified from Sample *et al.* (1996), was used to predict the exposure of mink in the AOC to BCCs based on a BCC's concentration in the water body supporting a mink's food web.
5. *What are the current levels of BCCs in lakeshore and inland populations of mink in and out of the RELO AOC?* Highly consistent patterns of BCC concentrations were observed across tissues and chemicals. The clear signal in the chemical data are that mink captured near Lake Ontario, and presumably eating organisms exposed to Lake Ontario water and its food web, have significantly higher BCC concentrations in their tissues than mink captured inland.
6. *Which BCCs, and at what levels, are known to cause adverse effects on populations or reproduction, or to cause deformities, in mink?* The answer to this question is chemical and

tissue specific. Jaw lesions associated with 40.2 ppb TEQ (toxic equivalents of 2,3,7,8-tetrachlorodibenzo-p-dioxin)/g liver appear to be the most sensitive bioindicator of the toxic effects of BCCs on mink.

7. *Are concentrations of BCCs in RELO AOC mink high enough to cause adverse effects?* The highest measured TEQ value for AOC lakeshore mink (and in the entire study) was 47.62 pg TEQ/g liver wet weight, which is slightly higher than the lowest LOAEL (40.2 pg TEQ/g liver) at which jaw lesions have been observed in 31-week mink kits.
8. *How do predicted levels of BCCs in mink tissues (based on concentrations in Lake Ontario water) compare with measured tissue residues in lakeshore mink specimens?* The bioaccumulation model (Sample *et al.* 1996) worked well for dioxin/furan TEQs and for total PCBs. In both cases, the predicted low and high values bounded measured values, except for the low estimate for PCBs which was very close to the lowest measured value in a lakeshore mink. The model did not predict tissue levels of mercury well.
9. *What is the most reliable and efficient way to monitor the health of RELO AOC mink populations in the future?* Mink jaw lesions have the lowest reported LOAEL in relation to mink reproduction and deformities, and these lesions are a simple, inexpensive bioindicator. Only the mink with the highest total PCB concentration and adipose TEQ and the third highest liver TEQ, vs. the mink with the next highest body burden of BCCs, had jaw lesions and it came from the lakeshore-AOC area.

Conclusion— Mink are reasonably abundant in the RELO AOC, and they are reproducing. It is unlikely that BCC sources in the AOC are now contributing to “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems.” Exposure to the Lake Ontario food web is associated with the highest levels of BCCs in mink. The bioaccumulation model used in this study should be used to predict concentrations of dioxins/furans and PCBs in mink as new data on the concentrations of these chemicals in Lake Ontario or Braddock Bay water become available. Once the model predicts concentrations below the LOAEL for jaw lesions, further mink monitoring should be done by sending teeth for aging and jaws for analysis of lesions. If age 1 mink, and older mink with no lesions, are found, confidence that mink exposed to the Lake Ontario food web are no longer at risk for population, reproductive or deformity problems will be high and delisting should proceed.

TABLE OF CONTENTS

EXECUTIVE SUMMARY	i
INTRODUCTION	1
The mink as a sentinel species	2
METHODS	3
RESULTS AND DISCUSSION	4
Are there differences in the relative abundance of lakeshore and inland mink populations in and out of the RELO AOC? (Appendix 1)	4
Are mink reproducing in and out of the RELO AOC? (Appendix 1).....	4
Can stable isotope analysis be used to evaluate mink diets, at lakeshore and inland areas in and out of the AOC, in terms of trophic levels ($\delta^{15}\text{N}$ ratio) and terrestrial and aquatic food sources ($\delta^{13}\text{C}$ ratio)? (Appendix 2)	5
Can stable isotopes be used to construct a food web/bioaccumulation model for mink in the RELO AOC to predict body burdens of BCCs in mink in relation to their diets? (Appendix 2).....	5
What are the current levels of BCCs in lakeshore and inland populations of mink in and out of the RELO AOC? (Appendix 3)	5
Which BCCs, and at what levels, are known to cause adverse effects on populations or reproduction, or to cause deformities, in mink? (Appendix 4).....	6
Are concentrations of BCCs in RELO AOC mink high enough to cause adverse effects? (Appendix 4)	7
How do predicted levels of BCCs in mink tissues (based on concentrations in Lake Ontario water) compare with measured tissue residues in lakeshore mink specimens? (Appendix 4).....	8
What is the most reliable and efficient way to monitor the health of RELO AOC mink populations in the future?	9
CONCLUSION	10
ACKNOWLEDGMENTS	11
LITERATURE CITED	11
FIGURES	14
Map 1. Rochester Embayment Area of Concern	14
Map 2. Sub-basins considered in the RELO RAP	15
APPENDICES	16
1. Video-trapping report	16
2. Stable isotopes and bioaccumulation model report	54
3. Bioaccumulative chemicals of concern in mink tissue report	87
4. Literature review and bioaccumulation modeling report.....	113
5. Mink jaw deformities in relation to total PCBs and TEQs in their tissues.....	140

RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, Trophic Relationships, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species

INTRODUCTION

In the 1980s the binational (Canada, U.S.) International Joint Commission (IJC) began the process of creating and implementing remedial action plans (RAPs) in 43 contaminated areas of concern (AOCs) throughout the Great Lakes Basin. The IJC established 14 “use impairments” that could cause a local area to be “listed” as an AOC, including “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems.” In 1988, Foley *et al.* reported that fish in Lake Ontario and the Genesee River had PCB concentrations within the range shown to cause reproductive failure in captive mink. This evidence, coupled with the perceived absence of mink within 2 miles of the lake, led to the inclusion of these two use impairments in the RAP (1993, 1997). This study (Haynes *et al.* 2002) was designed to determine if populations of mink on the shore of the Rochester Embayment of Lake Ontario (RELO) are negatively impacted by bioaccumulative chemicals of concern (BCCs) and, if so, whether the BCCs are originating in the Embayment watershed or elsewhere. The AOC includes the Embayment, a 35 square mile portion of Lake Ontario south of a line between Bogus Point in the town of Parma and Nine Mile Point in the town of Webster (both in Monroe County, New York); adjacent wetlands and bays; and the six mile reach of the Genesee River, from the Lower Falls to the mouth at Lake Ontario (Figure 1). The RAP also includes the sub-watersheds of Salmon Creek (western sub-basin), the Genesee River, and Irondequoit Creek (central sub-basin) (Figure 2).

The questions addressed by this study were:

1. Are there differences in the relative abundance of lakeshore and inland mink populations in and out of the RELO AOC?
2. Are mink reproducing in and out of the RELO AOC?
3. Can stable isotope analysis be used to evaluate mink diets, at lakeshore and inland areas in and out of the AOC, in terms of trophic levels and terrestrial and aquatic food sources?

4. Can stable isotopes be used to construct a food web/bioaccumulation model for mink in the RELO AOC to predict body burdens of BCCs in mink in relation to their diets?
5. What are the current levels of BCCs in lakeshore and inland populations of mink in and out of the RELO AOC?
6. Which BCCs, and at what levels, are known to cause adverse effects on populations or reproduction, or to cause deformities, in mink?
7. Are concentrations of BCCs in RELO AOC mink high enough to cause adverse effects?
8. How do predicted levels of BCCs in mink tissues (based on concentrations in Lake Ontario water) compare with measured tissue residues in lakeshore mink specimens?
9. What is the most reliable and efficient way to monitor the health of RELO AOC mink populations in the future?

The purpose of this final report is to summarize the key findings related to questions 1-8 above (Appendices 1-4), and to propose a plan for monitoring RAP progress by monitoring the health of RELO AOC mink populations in the future.

The mink as a sentinel species

The mink is commonly found along water edges and in wetlands wherever there is cover such as emergent vegetation, brush or forest. It is a predator and eats anything it can catch, including aquatic invertebrates, fish, frogs, birds, and small mammals (Illinois Natural History Survey 2001). Its position atop the aquatic food chain makes it highly susceptible to toxic pollutants in its environment due to the processes of bioaccumulation and biomagnification.

The sensitivity of mink to BCCs became evident in the 1960s when farmed mink exhibited reproductive problems and mortality after feeding on fish taken from the Great Lakes. In 1968, mink fed Lake Michigan coho salmon as 15% of their diet suffered 80% kit mortality (Aulerich and Ringer 1977). Feeding mink coho salmon from Lake Michigan had adverse effects almost identical to those of giving the mink 30 ppm PCBs (Aroclor mixture) in their feed (Aulerich et al. 1971). Both the PCBs and the Lake Michigan salmon caused reproductive problems such as reduced whelping and kit

survival and decreased kit weight. Other adverse effects were increased adult mortality and digestive and excretory system problems such as anorexia, bloody stools, gastric ulcers, and degeneration of liver and kidney.

Since mink are an economic resource, there was great interest in research into pollutants' effects on them, and by 1991 many lab studies had shown mink "particularly sensitive to toxic chemicals" (Wren 1991). These studies were done on laboratory animals that were otherwise healthy, well nourished, and living in a climate-controlled environment; wild populations likely are more sensitive due to stresses of hunger, weather, disease, or injury. By 1991 the accumulated evidence prompted this pronouncement from the editors of the Proceedings of the Expert Consultation Meeting on Mink and Otter: "The mink is the free-living mammal most sensitive to toxic substances such as PCBs and TCDD, and its diet provides an integrated exposure to contaminants in shoreline wetlands" (Addison *et al.* 1991). BCCs of concern in Rochester AOC (defined by G. N. Neuderfer, aquatic toxicologist, NYSDEC, Avon, NY) are PCBs, dioxins/furans, aldrin/dieldrin, chlordane, mirex/photomirex, DDT/metabolites, and methyl-mercury. Thus, the mink is an appropriate sentinel species for the RELO RAP.

METHODS

The methods for each phase of this study are described in Appendices 1 (*Are There Differences in the Relative Abundance of Lakeshore and Inland Mink In and Out of the Rochester Embayment of Lake Ontario Area of Concern*; Wellman and Haynes 2006a), 2 (*Age, Size, and Stable Isotope Data of Mink Populations, and a Predictive Model of Biaccumulation of Chemicals of Concern in the Rochester Embayment of Lake Ontario*; Wellman and Haynes 2006b), 3 (*Levels of Bioaccumulative Chemicals of Concern in Mink In and Out of the Rochester Embayment Area of Concern and On and Off the Shoreline of Lake Ontario*; Pagano and Haynes 2007), and 4 (*Bioaccumulative Chemicals of Concern in Mink: Adverse Effects Levels and Results of a Predictive Model for the Rochester Embayment of Lake Ontario*; Wellman and Haynes 2007).

It is important to clear up one matter arising from PI Haynes' confusion and subsequent miscommunication with NYS GLPF Administrator. The original proposal

(Haynes et al. 2002) stated that “Polychlorinated Biphenyls (99 zones, 132 congeners), Mirex/Photomirex, HCB, and DDE” would be analyzed by PI Pagano and that “Columbia Analytical Services, Inc. will conduct dioxin/furan (Houston, TX) and Methyl-mercury (Kelso, WA) analyses for this project.” Because there are 209 PCB congeners, many of which do not elute in the standard total PCB analysis (including the dioxin-like, co-planar PCB congeners), and because co-planar PCB analysis is also a separate, expensive activity from dioxin-furan analysis, it was never proposed to do analyses for co-planar PCBs for this project. Using well-established values from the literature (Appendix 4), TEQs from the dioxin-furan analyses were recalculated to include estimated contributions from co-planar PCBs. Thus, the predicted toxicity of BCCs in mink was fully accounted for in the study. Finally, polybrominated diethyl ethers (PDBEs) not proposed for analysis in the original proposal also were quantified in mink tissues.

RESULTS AND DISCUSSION

Are there differences in the relative abundance of lakeshore and inland mink populations in and out of the RELO AOC? (Appendix 1)

Video-trapping data tentatively suggest that there are no differences in the relative abundance of mink populations in and out of the AOC or between lakeshore and inland areas. However, the statistical power (probability of avoiding a Type II error) of the analyses was low due to high variability within and between sampling sites and small sample sizes. A Type II error is the conclusion that there is no significant difference between treatments (e.g., lakeshore vs. inland relative population size) when, in fact, a difference exists. To be confident about delisting a use impairment requires high confidence that the probability of a Type II error is low (e.g., lakeshore vs. inland and AOC: in vs. out comparisons). Video-trapping data alone do not provide a high level of confidence for delisting the “degradation of fish and wildlife populations” use impairment for mink.

Are mink reproducing in and out of the RELO AOC? (Appendix 1)

Video observations confirmed that mink are reproducing along the lakeshore in the AOC and at other locations studied. Mink less than one year old (based on counting annual age

rings in teeth) were physically trapped in all areas, implying reproduction in all areas. Although our observations cannot indicate whether reproduction is taking place at levels that would be considered “normal,” these data suggest that the “reproductive problems” part of the “bird or animal deformities or reproductive problems” use impairment can be delisted in the AOC.

Can stable isotope analysis be used to evaluate mink diets, at lakeshore and inland areas in and out of the AOC, in terms of trophic levels ($\delta^{15}\text{N}$ ratio) and terrestrial and aquatic food sources ($\delta^{13}\text{C}$ ratio)? (Appendix 2)

Analysis of $\delta^{15}\text{N}$ showed that mink in the study area feed on prey at an average trophic level of 2.5 (slightly higher along the lakeshore and in the AOC than elsewhere, with the highest level (2.8) along the lakeshore in the AOC), where trophic level 1 is plants, trophic level 2 is herbivores, and trophic level 3 is primary carnivores. The percent aquatic diet could not be determined for lack of $\delta^{13}\text{C}$ values for carbon sources (i.e., phytoplankton, submergent and emergent macrophytes) in the AOC wetlands.

Can stable isotopes be used to construct a food web/bioaccumulation model for mink in the RELO AOC to predict body burdens of BCCs in mink in relation to their diets? (Appendix 2)

Using trophic level determinations ($\delta^{15}\text{N}$) and estimated values from literature ranging from 50% to 90% aquatic diet, a food web bioaccumulation model, modified from Sample et al. (1996), was used to predict the exposure of mink in the AOC to BCCs based on a BCC's concentration in the water body (e.g., Lake Ontario) supporting the minks' food web.

What are the current levels of BCCs in lakeshore and inland populations of mink in and out of the RELO AOC? (Appendix 3)

Highly consistent patterns of BCC concentrations were observed across tissues and chemicals. Correlations among concentrations of the seven most notable chemicals analyzed were mostly high and significant in adipose and liver tissue. There were no significant differences in BCC concentrations in and out of the RELO AOC but BCC concentrations in mink captured near the Lake Ontario shore were significantly ($P < 0.05$) or suggestively ($0.05 < P < 0.1$) greater than concentrations in mink captured inland. The clear signal in the chemical data are that mink captured near Lake Ontario,

and presumably eating organisms exposed to Lake Ontario water and its food web, have significantly higher BCC concentrations in their tissues than mink captured inland.

Which BCCs, and at what levels, are known to cause adverse effects on populations or reproduction, or to cause deformities, in mink? (Appendix 4)

During the literature search for this project, the lowest observed adverse effect level (LOAEL) was for dioxin (CDD); 0.053 ppb in the diet of mink was associated with reduced kit survival at three weeks (Appendix 4—Appendix A-1). For PCB TEQ (toxic equivalents in relation to TCDD), the lowest LOAEL found was 26.9 ppb in liver associated with changes in retinol and retinyl ester concentrations in 27-week juvenile mink (Appendix 4—Appendix A-2). For mercury, the lowest LOAEL found was 1.06 ppm in liver associated with smaller litter sizes (Table 1). Because the relationship between retinol and retinyl esters and problems with mink populations in terms of reproduction and deformities is unknown, more-easily-detected jaw lesions associated with 40.2 ppb TEQ/g liver appear to be the most sensitive bioindicator of the toxic effects of BCCs on mink (Table 1).

Table 1. Selected endpoints and effects levels reported for mercury, PCBs, and TEQs in mink diets and tissues. (Values in italics were estimated using the average brain:liver ratios from Evans *et al.* 2000, Wobeser *et al.* 1976, and Wren *et al.* 1987a, b.) CDD = chlorinated dibenzo dioxins, CDF = chlorinated dibenzo furans, HCB = hexachlorobenzene.

Impairment	Endpoint	Toxin	Effect Level	Conc. (ppm or ug/g)		Reference
				Diet	Tissue	
Brain						
Population	Adult mortality	Hg	LC100	5 ppm	19.9 ppm	Aulerich et al. 1974
Reproduction	Whelping reduced	Hg in fish	LOAEL	0.5 ppm	<i>23.2 ug/g</i>	Dansereau et al. 1999
Reproduction	Litter size reduced	Hg in fish	LOAEL	0.22 ppm	<i>1.06 ppm</i>	Halbrook et al. 1997
Population	Hg intoxication	MeHg	LOAEL	1.1 ppm	8.2 ppm	Wobeser et al. 1976
Reproduction	Litter size reduced	MeHg	LOAEL	1.0 ug/g	2.0 ug/g	Wren et al. 1987a,b
Liver						
Reproduction	Kit survival 3 & 6 wks	PCBs	LOAEL	720 pg/g	2190 pg/g	Heaton et al. 1995, Tillit et al. 1996
		CDDs	LOAEL	60 pg/g	2626 pg/g	
		CDFs	LOAEL	13 pg/g	335 pg/g	
		TEQs	LOAEL	22.4 pg/g	208.3 pg/g	
Deformities	Jaw lesion in 31-wk kits	PCBs	LOAEL	0.96 ug/g	1.698 ug/g	Bursian et al. 2006a, b
Deformities	Jaw lesion in 27-wk kits	TEQs	LOAEL	9.2 pg/g	40.2 pg/g	Bursian et al. 2006c
		PCBs	LOAEL	1.1 ug/g	16 ug/g	
		TEQs	LOAEL	47 pg/g	75 pg/g	

Reproduction	Litter size	PCBs	LOAEL	1360 ppb	7250 ppb	Halbrook et al. 1999
Reproduction	P-1 Whelping reduced	PCBs	LOAEL	0.25 ppm	860 ng/g	Restum et al. 1998
	F-2 Kit mortality	PCBs	LOAEL	0.5 ppm	464 ng/g	
					Adipose	
Reproduction	Kit mortality	HCB	LOAEL	1 ppm	95 ppb	Rush et al. 1983

LOAELs have been determined for many organochlorine (OC) pesticides (Appendix 4—Appendix A-4) but according to Giesy *et al.* (1994) studies in the 1970s and 1980s determined that OC pesticides did not cause the effects seen in mink that ate Great Lakes fish. Because OC pesticide levels have decreased in the environment since then, they would be even less significant today, which probably accounts for the lack of recent studies regarding them.

Are concentrations of BCCs in RELO AOC mink high enough to cause adverse effects? (Appendix 4)

An estimate of the total environmental TEQ exposure, based on analysis of only dioxins and furans, would range from two to ten times the dioxin/furan TEQ measured in mink tissues (Appendix 4). The highest measured TEQ value for AOC lakeshore mink in our study was 47.62 pg TEQ/g liver wet weight (Appendix 3), which is slightly higher than the lowest LOAEL (40.2 pg TEQ/g liver) at which jaw lesions were seen in 31-week kits (Table 1). The lowest measured TEQ value in lakeshore mink (0.22 pg TEQ/g), even when multiplied by ten, is still an order of magnitude smaller than the LOAEL, indicating no risk (Table 2). However, the average (excluding high and low TEQ values for the analyzed mink) of 7.8 pg TEQ/g for lakeshore mink (Table 2), if multiplied by five, approaches the LOAEL for jaw lesions. This indicates that some lakeshore mink are at risk of developing jaw lesions known to lead to jaw deformities, osteolysis, and tooth loss (Render *et al.* 2001).

The highest measured TEQ for inland mink was 4.16 pg TEQ/g (Appendix 3). When multiplied by ten, the result is approximately equal to the 40.2 pg TEQ/g LOAEL for jaw lesions, indicating that the most exposed of the inland mink may be at low risk for developing jaw lesions. However, the lowest (0.00 pg TEQ/g) and average (0.25 pg TEQ/g) TEQ values for inland mink (excluding high and low TEQ values for the analyzed mink), even when multiplied by ten, indicate that the majority of inland mink are not at risk (Table 2).

Table 2. TEQ values (pg/g) for dioxins and furans from Lakeshore and Inland mink livers, showing high, low and average (excluding high and low) values for each category.

Location	Value	TEQ	TEQ*2	TEQ*10
Lakeshore	Low	0.22	0.44	2.2
	Average (8)	7.75	15.50	77.5
	High	47.62	95.24	476.2
Inland	Low	0.00	0.00	0.00
	Average (8)	0.25	0.50	2.50
	High	4.16	8.32	41.6

Total mercury concentrations in the brains of AOC mink averaged 0.281 ppm along the lakeshore and 0.158 ppm inland (Appendix 3—Table 7), levels 3-6 times lower than lowest LOAEL of 1.06 ppm reported to cause a reduction in litter size of mink. Therefore, it is unlikely that mercury is having an adverse effect on mink in the AOC.

How do predicted levels of BCCs in mink tissues (based on concentrations in Lake Ontario water) compare with measured tissue residues in lakeshore mink specimens? (Appendix 4)

The bioaccumulation model (Sample *et al.* 1996) worked well for dioxin/furan TEQs and for PCBs. In both cases, the predicted low and high values bounded measured values, except for the low estimate for PCBs which was very close to the lowest measured value in a lakeshore mink (Table 3). This was expected, as the AOC is neither the most polluted nor the cleanest portion of Lake Ontario (Luckey and Litton 2005; J. Vincent, pers. comm.).

The model did not predict tissue levels of mercury well; the measured values were up to three orders of magnitude higher than predicted values. The reason for this discrepancy is not known. One possibility is the fact that the model is based on the octanol-water partition coefficient, a concept which applies only to lipophilic compounds, which mercury is not. However, Sample *et al.* (1996) apparently intended the model to be used with mercury, as they provided BAF factors for it (as well as several other heavy metals). Another possibility is that the model predicts mercury concentrations in tissue based only on aquatic exposures; mink in our study might have had exposure to mercury through terrestrial sources unaccounted for by the model.

Further investigation and development of the model will be required if it is deemed necessary to predict mercury levels in mink of the Rochester Embayment.

Table 3. Predicted versus measured values for tissue residues of dioxin/furans (TEQs), methylmercury, and PCBs, based on water concentrations in Lake Ontario as reported by J. Vincent (2006, Environment Canada, pers. comm..) and Luckey and Litton (2005).

Value	Water Conc.		Tissue Level	
	pg/kg	BCC	Predicted ng/g	Measured ng/g
Low	0.00006	TEQs (liver)	0.0000552	0.00022
High	0.0024		0.0621	0.0213
Low	0.0	MeHg (brain)	0.0	90
High	18.0		4.70	1,550
Low	26.0	PCBs (liver)	19.2	13.6
High	915.0		160,000	5,870

What is the most reliable and efficient way to monitor the health of RELO AOC mink populations in the future?

Mink jaw lesions have the lowest reported LOAEL in relation to mink reproduction and deformities (Appendix 4), and these lesions are a simple, inexpensive (~\$40/sample) bioindicator of exposure to BCCs, particularly dioxins and furans. Jaws from 12 specimens collected in this study were sent for histological preparation (Kerrie Beckett, Woodlot Alternatives, Topsham, ME) and analysis (Steven Bursian, Dept. of Animal Science, Michigan State University) including the animals with the highest, typical and lowest BCC levels at the inland and lakeshore study areas in and out of the AOC. The only mink with jaw lesions (multiple squamous epithelial cysts or cell proliferations at multiple zones along the entire dental arcade, including bone lysis and cell atypia consistent with malignancy; Beckett et al. 2005, Bursian et al. 2006) was captured along the lakeshore in the AOC. It also had the highest total PCB concentration (by a factor of 2.46) and adipose TEQ (by a factor of 8.85) and the third highest liver TEQ (by a factor of 0.45) of the animals analyzed (Appendix 5). Therefore, it is still possible for a mink in the AOC to accumulate body burdens of BCCs that produce jaw lesions, the most sensitive indicator of exposure currently known. However, there is no evidence in the literature or from our observations of juvenile mink that current levels of

exposure along the lakeshore or inland, in or out of the AOC, are adversely affecting mink populations.

CONCLUSION

This study documented the presence of mink populations, and mink reproduction, in the RELO AOC. Except for a single lakeshore mink with the highest BCC concentrations in its tissues, analytical, modeling and literature review results all suggest that mink reproduction is unlikely to be impaired in the AOC. Therefore, it is unlikely that BCC sources in the AOC are now contributing to the “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems” use impairments identified in the RAP (1993, 1997). The results make clear that exposure to the Lake Ontario food web is associated with the highest levels of BCCs in mink in the AOC and elsewhere along the lakeshore.

The bioaccumulation model used in this study should be used to predict concentrations of dioxins/furans and PCBs in mink as new data on the concentrations of these chemicals in Lake Ontario or Braddock Bay water become available. We recommend that the USEPA or NYDEC sample the waters of Braddock Bay and lower Salmon Creek, the capture location of mink #17 with the highest concentrations of BCCs, during their future monitoring of Lake Ontario. Once the model predicts concentrations below the LOAEL for jaw lesions, further biological monitoring should be done by contracting with trappers to capture mink and sending their teeth for aging and their jaws for analysis of lesions. If age 1 mink, and older mink with no lesions, are found, confidence that mink exposed to the Lake Ontario food web are no longer at risk for population, reproductive or deformity problems will be high and delisting should proceed.

ACKNOWLEDGMENTS

Individuals who assisted the various phases of this project are acknowledged in Appendices 1-4. Support for the project was provided by the New York Great Lakes Protection Fund by grant C302399.

LITERATURE CITED

- Addison, E.M., G.A. Fox, and M.G. Gilbertson (eds.). 1991. Proceedings of the Expert Consultation Meeting on Mink and Otter. International Joint Commission, Windsor, ONT. ISBN 1-895085-32-2.
- Aulerich, R.J., and R.K. Ringer. 1977. Current status of PCB toxicity to mink, and effect on their reproduction. *Arch. Environ. Contamination Toxicol.* 6:279-292.
- Aulerich, R.J., R.K. Ringer, H.L. Seagran and W.G. Youatt. 1971. Effects of feeding coho salmon and other Great Lakes fish on mink reproduction. *J. Reproduction and Fertility* 19: 365-376.
- Beckett, K.J., S.D. Millsap, A.L. Blankenship, M.J. Zwiernik, J.P. Giesy, and S.J. Bursian. 2005. Squamous epithelial lesion of the mandibles and maxillae of wild mink (*Mustela vison*) naturally exposed to polychlorinated biphenyls. *Env. Tox. Chem.* 24: 674-677.
- Bursian, S.J., C. Sharma, R.J. Auerlich, B. Yamini, R.R. Mitchell, K.J. Beckett, C.E. Orazio, D. Moore, S. Svirsky, and D.E. Tillitt. 2006. Dietary exposure of mink (*Mustela vison*) to fish from the Housatonic River, Berkshire County, Massachusetts, USA: Effects on organ weights and histology and hepatic concentrations of polychlorinated biphenyls and 2,3,7,8-tetrachlorodibenzo-P-dioxin toxic equivalence. *Env. Tox. Chem.* 25: 1541-1550.
- Evans, R.D., E.M. Addison, J.Y. Villeneuve, K.S. MacDonald, and D.G. Joachim. 2000. Distribution of inorganic and methylmercury among tissues in Mink (*Mustela vison*) and Otter (*Lutra canadensis*). *Environmental Research Section A* 84: 133-139.
- Foley, R.E., S.J. Jackling, R.J. Sloan and M.K. Brown. 1988. Organochlorine and mercury residues in wild mink and otter: Comparison with fish. *Env. Tox. Chem.* 7: 363-374.
- Giesy, J.P., D.A. Verbrugge, R.A. Othout, W.W. Bowerman, M.A. Mora, P.D. Jones, J.L. Newsted, C. Vandervoort, S.N. Heaton, R.J. Aulerich, S.J. Bursian, J.P. Ludwig, G.A. Dawson, T.J. Kubiak, D.A. Best, and D.E. Tillitt. 1994. Contaminants in fishes from Great Lakes-influenced sections and above dams of three Michigan rivers. II: Implications for health of mink. *Archives of Environmental Contamination and Toxicology* 27: 213-223.

- Haynes, J.M., Pagano, J.J., and Wellman, S.T. 2002. New York State Great Lakes Protection Fund Application for State Assistance—RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species.
- Illinois Natural History Survey. 2001. Mink Species Account.
<http://www.inhs.uiuc.edu/dnr/fur/species/mink.html>
- Luckey, F., and S. Litten. 2005. Bioaccumulative Contaminants in Lake Ontario Surface Water, 1999. USEPA. New York, NY.
- Pagano, J.J., and J.M. Haynes. 2007. Levels of Bioaccumulative Chemicals of Concern in Mink In and Out of the Rochester Embayment Area of Concern and On and Off the Shoreline of Lake Ontario. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.
- RAP. 1993. Stage I Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- RAP. 1997. Stage II Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- Render, J.A., S.J. Bursian, D.S. Rosenstein, and R.J. Aulerich. 2001. Squamous epithelium proliferation in the jaws of mink fed diets containing 3,3',4,4',5-pentachlorobiphenyl (PCB 126) or 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *Veterinary and Human Toxicology* 43(1): 22-26
- Sample, B.E., D.M. Opresko, and G.W. Suter II. 1996. Toxicological Benchmarks for Wildlife: 1996 Revision. Oak Ridge National Laboratory. Oak Ridge, TN.
- Wellman, S.T., and Haynes, J.M. 2006a. Monitoring Mink Populations using Video Traps. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.
- Wellman, S.T., and Haynes, J.M. 2006b. Age, Size, and Stable Isotope Data of Mink Populations, and a Predictive Model of Bioaccumulation of Chemicals of Concern in the Rochester Embayment of Lake Ontario. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.
- Wellman, S.T., and Haynes, J.M. 2007. Bioaccumulative Chemicals of Concern in Mink: Adverse Effects Levels and Results of a Predictive Model for the Rochester Embayment of Lake Ontario. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.
- Wobeser, G., Nielsen, N.O., Schiefer, B., 1976a. Mercury and mink. II. Experimental methyl mercury intoxication. *Canadian Journal of Comparative Medicine* 40: 34–45.
- Wren, C.D., Hunter, D.B., Leatherland, J.F., Stokes, P.M., 1987a. The effects of polychlorinated biphenyls and methylmercury, singly and in combination, on mink. I. Uptake and toxic responses. *Archives of Environmental Contamination and Toxicology* 16: 441–447.

- Wren, C.D., Hunter, D.B., Leatherland, J.F., Stokes, P.M., 1987b. The effects of polychlorinated biphenyls and methylmercury, singly and in combination on mink. II. Reproduction and kit development. Archives of Environmental Contamination and Toxicology 16: 449–454.
- Wren, C.E. 1991. Cause-effect linkages between chemicals and populations of mink (*Mustela vison*) and otter (*Lutra canadensis*) in the Great Lakes basin. J. Tox. Env. Health 33:549-585.

FIGURES

Figure 1. Rochester Embayment of Lake Ontario Area of Concern (RELO AOC).

Figure 1: Rochester Embayment of Lake Ontario Area of Concern

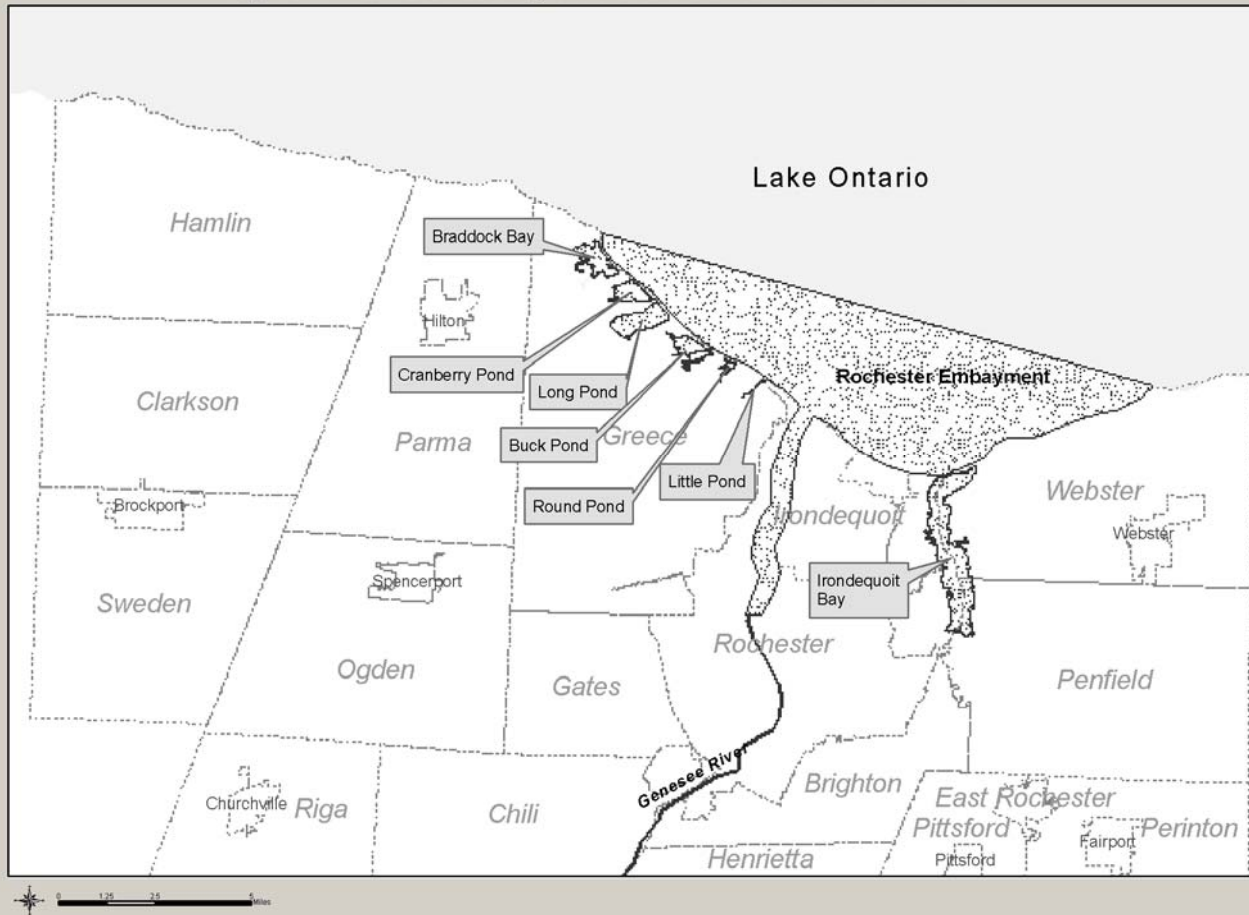
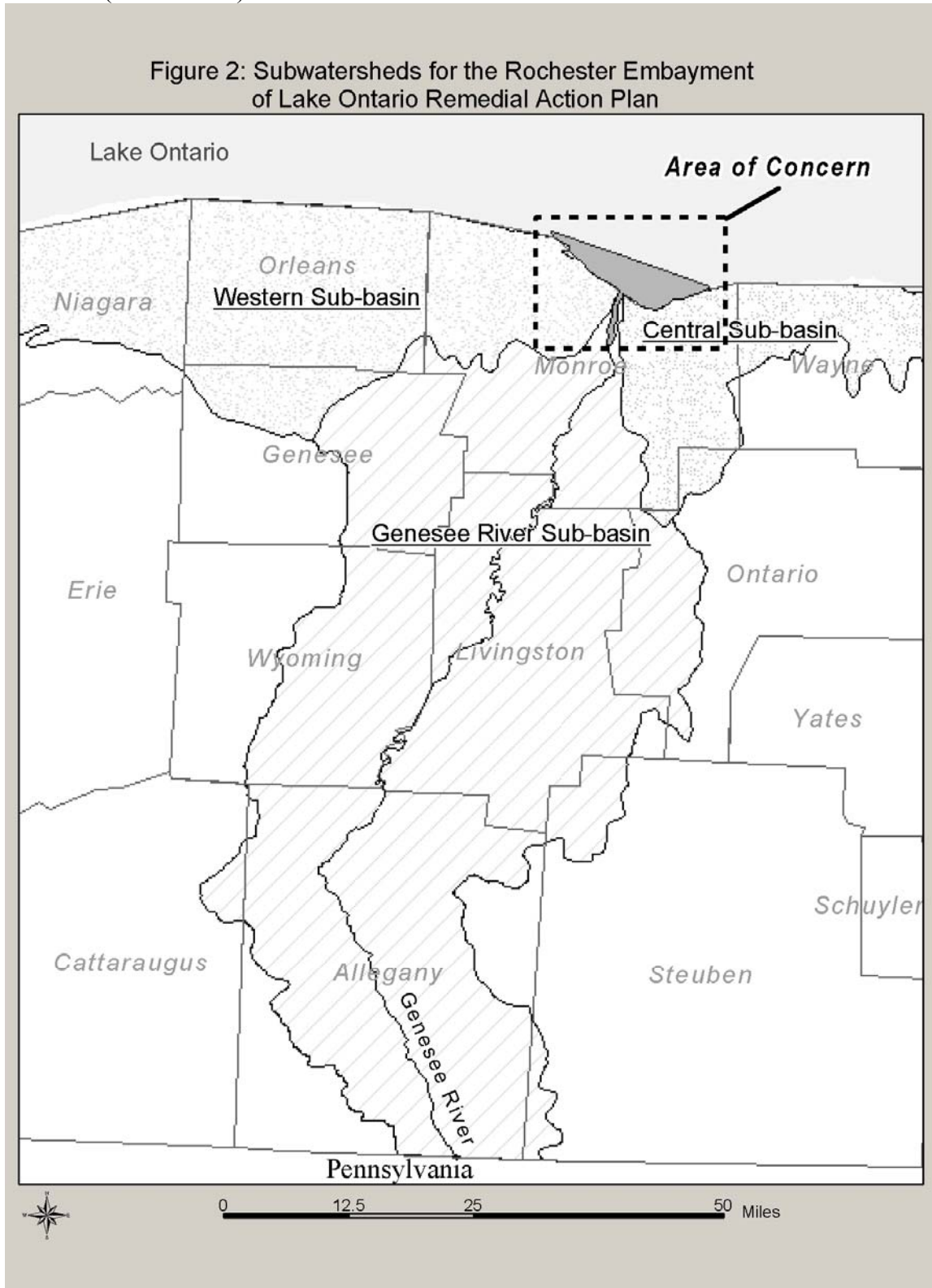


Figure 2. Sub-basins considered in the Rochester Embayment of Lake Ontario Remedial Action Plan (RELO RAP).



APPENDICES

Appendix 1

Are there Differences in the Relative Abundance of Lakeshore and Inland Mink Populations In and Out of the Rochester Embayment of Lake Ontario Area of Concern?: Monitoring Mink Populations Using Video Traps

Sara T. Wellman and James M. Haynes

Department of Environmental Science and Biology
State University of New York
College at Brockport
350 New Campus Drive
Brockport, NY 14420-2973

March 2006

OVERVIEW

This report is the first of four from project C302399, “*RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, Trophic Relationships, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species*,” funded by the New York Great Lakes Protection Fund in 2004. The project addresses use impairments related to water quality identified in the Remedial Action Plan for the Rochester Embayment of Lake Ontario (RELO RAP). This report deals with the development and use of video trapping systems that established the presence and reproduction of mink (*Mustela vison*) in and out of the RELO RAP Area of Concern (AOC). Three more reports will be written in 2006: (1) trophic positions (stable isotope analysis) and ages of mink (Wellman and Haynes, in preparation), (2) levels of bioaccumulative chemicals of concern (BCCs) in mink tissues (Pagano and Haynes, in preparation), and (3) a literature review of the effects BCCs on mink (Wellman, in preparation). Because the mink is the most sensitive species to BCCs known, the results of this study will determine if the fish and wildlife reproduction impairment for the RELO AOC can be recommended for delisting.

TABLE OF CONTENTS

INTRODUCTION	18
METHODS AND MATERIALS	18
Locations of Study Sites	18
MustelaVision System	19
System Requirements.....	19
System Components.....	19
System Placement	20
Field Service	20
Data Recording	20
Data Sheets	20
Definitions and Formulas.....	21
Data Analysis	22
RESULTS AND DISCUSSION	23
Relative Abundances	23
Relationship of Mink Passage Rates to Mink Abundances	23
Mink Passage Rates	24
Changes in Passage Rates from 2003 to 2004	25
Influence of Regional Descriptors	25
Influence of Site Descriptors	26
Ecological Observations	26
Mink Groups in the AOC.....	26
Observed Behaviors of Mink.....	27
CONCLUSION.....	29
ACKNOWLEDGMENTS	30
LITERATURE CITED	30
TABLES	
1. MustelaVision camera sites characterized by regional and site descriptors.....	33
2. Results of analysis of variance (GLM) of mink Passage Rates by Regional Descriptors	34
3. Results of analysis of variance (ANOVA) of mink Passage Rates by Site Descriptors	35
FIGURES	
1. Map showing placement of MustelaVision systems in four regions during 2003 and 2004.....	36
2. Mink Passage Rates calculated for each camera site during 2003 and 2004.....	37
3. Yearly changes (Delta Passage Rates) in mink passages for sites at which data were taken both years	38
APPENDICES	
A. MustelaVision Operations	39
B. Passage Rate Analyses	46
C. Regional Descriptor Analyses.....	46
D. Site Descriptor Analyses.....	50

INTRODUCTION

In the 1980s the binational (Canada, U.S.) International Joint Commission (IJC) began the process of creating and implementing remedial action plans (RAPs) in 43 contaminated areas of concern (AOCs) throughout the Great Lakes Basin. The IJC established 14 “use impairments” that could cause a local area to be “listed” as an AOC, including “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems.” In the Rochester AOC, both uses were defined as impaired because “very few” mink were then being trapped or observed within 2 miles of the lake (RAP 1993, 1997). This study was part of a project (Haynes *et al.* 2002) to determine whether lakeshore populations of mink along the Rochester Embayment of Lake Ontario (RELO) are negatively impacted by bioaccumulative chemicals of concern (BCCs) and, if so, whether the BCCs are originating in the embayment watershed or elsewhere.

The RELO AOC includes the Embayment, a 35 square mile portion of Lake Ontario south of a line between Bogus Point in the town of Parma and Nine Mile Point in the town of Webster (both in Monroe County, New York); adjacent wetlands and bays; and the six mile reach of the Genesee River from the Lower Falls to the mouth at Lake Ontario. The RAP also includes the subwatersheds of Salmon Creek, the Genesee River and Irondequoit Creek (RAP 1993, 1997; Figure 1).

The question addressed by this portion of the study was: Are there differences in the relative abundance of lakeshore and inland mink populations in and out of the RELO AOC? Our approach was to record the passage of mink using four “MustelaVision” videotrap systems in each of four regions designated as Lakeshore/AOC, Inland/AOC, Lakeshore/Out of AOC, and Inland/Out of AOC (Figure 1). Previous studies (cf. Gerrell 1970, Birks and Linn 1982, Eagle and Whitman 1987, Yamaguchi and MacDonald 2003) support our assumption that the number of mink passages recorded is related to relative population abundance (see Discussion). We tested the null hypothesis that there were no differences in passage rates among regions. We also went beyond the scope of our contract to look at the effect of several physical environmental variables on mink passage rates, and report on some observed mink behaviors.

METHODS AND MATERIALS

Locations of Study Sites

To maximize the chances of recording mink passages, we placed the Mustela-Vision systems in locations where mink were most likely to be found. Mink are semi-aquatic animals, found in wetlands and along water edges, especially with cover such as emergent vegetation, brush or forest (Linscombe *et al.* 1982, Allen 1986, Eagle and Whitman 1987, Dunstone 1993, Yamaguchi *et al.* 2003, Illinois Natural History Survey 2005, USDA Forest Service 2005).

Based on this information, the region chosen as Lakeshore/AOC was the Braddock Bay State Wildlife Management Area (BBWMA), a wetlands complex broadly connected to the RELO, separated only by narrow barrier beaches. The Inland/AOC region was around the Bergen Swamp, a smaller wetlands complex on Black Creek

(BLKCK), within the RELO watershed. The Inland/Out of AOC region was the Iroquois National Wildlife Refuge (INWR) and two connected state WMAs, a huge managed wetlands complex known to harbor abundant mink. Finally, the Lakeshore/Out of AOC region was along the Lake Ontario State Parkway (LOSPW) west of the RELO watershed, where creeks and small wetlands drain directly into Lake Ontario (Figure 1).

With the help of experienced trappers, we chose sites in each region most likely to be frequented by mink; in many cases we were guided by mink tracks or previous trappers' success. Each camera was placed near a water edge, either in a wetland or along a stream. Mink run along the edge of the water whenever possible (Burgess 1978, cited by Allen 1986; Eagle and Whitman 1987; Dunstone 1993; Yamaguchi *et al.* 2003; Yamaguchi and Macdonald 2003). Therefore, trappers usually set their traps at the corners of culverts and bridges where the minks' paths are funneled into these openings (Jamison 1983, Krause 1984, Geary 1985, National Trappers' Association 2005), and we looked for such tunnels when placing MustelaVision systems. Mink often use paths or "runways" through tall grass, cattails, and brush (Schladweiler and Storm 1969, Dunstone 1993, Racey and Euler 2003), and, according to trappers, ledges just under the water's surface along the bank, which also informed our choices of sites.

Because we were still searching for a suitable Lakeshore/Out of AOC region, no MustelaVision systems were placed in the LOSPW region in 2003. Systems were placed at 14 sites in the other three regions from June through October 2003. We considered the 2003 season our "exploratory season" during which we tried to find the best sites (i.e., most likely to observe mink passages) in each region, so either during 2003 or before the 2004 season, two systems in each of the original three regions were moved to potentially better locations. Thus, eight sites found in 2003, along with two new sites each in BBWMA and BLKCK and four new sites in the LOSP West region, were monitored from May to October 2004 (Table 1).

MustelaVision System

System Requirements

The MustelaVision system (Appendix A) was designed and built for this project by Jeffrey Wellman, an electrical engineer. We required a system usable in remote locations, powered by DC batteries, weatherproof, portable, lockable and affordable. To save battery power, videotape, and time for tabulating data, the system had to be triggered by the animal, rather than recording continuously. It also had to work day and night, because mink are considered predominantly nocturnal (cf. Birks and Linn 1982, Allen 1986, Eagle and Whitman 1987), and to operate quietly and invisibly to avoid disturbing the wary animals (Jamison 1983, Krause 1984, Barker 1991, Dunstone 1993).

System Components

The MustelaVision system (Appendix A-1a, b) consisted of an electronic camera head (Appendix A-1a, c) designed for security applications (Model PIC-I, SpyCameras ForLess.com), a 12-volt, 2-head videocassette recorder (Jensen KVC1500), a 12-volt DC deep-discharge battery, and a custom-built circuit board (Appendix A-1d) to protect the batteries from over-discharge which would shorten their useful lives. The camera head was attached to the VCR by a 50-foot cable, and it communicated with the VCR with an IR LED (infrared light emitting diode) that emulated the VCR's remote control unit.

The black and white Sony CCD (charge-coupled device) camera had a 464 X 625 pixel array, with 8-bit resolution, and a 92-degree field of view. The camera monitored an area 3 m wide by at least 12 m deep (depending on the camera angle relative to the ground). For image capture in the dark the camera head had six IR LEDs, providing a pool of illumination on the ground about 1 m wide by 2 m deep (again depending on camera angle. However, animals could be detected up to at least 10 m from the camera at night due to eye shine and their body heat against a cooler background.

The camera head had an IR motion detector that monitored a 104-degree by 15 m field. (Motion detection outside the viewing area meant fewer missed targets but more false triggers.) When the sensor detected motion it issued a “start recording” command to the VCR and started a 30-sec timer. If further motion was detected during the 30-sec period, the timer reset and recording time was extended. If no more motion was detected, after 30 seconds the camera head issued a “stop recording” command to the VCR.

System Placement

With one exception (a potential den site about 2 m from the water on Bald Eagle Creek’s west bank, LOSPW), each camera was placed on a stake within a meter of the water’s edge; often the stake was placed in the water. Each camera was aimed at the water’s edge to include in its field of view the pathway along which a mink would travel and the edge of the water in and along which it would forage. If the site included a tunnel, we aimed the camera at the opening through which mink would be forced to travel.

Along with mink habitat preferences, certain characteristics of the MustelaVision system dictated site choice and camera placement for optimal performance. Sites had to be near a road because of heavy batteries. To minimize tampering, the system had to be hidden in brush near a tree or other structure to which we could lock it. Also, we looked for high ground to avoid flooding and for shade to avoid overheating the electronics.

Camera angle was also important to avoid spurious triggers. We had to avoid sunbeams directed into the camera or reflected off water, vegetation near the camera that would trigger it during a breeze, and road and pedestrian traffic.

Field Service

Each MustelaVision system was serviced once per week. The batteries and videotapes were replaced, the camera lens was cleaned, the system was checked for functionality, and the field of view was checked for mink tracks and scats.

Data Recording

Data Sheets

System Log: Each MustelaVision system was assigned a letter to identify it, and had a separate log sheet (Appendix A-2a). During each field service session, we recorded on each System Log sheet the date of service, the ID numbers of the videotape cassette and battery, and comments such as mink tracks observed or operational problems.

Tape Log: Before viewing each videocassette, we recorded on its Tape Log (Appendix A-2b) which system the tape had been in, system location, dates during which the tape was in the system, and the total length of videotape recorded during that time. As we viewed the tape, we recorded periods of daylight and darkness, and all animals coming into the field of view of the camera.

Definitions and Formulas

Session: A Video Session was defined as the video recorded at one site on one cassette between service dates noted in System Log.

Mink Passage: A Mink Passage was defined as any time a mink came into the field of view of the camera and the camera was triggered and then turned off 30 sec after the mink left. Thus, if the mink passed out of the field of view and then back in before the camera stopped recording (no matter how many times), that was recorded as only one passage. In the rare cases in which multiple mink were recorded, their number was noted for that passage.

Trap Nights: Trap Nights were the number of 24-h periods observed on a video tape. In the video data log, a Trap Night was recorded for each period of contiguous dark shots, separated by daylight shots. If the camera was not triggered during a day, two consecutive nights could have been counted as one Trap Night. Thus, estimates were bounded by using Minimum and Maximum Trap Night calculations, in which the Min Trap Nights was the number of Trap Nights seen on the video during that session, and the Max Trap Nights was the number of nights between service dates in the System Log. If no nights were seen, but there were day shots, the Min Trap Nights was recorded as one. If the system triggered properly when started (recording our initial test hand wave, indicating that it was functioning properly), but otherwise did not trigger during the week (Session), the Trap Nights and Passages were recorded as zeros. If the system was non-functional throughout the Session (i.e. nothing on videotape at all, not even our hand wave), the Trap Nights and Passages were left blank and excluded from calculations.

Day vs. Night: A mink Passage was recorded as occurring during “Day” at any time that the natural illumination was sufficient to see outside the field of illumination of the camera head’s IR LEDs. When only objects illuminated by the camera’s LEDs could be seen, Passages were defined as “Night”. Light levels could not be used to define twilight because the camera had automatic brightness compensation, so that apparent light levels did not correspond to true ambient light levels.

Regional Descriptors: For the sake of easy reference during the following analyses, we define “Regional Descriptors” as those describing the four separate regions in which we worked, based on their characteristics on a landscape scale of many square kilometers. These Regional Descriptors were Inside vs. Outside the AOC (AOC In vs. Out); Inland vs. Lakeshore; and Landscape: Wetlands (large wetlands complex) vs. Mixed (habitat including uplands, streams, and small wetlands) (Table).

Site Descriptors: In order to evaluate the influence of ecological factors at each site on mink Passage Indices, we classified each camera site according to four factors. Habitat was classified as wetland, upland, or mixed, based on the types of vegetation visible around each camera site (many square meters). Cover was classified as cattail, brush, or forest, based on the vegetation at the camera’s specific location and in its field of view (a few square meters). We also recorded whether the water ran through a Tunnel (i.e., culvert or bridge) and whether each site had an underwater Ledge (Table).

Data Analysis

System Log sheets, Tape Log sheets, data keeping, and non-statistical calculations were done using Microsoft ® Excel 2000 by Microsoft Corporation.

Statistical analysis was done using Minitab™ Statistical Software Release 14.13 (Minitab Inc. 2005).

Data from each year at each site were originally kept separately because the habitat quality of each site could have varied from year to year due to weather or wetland management practices. We also kept the yearly data separate because we wanted to determine the potential impact on MustelaVision results of trapping we had contracted for in the AOC to collect tissue samples for other parts of the project.

Passage Rates (PRs) were calculated by dividing the number of mink Passages by the number of Trap Nights at a site. Since the Trap Nights had minimum and maximum values, corresponding maximum (Max) and minimum (Min) PRs were calculated for each site in each year. To determine whether the PRs changed from 2003 to 2004, we calculated “Delta PRs” at each site for which we had data from both years. To estimate the maximum possible change in passage rate (Max Delta PR) for each site, we subtracted that site’s 2003 Min PR from its 2004 Max PR; to estimate the minimum possible change in passage rate (Min Delta PR) we subtracted the 2003 Max PR from the 2004 Min PR. Thus, a positive Delta PR would indicate an increase in PR from 2003 to 2004. Using the Max Delta PRs from all sites as one data set, and the Min Delta PRs as a separate data set, we used one-sample T-tests to see if the mean difference (e.g., for either Max Delta PR or Min Delta PR) between years was different from zero. Means other than zero would have indicated a change in PRs from year to year. Since no differences were found, for subsequent analyses we combined the Passage Rate data from both years into two data sets (Max and Min PR) in which values for each year at each site were used as separate observations.

The INWR (Out of AOC study area) has a long history of targeted mink trapping, whereas the AOC does not. Therefore, we were concerned that when we contracted for mink trapping in the AOC (BBWMA and BLKCK regions) we would deplete a population already thought to be small. To assess the impact of our trapping in the AOC, we used two-sample T-tests to compare Min and Max PRs between the AOC and the INWR (we had no Delta PR data for LOSP West since no work was done there in 2003).

To evaluate the effects of the Regional Descriptors (AOC: In vs. Out, and Inland vs. Lakeshore) on the PRs, we used Minitab’s General Linear Model (GLM, a 2-way ANOVA with unbalanced cells, Tukey pairwise comparisons). We did an analysis for Max and Min PR. In a preliminary analysis, the site at Route 77 in the INWR in 2004 (Inland/Out of AOC) was an outlier with a standard residual greater than 4.7 (Figure 2). Therefore, we eliminated Min and Max PRs for that site in 2004. Also, because in the preliminary analysis the interaction between the Regional Descriptors AOC: In vs. Out, and Inland vs. Lakeshore was stronger than the effect of the descriptors themselves, we included a third Regional Descriptor, Landscape: Wetlands vs. Mixed Habitat, which exactly accounted for (i.e., the P-values were identical) the interactions in the earlier test. We estimated the power of the GLM using Minitab’s 2-Level Factorial power calculator (Factors = 3, Corners = 4, Replicates = 4, Effects = the differences between the means for each Regional Descriptor). Although this calculator was not designed for use with unbalanced cells, using the minimum number of replicates present in any of our regions yielded a conservatively low estimate of the actual power (Minitab support staff 2006, personal communication).

To evaluate the influences of the Site Descriptors on the variability of the PRs within each study Region, we did a one-way ANOVA for each Site Descriptor (Habitat: wetland, upland, mixed; Cover: cattail, brush, forest; Ledge: present, absent; Tunnel: present, absent). We also compared mink Passages during Day and Night, summed over all sites and both years, using a Chi-square test to determine whether there was any significant difference between the numbers of Passages during day and night.

RESULTS AND DISCUSSION

Relative Abundances

Relationship of Mink Passage Rates to Mink Abundances

Mink population density varies with habitat type; prey density, distribution and reliability; den availability; intraspecific aggression; and predation (Birks and Linn 1982, Linscombe *et al.* 1982, Allen 1986, Eagle and Whitman 1987, Dunstone 1993, Halliwell and MacDonald 1996, Sidorovich and Macdonald 2001, Yamaguchi *et al.* 2003). The sizes of minks' home ranges likewise vary with habitat quality, especially food supply, population structure and social stability of a population (Mitchell 1961, Allen 1986, Eagle and Whitman 1987). Thus, with reliably abundant food, mink populations are more dense and home ranges smaller, although in heavily trapped areas, the remaining males may have larger home ranges (Birks and Linn 1982, Eagle and Whitman 1987).

A number of studies show that mink are not strictly territorial and that their home ranges often overlap. Eagle and Whitman (1987) reported intrasexual territoriality in which home ranges of individuals of the same sex did not overlap, but females had home ranges inside males' territories. In contrast, Mitchell (1961) observed that adult male home ranges overlapped with juvenile males (even during the breeding season) but not with adult males. Gerrell (1970) reported that the home ranges of two of the four adult males he radio-tracked were visited by other adult males and females. The other two adult males visited other minks' home ranges, with one of them using the same den and core area used by an adult female two months earlier. He found that adult male home ranges often included home ranges of females with zones of overlap and zones monopolized by each mink. Occasionally both mink were present simultaneously in the zone of overlap, but usually intrusions occurred without direct confrontations when the owner was not present at that end of the home range. Linscombe *et al.* (1982) observed no active defense of any part of minks' home ranges from other mink of the same sex. Yamaguchi and MacDonald's (2003) radio-tracking study showed home range overlap ranging from 33% for female overlapping male to 88% for male overlapping male.

Mink use multiple dens within their home ranges, and move frequently between them as they cover their home range. Birks and Linn (1982) reported that the number of dens correlated with linear distance of home range, and that most den stays lasted less than one day. Allen (1986) reported stays ranging from a single night to a maximum of 40 days, with average distance between dens ranging from less than 90 m to 234 m in the U.S. In Sweden, Gerrell (1970) observed that mink usually used the nearest available den, with an average distance between dens used on consecutive days of 544 m. A juvenile female radio-tagged in east-central Minnesota by Schladweiler and Storm (1969) used 20 different dens prior to and during a 29-day study; only once was a den used for two

consecutive days. The straight-line distance between her daily dens varied from 99 to 849 m, averaging 353 m. Stevens *et al.* (1997) radio-tracked three males on large streams in eastern Tennessee, and found that the number of dens within home ranges varied from 8 to 24, and overnight movements of up to 4300 m were recorded.

Gerell (1970) reported that mink movements showed oscillations on two scales; small-scale movements, usually within <300m, were repeated in different parts of the home range until the entire home range was covered. All mink radio-tracked by Birks and Linn (1982) twice a day revealed more than 80% of their total home range within 5 days, and their entire home range within 10 days. Furthermore, Halliwell and MacDonald's (1996) statement that "most" territories were established by November implies that territories and home ranges may have been relatively fluid during the months of our study (May through October) when transients and dispersing juveniles would have been moving about.

Given the above, if mink are abundant enough, their home ranges or territories should abut one another, if not overlap. As population densities increase, home ranges become smaller, overlaps become greater, or both. Thus, because of the peripatetic nature of mink, higher population densities should result in higher rates of passage, whether from more mink passing through the same site, the same mink passing more often, or both. Thus, we are confident that our Passage Rate indices reflect relative mink numbers.

Mink Passage Rates

Mink Passages per Trap Night (Figure 2) varied from zero to 2.21 (INWR Route 77, 2004; Inland/Out of AOC) among sites and years, with a grand mean between 0.116 (Min PRs) and 0.216 (Max PRs). PRs were zero at nine of the 30 sampling sites; a Chi-square test showed no significant difference in the proportions of zero PRs between regions (Chi-Sq = 0.883, DF = 3, P = 0.830). The Passage Rates appeared to be higher and more variable in the AOC/Lakeshore (BBWMA) and the Out of AOC/Inland (INWR) regions, both of which are large wetlands complexes managed for the benefit of wildlife.

We suspect that mink had denned very close to the MustelaVision camera located on Route 77 in the INWR in 2004 (the statistical outlier), resulting in the unusually high PRs at this site (Figure 2). Several times in mid-August we recorded multiple mink traveling together there. This led us to suspect the presence of a den, since mink are normally solitary except during mating season (January through March, when we were not recording) and when mothers are raising their young from May through late summer (Mitchell 1961, Gerell 1970, Linscombe *et al.* 1982, Dunstone 1993, Illinois Natural History Survey 2005).

We tried to establish AOC Inland sites in the large Bergen Swamp complex, but there are no roads through it, so we had to choose sites on private lands outside the swamp. The Lakeshore Marshes Wildlife Management Area east of Sodus Bay would have been a close match in many ways to the BBWMA, but again there were no roads through the wetlands, so we had to resort to the LOSP West sites. (Roads were important primarily because they provide bridges or culverts at which to place cameras, and secondarily for convenience of servicing MustelaVision systems.) Thus, the AOC: In/Inland and the AOC: Out/Lakeshore regions consisted of more upland or mixed habitat than the large wetlands complexes of the AOC: In/Lakeshore and the AOC: Out/Inland regions, a circumstance that complicated our analyses (see below).

Changes in Passage Rates from 2003 to 2004

Changes in Passage Rates (Delta PRs) at sites for which we had two years' of data are shown in Figure 3. In 2003 and 2004, Delta PRs in the AOC (Lakeshore and Inland combined) were close to zero but they varied somewhat from zero in the Inland-Out of AOC area. However, the means of the Min and Max Delta PRs did not differ from zero (Min Delta PR: $P = 0.428$, Max Delta PR: $P = 0.511$; Appendix B: Table B-1), indicating that overall Passage Rates did not change between 2003 and 2004. This lack of change in PRs from year to year allowed us to combine both years' data for subsequent analyses.

There was no difference between the AOC and the Out of AOC regions in either Min Delta PR ($P = 0.554$) or Max Delta PR ($P = 0.938$) (Appendix B: Table B-2). The lack of differences among the Delta PRs for the BBWMA and the INWR between 2003 and 2004 was reassuring because we had contracted for mink trapping in the AOC (where mink had not been targeted previously) to get tissue samples required for other portions of the project. We were apprehensive about trapping the populations of concern, but the results showed no negative effect on the AOC populations, either Inland or Lakeshore.

The larger variation in Delta PRs in the INWR (Inland-Out of AOC) as compared to the BBWMA (Lakeshore-In AOC) may have been due to the fact that the INWR is a managed wetlands complex in which different areas were flooded in 2004 than in 2003. This could have changed the habitat quality in the areas around the MustelaVision sites, resulting in larger or smaller numbers of mink near each site. In contrast, the water levels in the BBWMA wetlands are naturally controlled by their connections to Lake Ontario, the level of which is tightly regulated; thus, habitat quality at MustelaVision sites there should have been more consistent from 2003 to 2004.

Influence of Regional Descriptors: Inland vs. Lakeshore, AOC: In vs. Out, and Landscape: Wetlands vs. Mixed

In the General Linear Model crossing the three Regional Descriptors, AOC: In vs. Out; Inland vs. Lakeshore; and Landscape: Wetlands vs. Mixed (Table 2), the descriptive statistics (Appendix C-1: Tables C-1a-c) and the main effects plots of the GLM (Appendix C-2: Figures C-1a-b) suggest that Passage Rates were lower inside the AOC than out of it, lower along the lakeshore than inland, and higher in the wetlands complexes than in mixed landscapes. However, the GLM itself (Appendix C-3: Table C-3) showed that AOC: In vs. Out had no significant effect (Max PR: $P = 0.404$; Min PR: $P = 0.446$), nor did Inland vs. Lakeshore (Max PR: $P = 0.251$; Min PR: $P = 0.342$). In contrast, the P-values for Landscape: Wetlands vs. Mixed were significant, at 0.026 and 0.042 for the Max and Min PRs, respectively.

The strong effect of Landscape (Table 2) was no surprise, as wetlands are known to be preferred habitat for mink (Allen 1986, Eagle and Whitman 1987, Dunstone 1993, Sidorovich and Macdonald 2001). The significance of the results for Landscape also make power calculations irrelevant (Chittenden 2002). The ability of the GLM to factor out this effect was important in evaluating our ability to answer the questions posed by this study.

The P-values (Table 2) for the Regional Descriptors AOC: In vs. Out (0.404-0.446) and Inland vs. Lakeshore (0.251-0.342) supported the null hypotheses of no difference between mink PRs for either descriptor. However, due to the small sample sizes the power of the GLM was low (≥ 0.254 for AOC: In vs. Out and ≥ 0.094 for

Lakeshore vs. Inland, although the actual power was somewhat higher because these numbers were calculated assuming only four sites in each region). In order to achieve a power of 0.8 for each test, given the differences between the means the number of replications (MustelaVision sites) in each region would have to have been 17 for AOC: In vs. Out, and 71 for Lakeshore vs. Inland. Although these results suggest that it may be possible to delist the “Degradation of Fish and Wildlife Populations” use impairment for mink in the Rochester Embayment, the low power of the tests (i.e., probability of finding no significant differences among treatments when differences exist) suggests that further evidence is needed before delisting can occur, such as results from the portion of this study on the levels of bioaccumulative chemicals of concern (BCCs) in mink tissues (Pagano and Haynes, in preparation).

Influence of Site Descriptors

None of Cover (brush, cattails, forest; Appendix D-1: Table D-1a), Habitat type (wetland, upland, mixed; Appendix D-1: Table D-1b), or underwater Ledge (presence or absence; Appendix D-1: Table D-1c) significantly affected PRs, but again, because of low sample sizes, statistical power to distinguish these effects was low (Table 3). While Habitat (wetland, upland, mixed) and Cover (cattail, brush, forest) had similar definitions, they applied to larger and smaller areas, respectively, around the MustelaVision sites. That the P-values for Cover were lower than those for Habitat implies that the likelihood of observing mink passage is more heavily influenced by site choice at a small scale, particularly the type of cover present at the camera site. This is similar to Bonesi and Macdonald’s (2004) finding that the habitat characteristics closest to the water had the strongest effect upon the duration of coexistence of otter and mink in England.

The Site Descriptor Tunnel had a highly significant effect on Passage Rates (Table 3; Appendix D-1: Table D-1d). This result was not unexpected. We were repeatedly told by trappers to place the cameras at culverts and bridges where mink following the water’s edge would enter the tunnel rather than leave the water to cross a road. Examination of the sites’ characteristics showed that the lack of a tunnel was the one thing that all sites with PRs of zero had in common (although not all sites without tunnels had PRs of zero). Unfortunately, the sites with tunnels were not evenly distributed between the Wetlands and the Mixed Landscapes. In the Wetlands regions, nine of 17 sites were tunnels, but in the Mixed regions only one of 12 sites was a tunnel (Chi-square = 6.196, DF = 1, P = 0.013). Thus, the effect of Tunnels on the PRs likely confounded the Wetlands effect in the Regional Descriptors GLM, indicating that careful camera site choice is an important factor in a study of this type, especially for animals with microhabitat preferences as specific as mink. Future studies should be certain that tunnels are fully represented in all experimental treatments or blocks.

Ecological Observations

Mink Groups in the AOC

The most exciting results were a hoped-for bonus of the study. We recorded four instances of mink families in the AOC: two in the BBWMA (Lakeshore-In AOC) and two in the BLKCK (Inland-In AOC) region. (We also recorded family groups at Route 77 in the INWR Inland-Out of AOC, but none in the LOSPW Lakeshore-AOC region.) At the Sackett Road site in the BLKCK region (AOC-Inland) on 23 June 2003, we recorded

one adult and two young mink traveling together. At the same site on 30 June 2004, we recorded two mink together. At Round Pond Creek in the BBWMA (AOC-Lakeshore) on 19 July 2004, we recorded one adult and four young. About two weeks later, on 5 August 2004, we recorded two animals traveling together at that site.

As mentioned above, mink are normally solitary except for mating pairs and mothers with kits. In two of the recordings of multiple mink in the AOC, it was obvious that there was one adult and several young. Since fathers take no part in raising the young (USDA Forest Service 2005), the adults observed with young were assumed to be their mothers. In the two cases in which we recorded two mink traveling together in the AOC, and their relationship was not obvious, we assumed that they were family members because these recordings were made during summer, before the young would have dispersed, rather than during mating season. This assumption is supported by Mitchell (1961) who reported that mink often travel in pairs, either two kits or a mother and daughter, until late fall.

This proof of reproduction of mink in the AOC, especially along the lakeshore, may justify delisting the current “Bird or Animal Deformities or Reproductive Problems” use impairment for the Rochester Embayment of Lake Ontario.

Observed Behaviors of Mink

Nocturnality of Mink

Many sources indicate that mink are primarily nocturnal (Birks and Linn 1982, Linscombe *et al.* 1982, Jamison 1983, Krause 1984, Geary 1985, Allen 1986, Eagle and Whitman 1987, Illinois Natural History Survey 2001, USDA Forest Service 2005). Our observations refuted this widely held belief. In 2003 and 2004, respectively, 65 of 109 mink passages (59.6%) and 71 of 116 (61.2%) were recorded as “Day,” i.e., the majority of mink passages were observed by natural light rather than the camera’s IR illumination. There was a statistical trend ($0.05 < P < 0.1$) toward a greater number of day than night Passages (Chi-square = 0.058, DF = 1, P = 0.809), which would not be the case if mink were mostly nocturnal. Given the observed ratio of Day to Night passages of 3:2, even if we had missed one-third of the night passages the conclusion of non-nocturnality would not be affected. This scenario is extremely unlikely, as the infrared VCR trigger is more sensitive at night because of the surroundings being cooler and the lack of IR from the sun. Also, since most animals’ eyes are highly reflective in the IR, and warm-blooded animals emit IR, we often saw them long before they entered the pool of IR that illuminated the entire animal on camera. Finally, the frequency at which we observed mice outside the IR pool of illumination at night causes us to believe that we would not often have missed a mink.

Although our definition of “Day” could apply during the twilight hours of dawn and dusk, it was obvious when watching the videos, judging by sun and shadow angles and knowing the orientation of the camera, that most of the “Day” passages took place in broad daylight, many at midday. Linscombe *et al.* (1982) cited reports by Gerell (1969) that females with kits were primarily diurnal and by Marshall (1936) that both sexes were most active between dawn and dusk in winter. However, neither of these reports would explain our findings, as less than 4% of our observations were of family units, and we were not recording during winter. Birks and Linn (1982) reported that 75% of inter-den or long distance movements were made at night, speculating that that may be why most are trapped at night and thus thought to be nocturnal.

Repeated Passages

On a number of occasions, mink were seen passing through the camera's field of view several times during the same day or night. They might then not be seen again for days or weeks, having apparently moved out of the area. At Cayuga Pool in the INWR (Inland-Out of AOC) on 27 June 2003, a mink crossed the field of view eight times while exhibiting searching behavior and covering the area thoroughly before disappearing. As it occurred in broad daylight, we were able to judge by shadow angles that this happened within a fairly short period of time (minutes rather than hours). This corresponds with Gerrell's (1970) reports of small-scale oscillatory movements superimposed upon larger-scale movements covering the home range.

Flight from a Predator

At the same location on the same day, a very different behavior was seen—a mink ran fast and straight through the entire field of view, disappearing in seconds. The probable explanation for this unusual behavior was following several feet behind the fleeing mink—the shadow of a large bird such, probably a hawk, was easily visible. Eagle and Whitman (1987) and Dunstone (1993) reported that mink are preyed upon by hawks, owls and eagles.

This potential for predation from overhead may explain why Cover was such an important factor in determining Passage Rates at a camera site. Mason and Macdonald (1983), Allen (1987), Eagle and Whitman (1987) and Yamaguchi *et al.* (2003) agree that mink prefer to stay under cover and avoid open areas. In retrospect, we realized that most of the sites with zero PRs had no cover immediately adjacent to the water's edge, though it may have been less than a meter away. At one of these sites, Cole Road in the BLKCK region in 2004, we recorded a hawk taking a duck from alongside the bank.

Predation on a Fish

At a stream site in the BLKCK region (Inland-In AOC) on 23 June 2004, we recorded a mink catching a fish. Although this was a night shot, and the event occurred at the edge of the IR field of illumination, recognition of the event was aided by the fact that this particular camera had a microphone. We heard a splash, and then saw the mink on the bank holding a fish; the mink had apparently dived into the water after sighting the fish, exactly as Dunstone (1993) and Eagle and Whitman (1987) reported.

Ice-breaking

At Bill's Point, a site in the BBWMA (Lakeshore-In AOC) on 25 November 2002, we recorded a mink breaking up a thin film of ice which was forming over an area where mink had frequently been seen swimming before. The mink swam under the ice and butted its head up against it until it broke; it repeated this behavior many times until most of the ice film was broken up, before swimming out of view.

CONCLUSION

The central question addressed by this study was: Are there differences in the relative abundance of lakeshore and inland mink populations in and out of the AOC? Our MustelaVision data tentatively suggest that there are no differences in mink populations inside and outside the AOC or between the lakeshore and inland areas, but the statistical power of our tests was low due to small sample sizes. We also showed that (1) landscape-scale features (wetland complexes) and microhabitat factors (tunnels) are key predictors

of mink presence or absence at a sampling site, (2) mink are successfully reproducing in the AOC, and (3) mink are not chiefly nocturnal. While the data in this report alone do not support delisting the RELO AOC use impairments for wildlife population degradation and reproductive problems, in combination with the chemical data also collected in this study (Pagano and Haynes, in preparation), we believe the time for delisting is approaching in the near future.

Many researchers have tried to estimate mink abundances. Some rely upon harvest records (cf. Linscombe 1982, Eagle and Whitman 1987), but this method can be confounded by trapping conditions, weather, number of trappers working the area, and other factors. Other studies rely on live-trapping (cf. Mitchell 1961, Halliwell and Macdonald 1996), but this can be fatal to mink. Mitchell reported that over 5% of trapped mink died upon capture, and Barker (1991) reported that, although released alive and apparently unharmed, mink may die of stress-related gastric hemorrhaging within a few days. Finally, some studies (cf. Mason and Macdonald 1983, Sidorovich and Macdonald 2001, Racey and Euler 2003) rely on the abundance of mink signs such as tracks, scats and scent posts, all of which can be easy to overlook and hard to relate to numbers of individuals if their home ranges overlap.

Our literature review found only two other studies that referred to the use of video cameras in population monitoring, neither of which was comparable to this study. Rutberg *et al.* (2004) used hand-held video cameras to record deer while driving around the perimeter of their study area, but this was only part of their effort, most of which focused on counting deer while “beating” or “driving” them with large numbers of people on foot. Westera *et al.* (2003) used video cameras to count fish attracted to bait stations in an effort to estimate abundances, but unlike our study they were not recording natural rates of passage. Hence we believe that we have developed a novel method that shows potential for monitoring relative population size, with appropriate care in camera placement, and has the added benefit of revealing the natural behaviors of the animals under study.

ACKNOWLEDGMENTS

We are extremely grateful to Jeffrey Wellman who put in many hours designing, building and maintaining the MustelaVision systems. We are also very grateful to Randy Baase who spent many hours in the field servicing each MustelaVision system weekly for months and who taught us much about mink. Thanks are also due to other trappers who taught us about mink: Matt Lochner (who also helped set up and service MustelaVisions) and Dan Frey, and to the landowners near the Bergen Swamp who allowed us to place MustelaVision systems on their property: “Doc” Fink, Dick Sands, Mel Reber and Al Burkhardt. We also thank Marc Chalupnicki for tabulating data and Albert Fulton for creating Figure 1. The government personnel who graciously allowed us to work in their territories include NYS DEC: Dan Carroll, Dave Woodruff and Heidi Bogner; NYS OPRHP: James Slusarczyk, and USFWS: Bob Lamoy and Paul Hess. We gratefully acknowledge grant C302399 from the New York State Great Lakes Protection Fund which made this study possible. Finally, we thank reviewers at NYS DEC and Dr. Daniel Simberloff, University of Tennessee at Knoxville, for their feedback, especially statistical, on an earlier version of this report.

LITERATURE CITED

- Allen, A.W. 1986. Habitat suitability index models: mink, revised. U.S. Department of the Interior Fish and Wildlife Service. Biol. Rept. 82 (10-127).
- Barker, I.H. 1991. Non-toxic Diseases of Mink and River Otter. Pages 19-20 *in* Addison, E.M., G.A. Fox, and M.G. Gilbertson (eds.), Proceedings of the Expert Consultation Meeting on Mink and Otter. International Joint Commission, Windsor ONT. ISBN 1-895085-32-2.
- Birks, J.D.S., and I.J. Linn. 1982. Studies on the home range of feral mink (*Mustela vison*). *Symposia of the Zoological Society of London* 49: 231–257.
- Bonesi, L., and D.W. Macdonald. 2004. Evaluation of sign surveys as a way to estimate the relative abundance of American mink (*Mustela vison*). *Journal of Zoology* 262(1): 65-72.
- Chittenden, M.E., Jr. 2002. Given a significance test, how large a sample size is large enough? *Fisheries* 27(8): 25-29.
- Dunstone, N. 1993. *The Mink*. T&AD Poyser. London.
- Eagle, T. C., and J. S. Whitman. 1987. Mink. Pages 614-625 *in* M. Novak, J. A. Baker, M. E. Obbard, and B. Mallock, eds. *Wild furbearer management and conservation in North America*. Ontario Trappers Assoc. and Ontario Ministry Nat. Resour.
- Geary, S.M. 1985. *Fur Trapping in North America, Revised Edition*. Winchester Press, New Century Publishers, Inc. Piscataway, NJ.
- Gerell, R. (1970). Home ranges and movements of the mink *Mustela vison* in southern Sweden. *Oikos* 21: 160-173.

- Halliwell, E.C., and D.W. Macdonald. 1996. American mink *Mustela vison* in the Upper Thames catchment: Relationship with selected prey species and den availability. *Biological Conservation* 76(1): 51-56.
- Haynes, J.M., Pagano, J.J., and Wellman, S.T. 2002. New York State Great Lakes Protection Fund Application for State Assistance—RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species.
- Illinois Natural History Survey. 2005. Mink Species Account. <http://www.inhs.uiuc.edu/dnr/fur/species/mink.html>
- Jamison, R. 1983. *The Trapper's Handbook*. DBI Books Inc. Northfield, IL.
- Krause, T. 1984. *National Trappers' Association Trapping Handbook*. United Graphics, Inc. Mattoon, IL.
- Linscombe, G., N. Kinler, and R.J. Aulerich. 1982. Mink (*Mustela vison*). Pages 629-643 in Chapman, G.A. and J.A. Feldhamer, eds. *Wild mammals of North America—Biology, management, economics*. John Hopkins University Press. Baltimore, MD
- Mason, C.F., and S.M. Macdonald 1983. Some factors influencing the distribution of mink (*Mustela vison*). *Zoological Society of London* 200: 281-283.
- Mitchell, J.L. 1961. Mink movements and populations on a Montana river. *J. Wildlife Management* 25(1): 48-54.
- National Trappers' Association. 2005. The Mink. www.nationaltrappers.com/Mink.html
- Racey, G.D., and D.L. Euler. 2003. Changes in mink habitat and food selection as influenced by cottage development in central Ontario." *J. Applied Ecology* 20(2): 387-401.
- RAP. 1993. Stage I Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- RAP. 1997. Stage II Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- Rutberg, A.T., R.E. Naugle, L.A. Thiele, and I.K.M. Liu. 2004. Effects of immunocontraception on a suburban population of white-tailed deer *Odocoileus virginianus*. *Biological Conservation* 116(2): 243-250.
- Schladweiler, J.L., and G.L. Storm. 1969. Den-use by mink. *J. Wildlife Management* 33(4): 1025-1026.
- Sidorovich, V., and D.W. Macdonald. 2001. Density dynamics and changes in habitat use by the European mink and other native mustelids in connection with the American mink expansion in Belarus. *Netherlands J. Zoology* 51(1): 107-126.
- Stevens, R.T., T.L. Ashwood, and J.M. Sleeman. 1997. Fall-early winter home ranges, movements, and den use of male mink, *Mustela vison*, in Eastern Tennessee. *Canadian Field-Naturalist* 111: 312-314.

- USDA Forest Service. 2005. Fire Effects Information System, Wildlife Species: *Mustela vison*. <http://www.fs.fed.us/database/feis/wildlife/mammal/muvi/all.html>
- Westera, M., P. Lavery, and G. Hyndes. 2003. Differences in recreationally targeted fishes between protected and fished areas of a coral reef marine park. *J. Experimental Marine Biology and Ecology* 294(2): 145-168.
- Yamaguchi, N., S. Rushton, D.W. Macdonald, and J.G. Kie. 2003. Habitat preferences of feral American mink in the Upper Thames." *J. Mammalogy* 84(4): 1356-1373.
- Yamaguchi, N., and D.W. Macdonald. 2003. The burden of co-occupancy: Intraspecific resource competition and spacing patterns in American mink, *Mustela vison*. *J. Mammalogy* 84(4): 1341-1355

Tables

Table 1. MustelaVision camera sites characterized by regional and site descriptors.

MustelaVision Camera Site	Region	Regional Descriptors			Site Descriptors				
		AOC: In		Lakeshore vs.	Landscape	Cover	Habitat	Ledge	Tunnel
		vs. Out	Inland						
Bogus Creek 2003	BBWMA	In	Lakeshore	Wetlands	Forest	Upland	No	No	
Bogus Creek 2004	BBWMA	In	Lakeshore	Wetlands	Forest	Upland	No	No	
Larkin Creek A 2003	BBWMA	In	Lakeshore	Wetlands	Forest	Mix	Yes	No	
Larkin Creek B 2004	BBWMA	In	Lakeshore	Wetlands	Forest	Mix	Yes	No	
Round Pond Creek 2003	BBWMA	In	Lakeshore	Wetlands	Brush	Mix	Yes	Yes	
Round Pond Creek 2004	BBWMA	In	Lakeshore	Wetlands	Brush	Mix	Yes	Yes	
Bill's Point 2003	BBWMA	In	Lakeshore	Wetlands	Cattails	Wetland	No	Yes	
Cranberry Pond Trib 2004	BBWMA	In	Lakeshore	Wetlands	Forest	Mix	No	Yes	
Black Creek/ Rte 19 2003	BLKCK	In	Inland	Mix	Forest	Upland	Yes	No	
Black Creek/ Rte 19 2004	BLKCK	In	Inland	Mix	Forest	Upland	Yes	No	
Black Creek/ W Sweden Rd 2003	BLKCK	In	Inland	Mix	Forest	Upland	Yes	No	
Black Creek Trib/ W Sweden Rd 2004	BLKCK	In	Inland	Mix	Brush	Upland	No	No	
Black Creek/ Mud City Rd 2003	BLKCK	In	Inland	Mix	Cattails	Wetland	Yes	No	
Black Creek/ Cole Rd 2004	BLKCK	In	Inland	Mix	Brush	Upland	Yes	No	
Black Creek Trib Sackett Rd 2003	BLKCK	In	Inland	Mix	Brush	Mix	No	No	
Black Creek Trib Sackett Rd 2004	BLKCK	In	Inland	Mix	Brush	Mix	No	No	
Rte 63 2003	INWR	Out	Inland	Wetlands	Cattails	Mix	No	No	
Rt 77 2003	INWR	Out	Inland	Wetlands	Cattails	Wetland	No	Yes	
Rt 77 2004	INWR	Out	Inland	Wetlands	Cattails	Wetland	No	Yes	
Cayuga Pool 2003	INWR	Out	Inland	Wetlands	Cattails	Wetland	No	No	
Cayuga Pool 2004	INWR	Out	Inland	Wetlands	Cattails	Wetland	No	No	
Sour Springs Road 2003	INWR	Out	Inland	Wetlands	Cattails	Wetland	Yes	Yes	
Sour Springs Road 2004	INWR	Out	Inland	Wetlands	Cattails	Wetland	Yes	Yes	
Feeder Road 2003	INWR	Out	Inland	Wetlands	Cattails	Mix	No	No	
Albion Rd 2003	INWR	Out	Inland	Wetlands	Cattails	Mix	No	Yes	
Albion Rd 2004	INWR	Out	Inland	Wetlands	Cattails	Mix	No	Yes	
Bald Eagle beaver den 2004	LOSPW	Out	Lakeshore	Mix	Brush	Mix	No	No	
Yanty Creek 2004	LOSPW	Out	Lakeshore	Mix	Brush	Mix	No	No	
Bald Eagle west bank 2004	LOSPW	Out	Lakeshore	Mix	Forest	Upland	No	No	
Yanty Culvert #2 2004	LOSPW	Out	Lakeshore	Mix	Cattails	Wetland	No	Yes	

Table 2. Results of analysis of variance (GLM) of mink Passage Rates by Regional Descriptors: AOC: In vs. Out; Inland vs. Lakeshore; and Landscape: Wetlands Complex vs. Mixed Habitat (Full MiniTab output in Appendix C). Bold indicates a significant difference. *The significance of the results for Landscape make those power calculations irrelevant.

Regional Descriptor	PR Used	N	Mean (SE)	P-value	Power
AOC: In vs. Out					
	Max			0.404	0.254
AOC: In		16	0.0768 (0.0238)		
AOC: Out		13	0.1675 (0.0611)		
	Min			0.446	0.248
AOC: In		16	0.0399 (0.0130)		
AOC: Out		13	0.0859 (0.0339)		
Lakeshore vs. Inland					
	Max			0.251	0.096
Inland		17	0.1424 (0.0479)		
Lakeshore		12	0.0821 (0.0309)		
	Min			0.342	0.094
Inland		17	0.0720 (0.0266)		
Lakeshore		12	0.0443 (0.0168)		
Landscape: Wetlands vs. Mixed					
	Max			0.026	0.112*
Mixed Habitat		12	0.0352 (0.0106)		
Wetlands Complex		17	0.1755 (0.0479)		
	Min			0.042	0.112*
Mixed Habitat		12	0.0180 (0.0056)		
Wetlands Complex		17	0.0905 (0.0267)		

Table 3. Results of ANOVAs of mink Passage Rates by Site Descriptors: Cover, Habitat, Ledge, and Tunnel (Full MiniTab output in Appendix D). Bold indicates a significant difference.

Site Descriptor	PR Used	N	Mean (SE)	P-value	Power
Cover	Max			0.166	0.268
Brush		8	0.0942 (0.0387)		
Cattails		12	0.1843 (0.0643)		
Forest		9	0.0490 (0.0255)		
	Min			0.193	0.259
Brush		8	0.0566 (0.0227)		
Cattails		12	0.0935 (0.0360)		
Forest		9	0.0200 (0.0100)		
Habitat	Max			0.245	0.279
Mix		13	0.1125 (0.0459)		
Upland		8	0.0512 (0.0289)		
Wetland		8	0.1918 (0.0757)		
	Min			0.341	0.197
Mix		13	0.0706 (0.0305)		
Upland		8	0.0204 (0.0115)		
Wetland		8	0.0842 (0.0337)		
Ledge	Max			0.872	0.052
Absent		18	0.1214 (0.0421)		
Present		11	0.1109 (0.0459)		
	Min			0.970	0.050
Absent		18	0.0610 (0.0235)		
Present		11	0.0597 (0.0244)		
Tunnel	Max			0.004	0.743
Absent		19	0.0557 (0.0227)		
Present		10	0.2348 (0.0658)		
	Min			0.001	0.874
Absent		19	0.0222 (0.0075)		
Present		10	0.1333 (0.0387)		

Figures

Figure 1. Map showing placement of MustelaVision systems in four regions during 2003 and 2004. Each red triangle is the site of one MV system. In LOSP West, there were actually four sites, but the triangle symbols are almost superimposed in pairs due to their proximity. RELO is the Rochester Embayment of Lake Ontario. (Map courtesy of Albert Fulton 2005.)

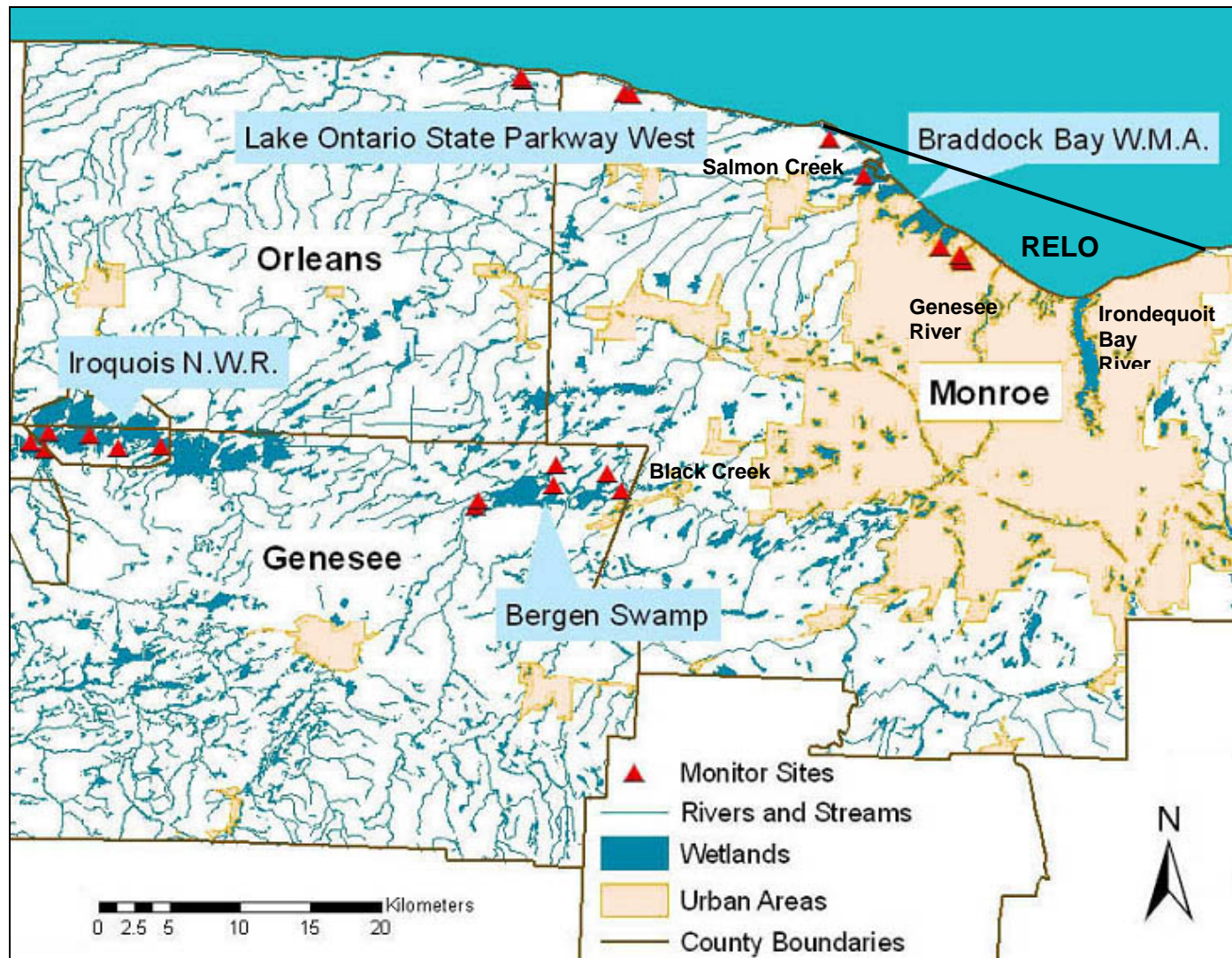


Figure 2. Mink Passage Rates calculated for each camera site during 2003 and 2004. The bars show the range between the Min and Max Passage Rates, based on the Max and Min number of Trap Nights, respectively, at each site. The sites are grouped by Region (In AOC vs. Out of AOC, and Inland vs. Lakeshore) labeled at the bottom. Data from both years at one site are shown side by side.

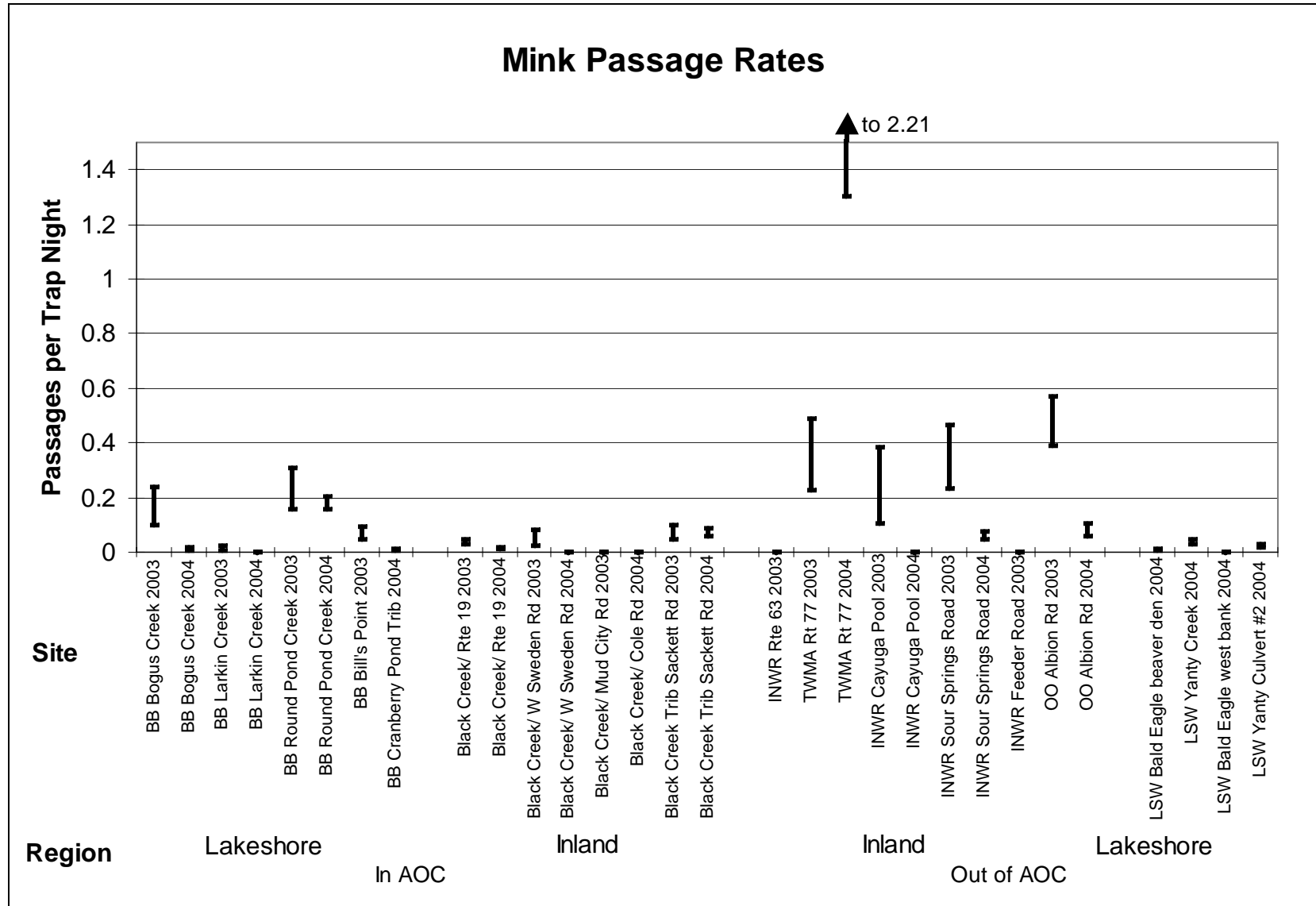
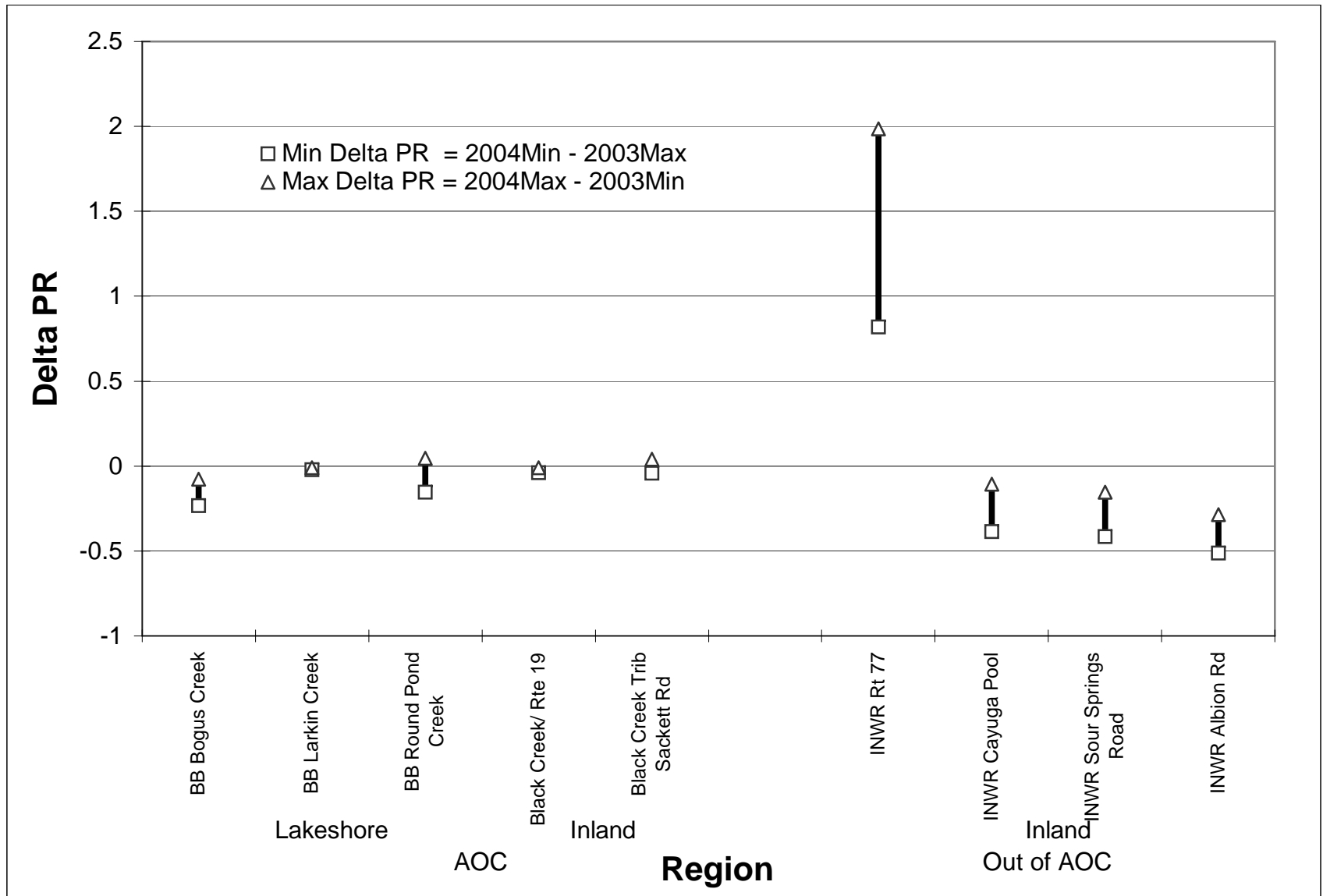


Figure 3. Yearly changes (Delta Passage Rates) in mink passages for sites at which data were taken both years. BB = Braddock Bay Wildlife Management Area, INWR = Iroquois National Wildlife Refuge.



Appendices

Appendix A: MustelaVision Operations

Appendix A-1: MustelaVision System

Figure A-1a. MustelaVision system: camera head on stake at right, VCR on platform in front of battery, protective circuit board mounted underneath VCR platform.



Figure A-1b. Schematic of a MustelaVision system. The remote monitor is used only during field testing and camera alignment.

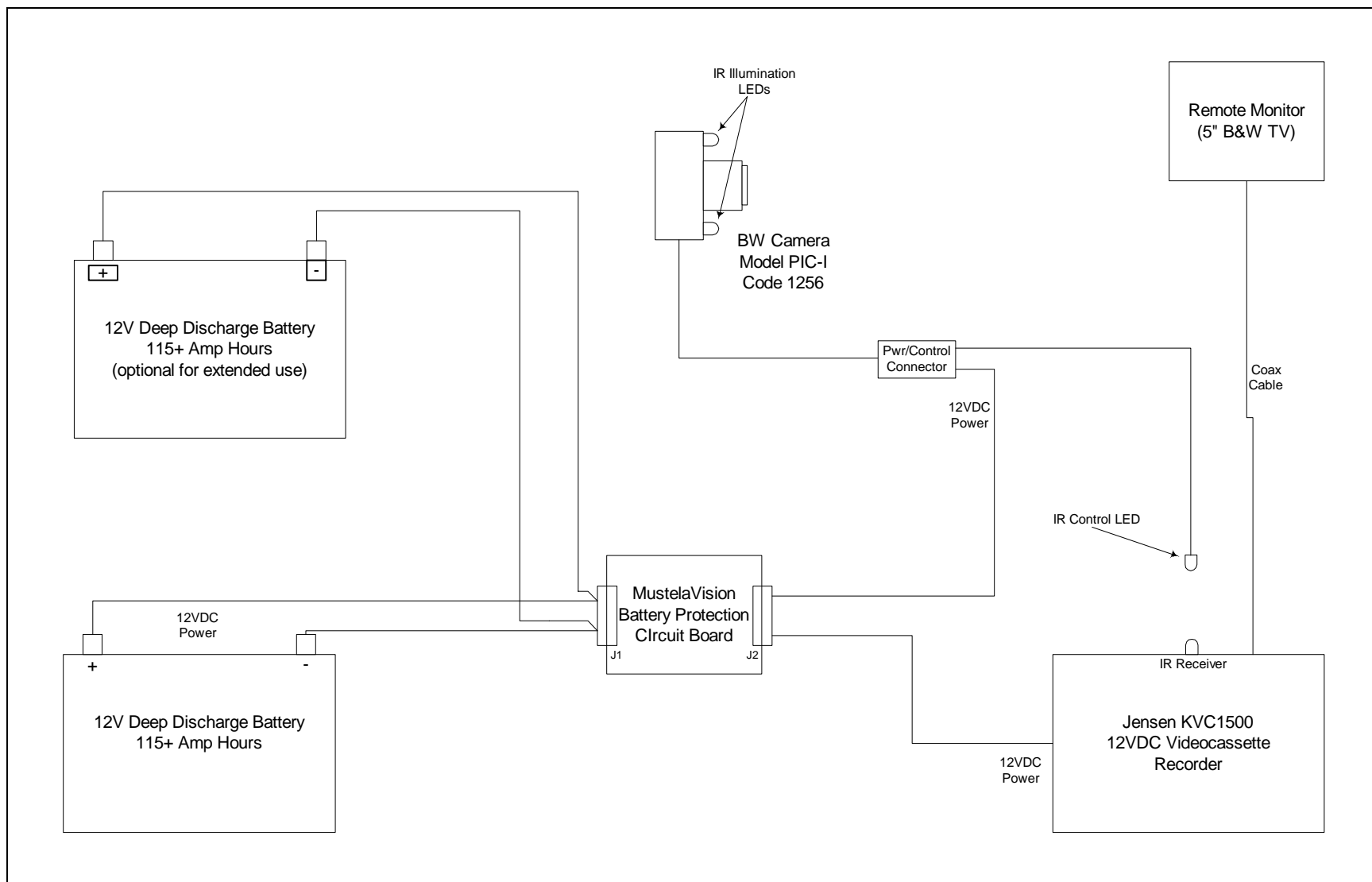
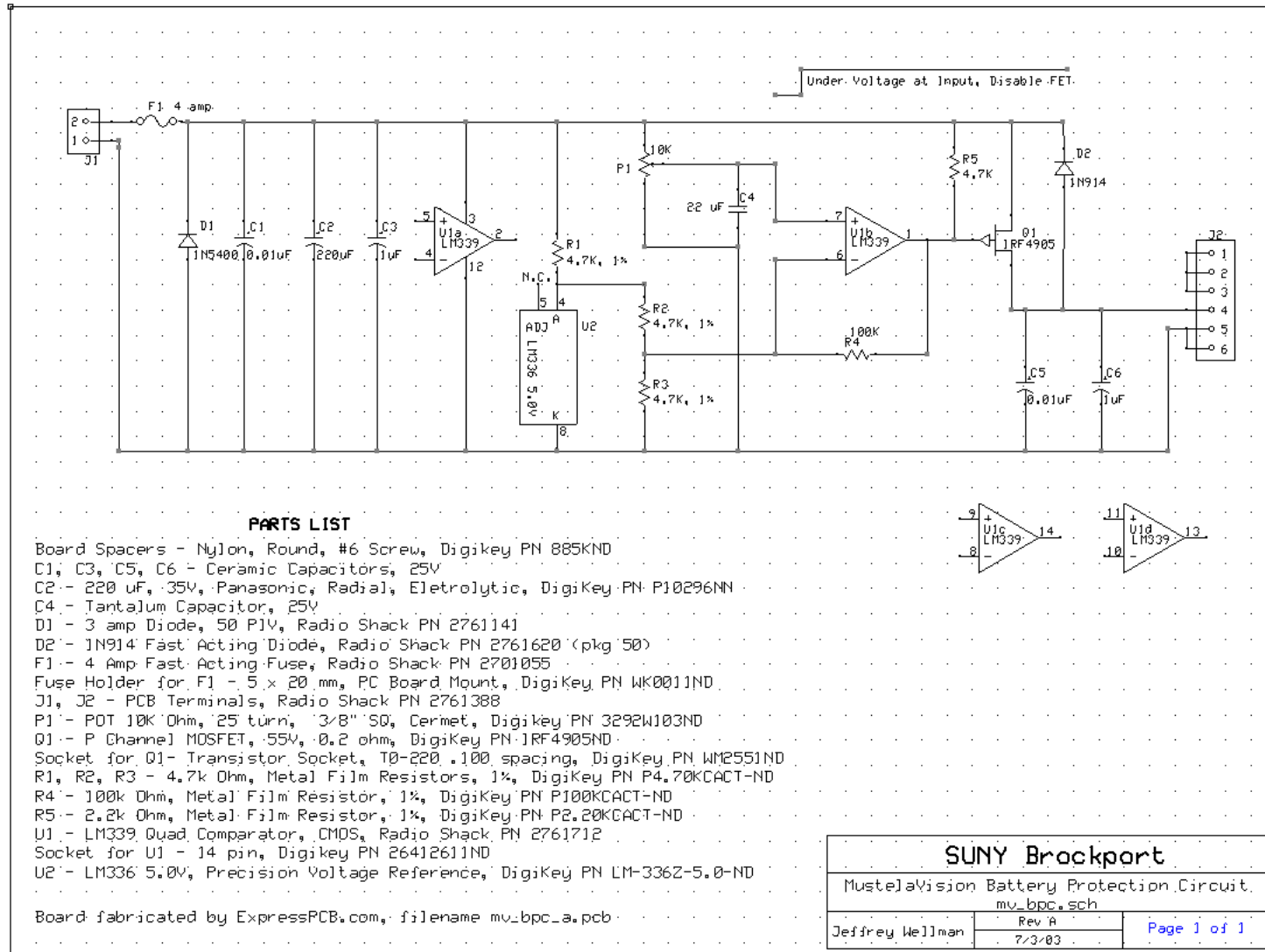


Figure A-1c. Back view of a MustelaVision camera head mounted on stake at the Round Pond Creek site in the BBWMA.



Figure A-1d. Battery protection circuit board, which protected against over discharge of battery, reversal of battery hook-up, and component fusing.



Appendix A-2: Data Record Sheets

Figure A-2a. Sample System Log. During each field service session, we recorded the date of service, the ID numbers of the videotape cassettes and batteries inserted, and any comments such as mink tracks observed, operational failures, or other pertinent information.

**MustelaVision
System Log C**

Location (Creek/Road)	Tape #	Battery #	2003 Start Date	End Date	Tracks/Comments	# Mink Sights
RPC	5	1	6/14/03	6/21		0
	10	10	6/21	6/26		0
	20	2	6/26	7/2	mink tracks	1
	20	10	7/2	7/8		1
	39	23	7/8	7/14	battery down. VCR won't record, nil on tape 39	0
	14	3	7/14	7/20	did record (tape full of veg/wind)	0
	9	21	7/20	7/27		
	39	2	7/28	8/4	tracks: mink + weasel	2
	3	23	8/4	8/11		1
	15	12	8/11	8/18		0
	17	26	8/18	8/25		
	15	4	8/25	9/1		0
	18	26	9/1	9/8		1
	13	4	9/8	9/16		0
	3	12	9/16	9/23	weasel + small mink	1
6	26	9/23	10/1		0	
20	13	10/1	10/7		0	
12	14	10/7	10/14		1	
37	11	10/14	10/21		0	
15	13	10/21	10/27	pulled system	0	
2004			2004			
Sour Springs Rd	1	22	6/25	7/1		0
	8	28	7/1	7/8		1
	7	39	7/8	7/15		0
	39	1	7/15	7/22		0
	38	36	7/22	7/29		0
	16	47	7/29	8/5	Fuse blew	0
	18	37	8/5	8/12	Replaced fuse	?
	25	39	8/12	8/19		0
	19	66	8/19	8/26		0

System Log C

Figure A-2b. Sample Tape Log. The start and end dates for each session generated the Max Trap Nights (TN) for that session, and the day (D) and night (N) periods, recorded as they occurred in the video, yielded the Min TN for that session.

**MustelaVision
Tape Log 36**

Location (Creek/Road)	System	2003 Date		Tape Counter		Trigger		Animal
		Start	End	Session	Cue	Day	Night	
INWR Feeder Rd	H	7/7 DD	7/13 N	19:		1		Heron
Bogus Cr./LOSP	A	7/28 DD	8/4 NN	3:16:55		11		Ducks Heron Spider Raccoons
Tonawanda Rt 77	E	8/14 DDDDDD	8/21 NNNNN	1:42:	* 21:31	1	✓	Heron Iron woods (20+ min) Mink Mink Day - Tonawanda Rt77 08-14-03 Mink " " " " " - B ^A
					* 22:27	1	✓	"Mink-Night-Tonawanda Rt77 08-14-03
					* 26:26	1	✓	2 Minks 2 Minks-Night-Ton 77-08-14-03
					* 27:57	1	✓	Mink Mink Day Tonawanda Rt77 08-15-03
					* 59:32	1	✓	
INWR Sour Springs	G	8/28 DDDD	9/4 NNN	1:08:	* 7:40	1	✓	Ducks Mink Mink - INWR - Sour Springs 08-26-03
						1	✓	Rail? Mink Mink-Tail-Night- INWR - Sour Springs 08-28-03
					54:04	1	✓	" Couldn't capture Ducks under attack - INWR - Sour Springs 08-28-03 Ducks being attacked - INWR - Sour Springs 08-28-03
					57:26	1	✓	Mink? (fur in LR corner) Muskrat?
					1:05:22		✓	
Tonawanda Rt 77	E	9/25 DDDD	10/2 NNNN	29:	* 12:26	9/26	✓	Muskrats weasel or Mink Mouse
					* 13:31	9/27	✓	Mink-Tonawanda Rt77-day-09-27-03
					* 20:05	9/29	✓	Weasels v - 09-29-03
						1		Heron

Appendix B: Passage Rate Analyses (Delta PRs)

Table B-1. Mean Delta PRs

One-Sample T: Dmin, Dmax

Test of mean Delta PR = 0 vs mean Delta PR not = 0

Variable	N	Mean	StDev	SE Mean
Dmin	9	0.159	0.693	0.231
Dmax	9	-0.109	0.392	0.131

Variable	95.0% CI	T	P
Dmin	(-0.374, 0.691)	0.69	0.511
Dmax	(-0.410, 0.192)	-0.83	0.428

Table B-2. Delta PRs: AOC: In (trapped for this study) vs. Out (trapped historically)

Two-Sample T-Test and CI: Dmax, Area

Area	N	Mean	StDev	SE Mean
AOC	5	-0.0970	0.0919	0.041
Out	4	-0.124	0.630	0.32

Difference = mu (AOC) - mu (Out)

Estimate for difference: 0.027

95% CI for difference: (-0.984, 1.038)

T-Test of difference = 0 (vs not =): T-Value = 0.08 P-Value = 0.938 DF = 3

Two-Sample T-Test and CI: Dmin, Area

Area	N	Mean	StDev	SE Mean
AOC	5	-0.0018	0.0497	0.022
Out	4	0.36	1.09	0.54

Difference = mu (AOC) - mu (Out)

Estimate for difference: -0.361

95% CI for difference: (-2.092, 1.369)

T-Test of difference = 0 (vs not =): T-Value = -0.66 P-Value = 0.554 DF = 3

Appendix C: Regional Descriptor Analyses

Appendix C-1. Descriptive Statistics: Regional Descriptors

Table C-2a. Descriptive Statistics: AOC: In vs. Out

		AOC:In						
Variable	vs. Out	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	In	16	0	0.0768	0.0238	0.0950	0.00000	0.00281
	Out	13	0	0.1675	0.0611	0.2203	0.00000	0.00000
Min	In	16	0	0.0399	0.0130	0.0522	0.00000	0.00210
	Out	13	0	0.0859	0.0339	0.1222	0.00000	0.00000
Variable	AOC	Median	Q3	Maximum				
Max	In	0.0340	0.0960	0.3077				
	Out	0.0471	0.4249	0.5714				
Min	In	0.0147	0.0536	0.1552				
	Out	0.0317	0.1671	0.3902				

Table C-2b. Descriptive Statistics: Inland vs. Lakeshore

		Lakeshore						
Variable	vs.Inland	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	Inland	17	0	0.1424	0.0479	0.1977	0.00000	0.0000
	Lakeshore	12	0	0.0821	0.0309	0.1069	0.00000	0.0116
Min	Inland	17	0	0.0720	0.0266	0.1097	0.00000	0.0000
	Lakeshore	12	0	0.0443	0.0168	0.0583	0.00000	0.00774
Variable	Lakeshore	Median	Q3	Maximum				
Max	Inland	0.0755	0.2449	0.5714				
	Lakeshore	0.0251	0.1751	0.3077				
Min	Inland	0.0290	0.0827	0.3902				
	Lakeshore	0.0119	0.0834	0.1552				

Table C-2c. Descriptive Statistics: Landscape: Wetlands vs. Mix

		Landscape						
Variable	Landscape	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	Mix	12	0	0.0352	0.0106	0.0368	0.00000	0.00000
	Wetlands	17	0	0.1755	0.0479	0.1974	0.00000	0.00562
Min	Mix	12	0	0.01802	0.00557	0.01930	0.00000	0.00000
	Wetlands	17	0	0.0905	0.0267	0.1102	0.00000	0.00420
Variable	Landscape	Median	Q3	Maximum				
Max	Mix	0.0240	0.0744	0.0968				
	Wetlands	0.0938	0.3462	0.5714				
Min	Mix	0.01220	0.03106	0.05556				
	Wetlands	0.0494	0.1551	0.3902				

Appendix C-2: Main Effects Plots

Figure C-1a. Main effects plot showing effects of Regional Descriptors on Max PR.

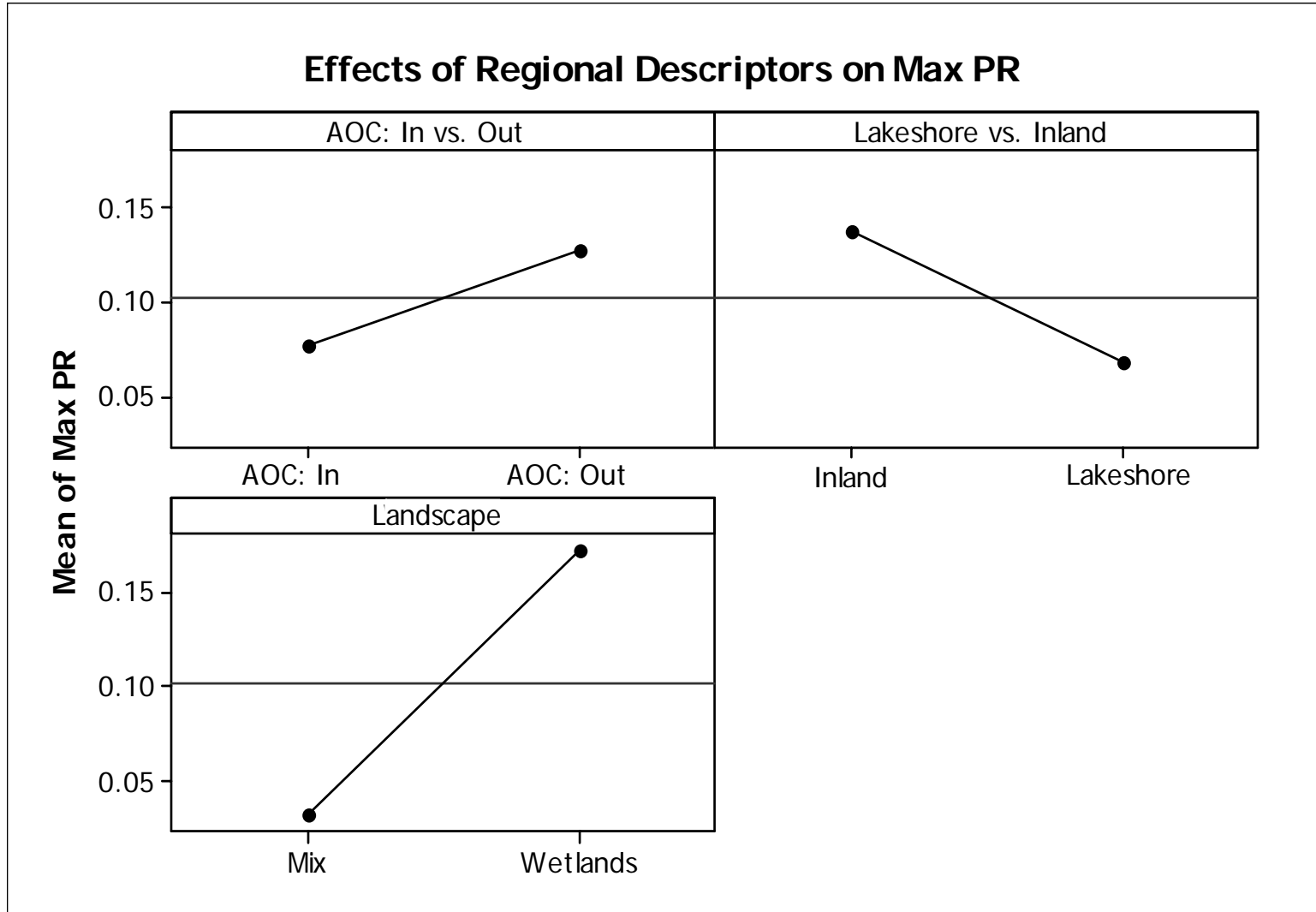
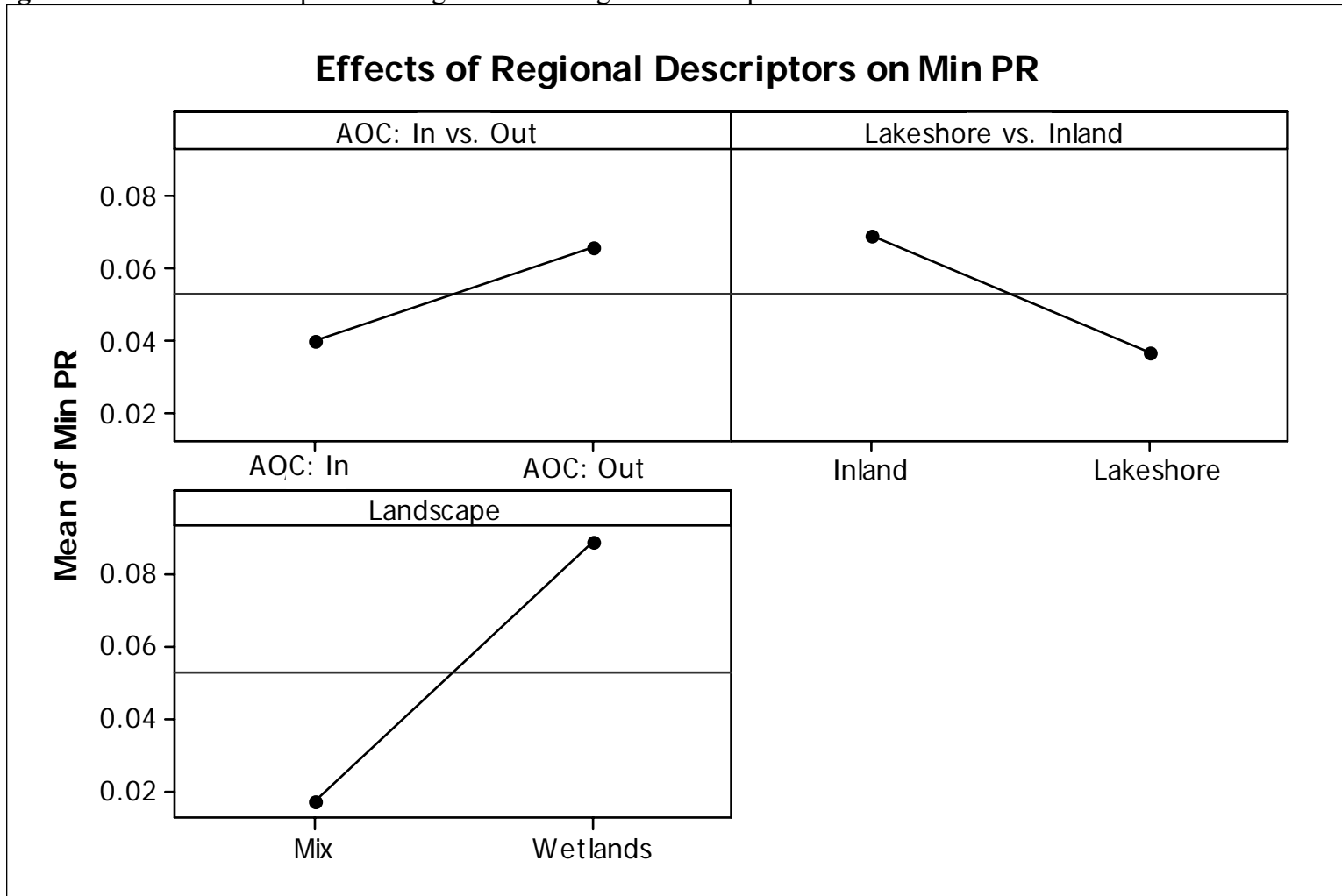


Figure C-1b. Main effects plot showing effects of Regional Descriptors on Min PR.



Appendix C-3. General Linear Model: Regional Descriptors

Table C-3. General Linear Model: Regional Descriptors

Factor	Type	Levels	Values
AOC	fixed	2	AOC, Out
Shore	fixed	2	Inland, Shore
Wetlands	fixed	2	Mix, Wetlands

Analysis of Variance for Max, using Adjusted SS for Tests

Source	DF	Seq SS	Adj SS	Adj MS	F	P
AOC	1	0.05893	0.01658	0.01658	0.72	0.404
Lakeshore	1	0.01323	0.03176	0.03176	1.38	0.251
Wetlands	1	0.12819	0.12819	0.12819	5.56	0.026
Error	25	0.57615	0.57615	0.02305		
Total	28	0.77651				

S = 0.151810 R-Sq = 25.80% R-Sq(adj) = 16.90%

Unusual Observations for Max

Obs	Max	Fit	SE Fit	Residual	St Resid
24	0.571429	0.231957	0.050603	0.339472	2.37 R

R denotes an observation with a large standardized residual.

Analysis of Variance for Min, using Adjusted SS for Tests

Source	DF	Seq SS	Adj SS	Adj MS	F	P
AOC	1	0.015166	0.004399	0.004399	0.60	0.446
Shore	1	0.002547	0.006896	0.006896	0.94	0.342
Wetlands	1	0.033783	0.033783	0.033783	4.59	0.042
Error	25	0.183830	0.183830	0.007353		
Total	28	0.235325				

S = 0.0857507 R-Sq = 21.88% R-Sq(adj) = 12.51%

Unusual Observations for Min

Obs	Min	Fit	SE Fit	Residual	St Resid
24	0.390244	0.117964	0.028584	0.272280	3.37 R

R denotes an observation with a large standardized residual.

Appendix D: Site Descriptor Analyses

Appendix D-1. Descriptive Statistics

Table D-1a. Descriptive Statistics: Cover

Variable	Cover	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	Brush	8	0	0.0942	0.0387	0.1095	0.000000000	0.00313
	Cattails	12	0	0.1843	0.0643	0.2227	0.000000000	0.000000000
	Forest	9	0	0.0490	0.0255	0.0765	0.000000000	0.00562
Min	Brush	8	0	0.0566	0.0227	0.0643	0.000000000	0.00188
	Cattails	12	0	0.0935	0.0360	0.1248	0.000000000	0.000000000
	Forest	9	0	0.0200	0.0100	0.0300	0.000000000	0.00420

Variable	Cover	Median	Q3	Maximum
Max	Brush	0.0673	0.1759	0.3077
	Cattails	0.0846	0.4450	0.5714
	Forest	0.0196	0.0655	0.2407
Min	Brush	0.0397	0.1302	0.1552
	Cattails	0.0471	0.1969	0.3902
	Forest	0.00847	0.0247	0.0963

Table D-1b. Descriptive Statistics: Habitat

Variable	Habitat	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	Mix	13	0	0.1125	0.0459	0.1655	0.000000000	0.00562
	Upland	8	0	0.0512	0.0289	0.0819	0.000000000	0.000000000
	Wetland	8	0	0.1918	0.0757	0.2142	0.000000000	0.00746
Min	Mix	13	0	0.0706	0.0305	0.1101	0.000000000	0.00376
	Upland	8	0	0.0204	0.0115	0.0324	0.000000000	0.000000000
	Wetland	8	0	0.0842	0.0337	0.0954	0.000000000	0.00385

Variable	Habitat	Median	Q3	Maximum
Max	Mix	0.0471	0.1538	0.5714
	Upland	0.0189	0.0744	0.2407
	Wetland	0.0846	0.4450	0.4857
Min	Mix	0.0317	0.1065	0.3902
	Upland	0.00871	0.0268	0.0963
	Wetland	0.0471	0.1969	0.2299

Table D-1c. Descriptive Statistics: Ledge

Variable	Ledge	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	No	18	0	0.1214	0.0421	0.1788	0.000000000	0.000000000
	Yes	11	0	0.1109	0.0459	0.1524	0.000000000	0.000000000
Min	No	18	0	0.0610	0.0235	0.0999	0.000000000	0.000000000
	Yes	11	0	0.0597	0.0244	0.0810	0.000000000	0.000000000
Variable	Ledge	Median	Q3	Maximum				
Max	No	0.0385	0.1391	0.5714				
	Yes	0.0476	0.2022	0.4651				
Min	No	0.0236	0.0676	0.3902				
	Yes	0.0204	0.1550	0.2299				

Table D-1d. Descriptive Statistics: Tunnel

Variable	Tunnel	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Max	No	19	0	0.0557	0.0227	0.0991	0.000000000	0.000000000
	Yes	10	0	0.2348	0.0658	0.2081	0.0112	0.0641
Min	No	19	0	0.02219	0.00753	0.03281	0.000000000	0.000000000
	Yes	10	0	0.1333	0.0387	0.1225	0.00847	0.0374
Variable	Tunnel	Median	Q3	Maximum				
Max	No	0.0182	0.0833	0.3846				
	Yes	0.1538	0.4703	0.5714				
Min	No	0.00840	0.03175	0.10753				
	Yes	0.1065	0.2275	0.3902				

Appendix D-2. One-Way ANOVAs: Site Descriptors

Table D-2a. Cover

Factor Type Levels Values
Cover fixed 3 Brush, Cattails, Forest

One-way ANOVA: Max versus Cover

Source	DF	SS	MS	F	P
Cover	2	0.1000	0.0500	1.92	0.166
Error	26	0.6765	0.0260		
Total	28	0.7765			

S = 0.1613 R-Sq = 12.88% R-Sq(adj) = 6.18%

One-way ANOVA: Min versus Cover

Source	DF	SS	MS	F	P
Cover	2	0.02794	0.01397	1.75	0.193
Error	26	0.20739	0.00798		
Total	28	0.23533			

S = 0.08931 R-Sq = 11.87% R-Sq(adj) = 5.09%

Table D-2b. Habitat

Factor Type Levels Values
Habitat fixed 3 Mix, Upland, Wetland

One-way ANOVA: Max versus Habitat

Source	DF	SS	MS	F	P
Habitat	2	0.0797	0.0398	1.49	0.245
Error	26	0.6968	0.0268		
Total	28	0.7765			

S = 0.1637 R-Sq = 10.26% R-Sq(adj) = 3.36%

One-way ANOVA: Min versus Habitat

Source	DF	SS	MS	F	P
Habitat	2	0.01869	0.00935	1.12	0.341
Error	26	0.21663	0.00833		
Total	28	0.23533			

S = 0.09128 R-Sq = 7.94% R-Sq(adj) = 0.86%

Table D-2c. Ledge

Factor Type Levels Values
Ledge fixed 2 No, Yes

One-way ANOVA: Max versus Shelf

Source	DF	SS	MS	F	P
Shelf	1	0.0008	0.0008	0.03	0.872
Error	27	0.7758	0.0287		
Total	28	0.7765			

S = 0.1695 R-Sq = 0.10% R-Sq(adj) = 0.00%

One-way ANOVA: Min versus Shelf

Source	DF	SS	MS	F	P
Shelf	1	0.00001	0.00001	0.00	0.970
Error	27	0.23531	0.00872		
Total	28	0.23533			

S = 0.09336 R-Sq = 0.01% R-Sq(adj) = 0.00%

Table D-2d. Tunnel

Factor Type Levels Values
Tunnel fixed 2 No, Yes

One-way ANOVA: Max versus Tunnel

Source	DF	SS	MS	F	P
Tunnel	1	0.2101	0.2101	10.02	0.004
Error	27	0.5664	0.0210		
Total	28	0.7765			

S = 0.1448 R-Sq = 27.06% R-Sq(adj) = 24.36%

One-way ANOVA: Min versus Tunnel

Source	DF	SS	MS	F	P
Tunnel	1	0.08089	0.08089	14.14	0.001
Error	27	0.15444	0.00572		
Total	28	0.23533			

S = 0.07563 R-Sq = 34.37% R-Sq(adj) = 31.94%

Appendix 2

Age, Size, and Stable Isotope Data of Mink Populations, and a Predictive Model of Bioaccumulation of Chemicals of Concern in the Rochester Embayment of Lake Ontario

Sara T. Wellman and James M. Haynes

Department of Environmental Science and Biology
State University of New York
College at Brockport
350 New Campus Drive
Brockport, NY 14420-2973

June 2006

OVERVIEW

This report is the second of four resulting from project C302399 funded by the New York Great Lakes Protection Fund in 2004 to address use impairments related to water quality identified in the Remedial Action Plan for the Rochester Embayment of Lake Ontario (RELO RAP). It deals with ages, sizes and trophic positions (stable isotope analysis) of mink (*Mustela vison*) in the study area, and provides a predictive model for exposure levels of mink to bioaccumulative chemicals of concern (BCCs). The previous report (Wellman and Haynes 2005) addressed the development and use of videocapture (MustelaVision) systems that established the presence and reproduction of mink in and out of the RELO RAP Area of Concern (AOC). Two more reports will be written in 2006: (1) levels of BCCs in mink tissues (Pagano and Haynes, in preparation), and (2) a literature review of the effects of BCCs on mink (Wellman, in preparation). Because mink are the most sensitive known species to BCCs, the results of this four-part project will determine whether delisting the fish and wildlife reproduction impairment for the RELO AOC can be recommended.

TABLE OF CONTENTS

INTRODUCTION	56
MATERIALS AND METHODS.....	56
Specimen collection.....	57
Collection.....	57
Processing.....	57
Handling & Shipping	58
Isotope Analysis.....	58
Aging	58
Data Analysis.....	58
Modeling.....	59
RESULTS AND DISCUSSION	58
Mink Age, Length and Weight	59
Age vs. Regional Descriptors	60
Isotopes	61
Isotope Data vs. Age.....	61
Isotope Data vs. Regional Descriptors.....	61
Construction of Bioaccumulation Model.....	61
Calculation of Trophic Level.....	61
Calculation of Aquatic Portion of Diet.....	62
Modeling Exposure of Mink in the AOC to BCCs.....	63
SUMMARY	65
ACKNOWLEDGMENTS	65
LITERATURE CITED	65
TABLES	
Table 1. Table of Age versus Regional Descriptors.....	69
Table 2. Table of $\delta^{15}\text{N}$ versus Regional Descriptors	70
Table 3. Table of $\delta^{13}\text{C}$ versus Regional Descriptors.....	71
Table 4. Predicted daily exposure levels of mink in the AOC	72
FIGURES	
Figure 1. Map showing the four regions referred to in the study.	73
Figure 2. Ages of mink trapped.	73
Figure 3. Plot of individual mink ages in each region.	73
Figure 4. Stable isotope values of carbon sources in a Lake Superior wetland.....	74
APPENDICES	
A. Mink age, length and weight relationships.	76
B. Age versus Regional Descriptors	77
C. Isotope data versus Age.	81
D. Isotope Data versus Regional Descriptors	83

Age, Size, and Stable Isotope Data of Mink Populations, and a Predictive Model of Bioaccumulation of Chemicals of Concern in the Rochester Embayment of Lake Ontario

INTRODUCTION

In the 1980s, the binational (Canada, U.S.) International Joint Commission (IJC) began the process of creating and implementing remedial action plans (RAPs) in 43 contaminated areas of concern (AOCs) throughout the Great Lakes Basin. The IJC established 14 “use impairments” that could cause a local area to be “listed” as an AOC, including “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems.” In the Rochester AOC, both uses were defined as impaired because “very few” mink were then being trapped or observed within 2 miles of the lake (RAP 1993, 1997). This study was part of a project (Haynes *et al.* 2002) to determine if populations of mink on the shore of the Rochester Embayment of Lake Ontario (RELO) are negatively impacted by bioaccumulative chemicals of concern (BCCs) and, if so, whether the BCCs are originating in the embayment watershed or elsewhere.

The RELO AOC includes the Embayment, a 35 square mile portion of Lake Ontario south of a line between Bogus Point in the town of Parma and Nine Mile Point in the town of Webster (both in Monroe County, New York); adjacent wetlands and bays; and the six mile reach of the Genesee River, from the Lower Falls to the mouth at Lake Ontario. The RAP also includes the subwatersheds of Salmon Creek, the Genesee River and Irondequoit Creek (RAP 1993, 1997; **Figure 1**).

The initial questions addressed by this portion of the study were: 1) Can stable isotope analysis be used to evaluate mink diets, at lakeshore and inland areas in and out of the AOC, in terms of trophic levels and terrestrial and aquatic food sources? 2) Can stable isotope results be used to construct a food web/bioaccumulation model for mink in the Rochester AOC to predict body burdens of BCCs in mink in relation to their diets?

Stable isotopes (SIs) of carbon and nitrogen are often used to evaluate trophic webs of ecosystems to give lifetime, integrated estimates of both trophic level and dietary sources for organisms. Both ^{12}C and ^{14}N have stable, heavier isotopes (^{13}C and ^{15}N) which occur naturally, and the heavier and lighter isotopes are differentially absorbed and metabolized by organisms. Usually the lighter isotopes are excreted preferentially, leading to a relative enrichment of the heavier isotopes in organisms relative to their environment or diet. These enrichments are measurable through mass spectrometry, and are reported in parts per thousand ($\delta\text{‰}$) relative to a standard:

$$\delta X = [(R_{\text{sample}} - R_{\text{standard}}) / R_{\text{standard}}] \times 10^3$$

where X is ^{13}C or ^{15}N and R is the corresponding ratio $^{13}\text{C}/^{12}\text{C}$ or $^{15}\text{N}/^{14}\text{N}$. The standard for carbon is PeeDee Belemnite (PDB) limestone, and the standard for nitrogen is atmospheric nitrogen (Fry 1991).

Selective excretion of ^{14}N over ^{15}N by animals results in an increase of approximately 3.4 ‰ in the $\delta^{15}\text{N}$ at each trophic level; thus, ^{15}N analysis can determine the trophic level at which an animal feeds (Peterson and Fry 1987, Cabana and Rasmussen 1994). Carbon is also enriched between trophic levels, but at a much lower rate, between 0 and 1 ‰ . Because freshwater algae have a much less negative $\delta^{13}\text{C}$ than terrestrial plants (e.g., terrestrial leaves $\delta^{13}\text{C} = -27$ to -31‰ versus algae $> -17\text{‰}$; Collier and Lyon 1991), ^{13}C analysis can differentiate

between these as original sources of carbon in a diet, indicating whether the diet is primarily of aquatic or terrestrial origin.

Once trophic level and percent aquatic diet are known, the exposure level for each BCC can be calculated using a model adapted from Sample *et al.* (1996). The model takes into account the concentration of the BCC in the water, daily food and water ingestion rates, proportion of the diet originating from aquatic carbon sources, body weight of the animal, and bioaccumulation factors (BAF) for each BCC. The BAF is dependent upon the trophic level and the octanol-water partition coefficient of the compound (Sample *et al.* 1996).

Our approach was to conduct stable isotope analysis for ^{13}C and ^{15}N on tissues from the same mink collected for BCC analyses (Pagano and Haynes, in preparation). We tested the null hypotheses that there are no differences in stable isotope ratios ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) among regions (AOC: In vs. Out, Lakeshore vs. Inland). We used our results for trophic level of prey in mink diets to model the bioaccumulation of selected BCCs in mink in the AOC for later comparison with the results of Pagano's and Haynes' (in preparation) study.

Because it was desirable to know the ages of the mink for the Pagano and Haynes study (in preparation), we had the minks' teeth aged and used those results to answer further questions, such as: How do body length and weight relate to ages of trapped mink? Do the ages of mink trapped vary in and out of the AOC and between lakeshore and inland areas? Do stable isotope values in mink vary with age?

MATERIALS AND METHODS

Specimen collection, processing and handling

Collection

Mink carcasses were collected from trappers (after skinning) in five areas. We divided the study area into four Regions: Inland/AOC, Lakeshore/AOC, Inland/Out of AOC, and Lakeshore/Out of AOC. Both Lakeshore regions were identical to those defined for the MustelaVision videocapture study (Wellman and Haynes 2005) — Lakeshore/AOC (Wetlands) was the Braddock Bay Wildlife Management Area (BBWMA), and Lakeshore/Out of AOC (Mixed) was the Lake Ontario State Parkway west of Route 19 (LOSPW). However, for this study Inland/AOC (Mixed) included any animals taken in the AOC watershed more than 5 km from the lakeshore, and Inland/Out of AOC included animals taken from the Tug Hill Plateau, as well as from the Iroquois National Wildlife Refuge area, to provide two presumably “clean” control areas. Thus, Inland/Out of AOC was the only region that included both Wetlands and Mixed habitats. For the purpose of analysis, we described each collection area using the Regional Descriptors AOC: In versus Out; Lakeshore versus Inland; and Landscape: Wetlands (large wetlands complex) versus Mixed Habitat.

Carcasses were put in plastic bags and frozen by the trappers as soon as possible. The trappers completed log sheets indicating the date and location of capture for each animal, as well as the trapper's name and contact information. Carcasses were assigned specimen numbers in the order in which they were collected, and the specimen number, date and location of capture were written on the plastic bags with a permanent marker.

Processing

We thawed the frozen mink carcasses overnight in a refrigerator before processing them. Because some trappers removed the tails when skinning the carcasses, we removed all other tails before weighing and measuring to obtain comparable measures of body weight and length. We

placed carcasses in hexane-rinsed aluminum pans or aluminum foil for resection, and all utensils used were rinsed with hexane before each use. We placed muscle tissue from the left thigh of each carcass into a hexane-rinsed glass specimen bottle, labeled with the specimen number and tissue type, and froze it. We extracted two canine teeth from each mink and placed them in similarly labeled specimen envelopes. We recorded the body weight, tail-less body length, and weight of each tissue sample (except teeth) on a separate sheet for each mink, along with its specimen number and collection record.

Handling & Shipping

We shipped the muscle tissue samples, frozen and packaged with dry ice, to Cornell University's Stable Isotope Laboratory (COIL). We let the teeth dry in their paper envelopes and shipped them to Matson's Laboratory (Milltown, MT) for aging.

Isotope Analysis

At COIL, stable isotope analyses for ^{13}C and ^{15}N were conducted using a continuous flow Elemental Analyzer (NC2500, CE Elantech, New Jersey) interfaced with an Isotope Ratio Mass Spectrometer (IRMS) (Delta Plus, Thermo Electron Corp., Germany). Strict quality control procedures included standards to: (1) test for instrument linearity and define instrument response for the determination of elemental composition, and (2) measure stability of precision and accuracy over the length of a run (Arthur Kasson, COIL, personal communication).

Aging

Matson's Laboratory aged the mink teeth using a standardized species- and tooth-specific cementum analysis. Mammalian teeth, like trees, show seasonal growth rings when properly stained (Matson 1981). Matson's assumed a birth date of April 1 for all mink, and reported ages in whole years only. We then calculated the additional partial year for each mink between April 1 and its capture date, and added that to Matson's result to obtain the age of each mink.

Data Analysis

We used Microsoft® Excel 2000 for data management and non-statistical calculations. For statistical analyses, we used Minitab™ Statistical Software Release 14.13 (2005). We conducted regression analyses to evaluate the relationships between age and both body length and weight. We computed descriptive statistics for age versus the Regional Descriptors (AOC: In vs. Out, Lakeshore vs. Inland, and Landscape: Wetland vs. Mixed Habitat) and then used Minitab's General Linear Model (GLM, a 2-way ANOVA with unbalanced cells, Tukey's pairwise comparisons) to analyze the relationships between age and the Regional Descriptors. We also estimated historical trapping pressure in each area based on our conversations with DEC employees and trappers, and assigned Trapping Pressure values as a covariate in the GLM. Trapping Pressures were: 1 = mink not previously targeted by trappers (Lakeshore/Out of AOC and Inland/AOC), 2 = mink trapped historically (Lakeshore/AOC and Inland/Out of AOC). We then conducted regression analyses for each isotope ($\delta^{13}\text{C}$ and δN) versus age, and finally computed descriptive statistics and GLMs for each isotope versus the Regional Descriptors.

We estimated the power of the GLMs using Minitab's 2-Level Factorial power calculator (Factors = 3, Corners = 4, Replicates = minimum number for any level of the factor of concern (Regional Descriptor or Trapping Pressure), Effects = the smallest differences between the means for each factor). Although this calculator is not designed for unbalanced cells, using the minimum number of replicates among treatments produced a conservatively low estimate of the actual power (Minitab support staff 2006, personal communication).

Modeling

Trophic level is calculated by dividing the $\delta^{15}\text{N}$ value of an organism by the change in $\delta^{15}\text{N}$ per trophic level, usually 3.4‰ (Minigawa and Wada 1984, Vander Zanden and Rasmussen 1999, Doucett 1999). Calculating percent aquatic diet using $\delta^{13}\text{C}$ requires 1) determining the $\delta^{13}\text{C}$ value in tissue, 2) estimating the difference between the $\delta^{13}\text{C}$ values in the tissue and in the diet, and 3) calculating the relative contributions of aquatic and terrestrial sources required to yield the estimated $\delta^{13}\text{C}$ of the diet (DeNiro and Epstein 1978). COIL's analysis provided the data for step 1. Literature review provided appropriate estimated values for step 2. The equation for step 3, calculating the proportion of a diet (%_A) originating from one of two dietary sources of carbon with different $\delta^{13}\text{C}$ values, is

$$\%_A = \frac{\delta^{13}\text{C}_{\text{animal}} - \delta^{13}\text{C}_B - f \cdot x}{\delta^{13}\text{C}_A - \delta^{13}\text{C}_B} \times 100,$$

where $\delta^{13}\text{C}_{\text{animal}}$ is the stable-isotope ratio in the animal, $\delta^{13}\text{C}_A$ and $\delta^{13}\text{C}_B$ are the stable-isotope ratios of the two carbon sources, f is the trophic fractionation between the animal and its diet, and x is the trophic position of the animal (adapted from Doucett 1999).

Once the trophic level and aquatic portion of an animal's diet are known, the animal's exposure to a BCC can be modeled knowing the concentration of the compound in ambient water. The equation to predict the daily exposure level of an animal to a BCC in water is

$$\text{Exp} = \frac{C_w [W + (F \times P_{\text{aq}} \times \text{BAF})]}{bw},$$

where Exp is the exposure from both food and water, C_w is the concentration of the BCC in the water, W and F are the daily water and food consumption rates in L/day and g/day, respectively, P_{aq} is the aquatic proportion of the diet, BAF is the bioaccumulation factor of the chemical of concern (based on the trophic level of the animal and the octanol-water coefficient k_{ow} , a measure of hydrophobicity of the compound), and bw is the body weight of the animal in grams (adapted from Sample *et al.* 1996).

RESULTS AND DISCUSSION

Mink Age, Length and Weight

Although we used 41 mink in this study, 12 were not aged due to damaged or lost teeth. The ages of the mink ranged from 0.60 to 4.75 years; 41% (12/29) of those aged were less than one year old, while only 2 mink (7%) were over 4 years old (Figures 2, 3). Eagle and Whitman (1997) stated that wild mink rarely live longer than 3 years, and cited a study (Adams and Chapman 1981) in which only one of 169 trapped mink had reached the age of four years. Mitchell (1961) reported almost complete turnover of a population in Montana within three years. In contrast, Aulerich *et al.* (1999) reported life spans of 7-11 years in ranch mink.

Neither body length ($r = 0.007$, $P = 0.655$) nor body weight ($r = 0.019$, $P = 0.476$) were correlated with the age of the mink trapped (Appendix A: Figures A1, 2), a result explained by earlier studies. Mitchell (1961) reported that juvenile females attained their adult weight by mid-August, and males sometime during their first winter. Dunstone (1993) reported that mink reach adult body size by about 10 months (their first breeding season) although males may continue to gain weight after the first year (and body weight may cycle with seasons). Aulerich *et al.* (1999) reported that mink reach 95% or more of their adult body length by 16 weeks of age (September).

Trapping season in the study area does not open until late November, except for the Tug Hill Plateau, where it opens in October (our contracted trapper for the Rochester Embayment also started in October with a special collector's permit). Thus, the mink should have been close to their adult weight by their first trapping season, and a correlation between weight and age would not be expected.

Females in our study averaged 32.6 (\pm 1.0) cm body length and 456.8 (\pm 42.0) g body weight, while mean male body length was 37.4 (\pm 2.7) cm and mean body weight 781.5 (\pm 26.8) g. These means are somewhat smaller than reported by Mitchell (1961) in Montana, where males averaged 1150 g and non-pregnant females 600 g. The average male body length and weight in our study were 15% and 71% greater, respectively, than the female means. Dunstone (1993) reported that males are typically about 75% heavier than females, and Aulerich *et al.* (1999) reported male body weights 68% and 85% higher than females of the same age.

Age vs. Regional Descriptors

None of the Regional Descriptors (or their interactions) had any effect on the age of mink trapped (P-values ranged from 0.304 to 0.404; Table 1, also Appendix B: Table B2). However, in examining the ages of the mink from each area (Figure 3), and the descriptive statistics (Appendix B, Table B1), we noticed that the largest difference between the mean ages occurred for the Landscape descriptor and that none of the mink from the Iroquois National Wildlife Refuge (INWR, Inland/Out of AOC) area had reached one year of age (Appendix A, Figure A2). Also, seven of the eight mink older than three years were trapped in the Inland/AOC and Lakeshore/Out of AOC areas where, to our knowledge, mink had not previously been trapped. Despite the small sample size from INWR (only four of those five mink could be aged), we hypothesized that trapping pressure might have an effect on the ages of the mink trapped, and assigned the Trapping Pressure levels to each area, as described above, to enable further investigation.

The descriptive statistics (Appendix B, Table B1) indicated that Trapping Pressure did have an effect on the ages of mink trapped. In previously non-trapped areas (Lakeshore/Out of AOC and Inland/AOC) the mean age (and standard error) was 2.6 (0.37), the median 3.0, and the maximum 4.8 years. In trapped areas (Lakeshore/AOC, Inland/Out of AOC) the mean age was 1.4 (0.27), the median 0.8, and the maximum 3.6.

Having observed these differences, we used Trapping Pressure as a covariate in the GLM (Appendix B, Table B3), (which forced us to drop the Landscape Descriptor and the AOC X Lakeshore Interaction due to empty cells). The results were that the P-value for Trapping Pressure was 0.017, while the P-values for the Regional Descriptors rose to 0.634 or higher (Table 1). The low power levels (Table 1) calculated for AOC: In vs. Out and Lakeshore vs. Inland are due to the small differences between the means for each factor (0.4 and 0.3 years, respectively), which can be attributed to the length of the trapping season (0.2 years in western NY to 0.5 years on the Tug Hill Plateau and for our contracted trapper with his special permit). In contrast, the difference in ages between historically trapped and non-trapped mink was 1.3 years, significant especially when compared to the short (3-4 yr) life spans of wild mink.

Our conclusion that Trapping Pressure is a biologically significant factor in the ages of the mink trapped is supported by Eagle and Whitman (1997). They reported higher proportions of juveniles in heavily trapped populations than in untrapped populations and hypothesized that reproduction or juvenile survival may be suppressed in untrapped areas that may reach their carrying capacity for mink. Mitchell (1961) also found juvenile to adult female ratios higher in a

commercially trapped area than in a historically non-trapped area. Unfortunately, we have no way of knowing whether the trapped mink in heavily trapped areas truly represented the population structure, or whether older mink are more trap-wary and less likely to be caught and counted. In historically non-trapped areas, trap-wariness should not be a factor and the trapped mink might better represent the population structure.

The presence of mink less than one year old in each area implies reproduction in all areas. It is possible that the young of the year trapped in an area were recently dispersed newcomers, as Gerell (1970) reported one dispersing mink traveling 45 km and another averaging 800 m per day over 27 days. However, the MustelaVision study (Wellman and Haynes 2005) recorded family units in all areas except Lakeshore/Out of AOC, confirming reproduction in the AOC.

Isotopes

Isotope Data vs. Age

Although the P-value of 0.043 indicated a significant correlation between age and $\delta^{15}\text{N}$, the R^2 of 0.144 is small enough to conclude that there is no real effect (Appendix C1). For example, Mink #58 (AOC: In/Inland/Mixed) had a high $\delta^{15}\text{N}$ (14.85) but was only 0.87 years (10.4 months) old. Minigawa and Wada (1984) found no relationship between age and $\delta^{15}\text{N}$ in marine mussels or in tilapia. Age also had no effect on $\delta^{13}\text{C}$ ($R^2 = 0.008$; $P = 0.641$; Appendix C2). Our results were similar to, but the reverse of, Kiriluk *et al.*'s (1995) findings that the correlation between $\delta^{15}\text{N}$ and age in lake trout was not significant, but $\delta^{13}\text{C}$ and age were weakly correlated ($r^2 = 0.22$, $P = 0.0001$).

Isotope Data vs. Regional Descriptors

Two Regional Descriptors, AOC: In vs. Out and Lakeshore vs. Inland, had significant effects on $\delta^{15}\text{N}$ values of the mink (Table 2; also Appendix D2). Mink in the AOC had higher $\delta^{15}\text{N}$ values than mink out of the AOC ($P = 0.025$), and Lakeshore mink had higher $\delta^{15}\text{N}$ values than Inland mink ($P = 0.002$). Landscape had no effect ($P = 0.613$, power = 0.711). The highest mean $\delta^{15}\text{N}$ of the four regions was found in the AOC (Lakeshore/Wetlands), where the mean $\delta^{15}\text{N} = 13.2$ ‰ (SE = 0.5). The highest individual $\delta^{15}\text{N}$ value (16.9 ‰, Mink #17) was also found in the Lakeshore/AOC area, while the lowest was found in the AOC/Inland area (9.2 ‰, Mink #24) (Appendix D1).

None of the Regional Descriptors had significant effect on the $\delta^{13}\text{C}$ values of the mink studied (Table 3 and Appendix D2). The highest (most positive) $\delta^{13}\text{C}$ value (-28.29 ‰) was found in Mink #17 (Lakeshore/AOC) while the lowest (-19.89 ‰) was Mink #5 (Inland/Out of AOC) (Appendix D2). The low power levels are due to the small differences between the means for each factor (Table 3).

Construction of Bioaccumulation Model

Calculation of Trophic Level

Using the $\delta^{15}\text{N}$ value of 11.9 (grand mean of 41 mink in our study) and the commonly accepted value of 3.4‰ $\delta^{15}\text{N}$ per trophic level, the average trophic level of mink in our study was 3.50. If we use 3.5‰ $\delta^{15}\text{N}$ per trophic level, as reported by Cabana and Rasmussen (1994) for the Lake Ontario food web, the trophic level of our mink averaged 3.40. The higher mean $\delta^{15}\text{N}$ of 13.2 for mink in Lakeshore/AOC resulted in higher values for the trophic level of those mink (3.87 or 3.76, using 3.4‰ or 3.5‰ $\delta^{15}\text{N}$ per trophic level, respectively). All of these values agree well with estimates found in the literature; USEPA (1995a) reported estimates for mink prey levels ranging from 2.5 to 2.9, which would imply the minks' trophic level to be 3.5 to 3.9.

For the purpose of the model, we chose 3.8 as the trophic level of mink, for several reasons. As the ultimate purpose of the project is to protect mink populations in the AOC, we wanted to represent the mink in the AOC at greatest risk, those along the lakeshore. We chose an intermediate value between those based on the two estimates of the change in δN per trophic level, because, although Cabana and Rasmussen (1994) studied Lake Ontario, they analyzed only the pelagic food web. Therefore, their estimate is not fully appropriate for the diet of mink that feed in the littoral zone of the lake, associated wetlands, or in streams.

The mean $\delta^{15}N$ in mink from the Lakeshore was 1.5‰ higher than the mean from Inland areas. This represents almost one-half of a trophic level difference between Lakeshore and Inland mink. The mean $\delta^{15}N$ for mink in the AOC was 1.1‰ higher than the mean out of the AOC, about one-third of a trophic level. If the Lakeshore minks' diet includes a higher proportion of aquatic-based prey, then inferring a higher trophic level for Lakeshore than Inland mink may be confounded by the fact that aquatic primary producers typically have $\delta^{15}N$ values 1-3‰ higher than terrestrial values (Figure 4). However, the hypothesis that Lakeshore mink feed at a slightly higher trophic level than Inland mink is supported by BCC analysis of the mink tissues (James Pagano, SUNY Oswego, personal communication).

Calculation of Aquatic Portion of Diet

In reference to the three steps involved in calculating the aquatic portion of the diet of an animal using $\delta^{13}C$, the calculated mean $\delta^{13}C$ in mink muscle tissue from the Lakeshore/AOC was -25 (Table 3). For step 2 of the calculation, since the mink tissue used was thigh muscle, we relied on DeNiro and Epstein's (1978) report that the $\delta^{13}C$ of thigh muscle from mice fed two different diets was depleted (more negative) by 1.9 ± 0.5 ‰ from the $\delta^{13}C$ value of the diets. Adding this value (+1.9‰) to the $\delta^{13}C$ value of the mink tissue (-25) yielded a value of -23.1‰ $\delta^{13}C$ for the diet of mink in BBWMA.

DeNiro and Epstein (1978) examined insects, nematodes, snails and mice, and found that $\delta^{13}C$ values varied significantly between tissues of the same animal such that no single tissue truly represents the $\delta^{13}C$ value of the whole animal. They also reported that $\delta^{13}C$ values differ among conspecifics raised on different diets, but that differences between an animal and its diet are similar within a species regardless of diet. Thus, mouse $\delta^{13}C$ values would not be applicable to mink, but since the ^{13}C fractionation is due to metabolic processes that are similar in all mammals, herbivores and carnivores, our estimate for the ^{13}C depletion from diet to thigh muscle should be satisfactory. Focken and Becker (1998) cautioned that the lipid content of tissue in their study had such a strong influence on $\delta^{13}C$ ratios that the trophic shift was not constant among, or even within, species, and that within-species differences were sometimes higher than levels commonly assumed for trophic level shift. However, the mouse data was the best estimate we found.

We had several difficulties with step 3 of the calculation of the aquatic portion of the diet of mink. The formula,

$$\%_A = \frac{\delta^{13}C_{animal} - \delta^{13}C_B - f \cdot x}{\delta^{13}C_A - \delta^{13}C_B} \times 100,$$

works quite well if there are only two dietary sources of carbon with $\delta^{13}C$ values separated by 5‰ to 10‰ (with sample sizes of 50 to 15, respectively; Doucett 1999). So, if the $\delta^{13}C$ values of terrestrial and aquatic carbon sources in our study differed by at least 6‰, we should have had no problem in calculating the aquatic portion of the diet. For example, Balasse *et al.* (2005) were able to use the difference between terrestrial vegetation δC values (mean of -27 ‰) and seaweed

(ranging from -18.5 to -13.1 ‰) to determine that seaweed made a significant contribution to the diet of coastal sheep in Scotland.

The first problem was that wetlands have more than two sources of ^{13}C , including phytoplankton, C_3 vascular plants (terrestrial, emergent, floating-leaved, submersed), and epiphytic and filamentous algae. The second problem was variation within, and overlap among, $\delta^{13}\text{C}$ values in these sources. Figure 4 shows $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of vegetation and algae in a wetland associated with Lake Superior, along with values for phytoplankton from Lake Superior (Keough *et al.* 1996). Fry (1991) reported that $\delta^{13}\text{C}$ values for terrestrial plants range from -35 to -25 ‰, while algae range from -34 to -18 ‰. Other researchers (Peterson *et al.* 1985, France 1995, Albuquerque *et al.* 1997, Doucett 1999, Cloern *et al.* 2002, Choi *et al.* 2005) each reported different ranges of values for the sources mentioned above, with many of the ranges overlapping even in the same study. Peterson and Fry (1987) explained that the ^{13}C content of freshwater organisms depends on the source of the dissolved CO_2 in the water and upon the ^{13}C fractionation by those organisms. Heaton (1999) reported that plant $\delta^{13}\text{C}$ values were affected up to 2‰ by factors such as growing conditions (e.g., light, temperature, water and nutrient availability, air flow), variation between parts of the same plant, variation between individuals of the same species in the same location (genetic diversity, microhabitat, age), and seasonal or annual variations. DeNiro and Epstein (1978) noted that the $\delta^{13}\text{C}$ values of a single plant species can vary 5‰ or more in a growing season, and Kiriluk *et al.* (1995) found that Lake Ontario net plankton $\delta^{13}\text{C}$ values ranged from -33.10 in May to -21.92 in September 1992. Cloern *et al.* (2002) concluded that isotope studies to determine primary producers are confounded by overlap of the isotopic ratios of the primary producers and changes in the isotopic composition of plant matter as it degrades. They warned of the danger in applying isotopic data from one ecosystem to another, even in congeneric species. For these reasons, we reluctantly concluded that we could not use $\delta^{13}\text{C}$ values of our mink to determine the aquatic portion of their diet without having also analyzed samples of the vegetation (and the minks' known prey species) in the AOC.

Modeling Exposure of Mink in the AOC to BCCs

Although we were unable to calculate the proportion of aquatic foods in the diet of the mink in our study, we found several estimates in the literature. Although most diet studies only report frequencies of occurrence of diet items in scats, digestive tracts, or dens (e.g., USEPA 1993 summarizes the results of 19 such studies), USEPA (1995a) points out that this is not a good representation of biomass assimilated by the mink. However, USEPA (1995b) cited a study by Alexander (1977) reporting that the aquatic portion of minks' diets was 75% to 90%, based on wet weight of stomach contents year-round. Sample and Suter (1999) averaged the results of five studies to conclude that the aquatic portion of minks' diet is 54.6%. (The standard deviation for that average was reported as $\pm 0.21\%$, which seems in error, as the average included Alexander's 1977 study; it is much more likely that the standard deviation was 21%). USEPA (1995b) used both 90% and 50% to calculate Wildlife Values for DDT, Hg, 2,3,7,8-TCDD, and PCBs; therefore, we chose the same bounds on the aquatic portion of the diet of mink.

Other values needed for the model are the body weight of the animal (g), daily consumption rates of food (g/day) and water (L/day), the bioaccumulation factor (BAF) of the chemical of concern (which also requires knowing the k_{ow} of the compound), and the concentration of the BCC in the water. The mean body weight of females in our study was 456.8 (± 42.0) g, while males averaged 781.5 (± 26.8) g. Because we had six females and 35 males, we averaged the male and female means for a representative average body weight of 620 g. We then had to correct for the absence of tails and pelts on the mink, since we presumed that the body

weight in the model would have included these. The tails that we removed from mink averaged 1% of the body weight of the mink, and Aulerich *et al.* (1999) gives the weight of a mink skin as about 17% of the body weight. Those body weights from July through pelting would have included skin and tail, so taking the inverse of 0.82 gave us the multiplying factor of 1.22 to get from our tail-less, skinned carcasses to a whole body weight of about 760g.

Several sources give daily food and water consumption rates along with body weights of mink. Sample and Suter (1999) cited Bleavin and Aulerich's (1981) value of 137 g of food per day and estimated daily intakes of 0.099 L of water, using a model by Calder and Brown (1983), for mink averaging 970 g body weight. USEPA (1995b) estimated intakes of 177 g of food (using an allometric model by Nagy 1987) and 0.081 L water per day (using Calder and Brown's 1983 model) for mink with a body weight of 800 g. For captive adult males averaging 2200 g, Aulerich *et al.* (1999) reported that they drank 0.127 L/day and daily food consumption ranged from 147 g to 275 g, depending upon the caloric content of the food and the season. Since our largest mink weighed only 1111 g, and we wanted to make our model conservative (protective of the AOC mink) but not unrealistic, we discounted Aulerich's consumption rates as too high, and chose the larger of the remaining two values for daily food and water intakes. Thus, for our model, the daily food and water consumption rates were 177 g and 0.1 L (= 100g), respectively.

To demonstrate the model, we used several BCCs for which we could find literature values for the concentrations in Lake Ontario. We chose 2,3,7,8-TCDD (dioxin), with a relatively high k_{ow} of 6.53, and lindane, with a moderately low k_{ow} of 3.73, reasoning that compounds with k_{ow} s lower than lindane would have low potential for biomagnification. We also modeled benzo(a)pyrene, dieldrin, and mercury, for which Booty *et al.* (2005) reported k_{ow} values and Lake Ontario water concentrations; they also modeled the Lake Ontario concentration of TCDD. The concentration of lindane in Lake Ontario was 0.24 ng/L in 1992 (Williams *et al.* 2001, cited by Marvin *et al.* 2004). The k_{ow} s and BAFs were taken from Sample *et al.* (1996), who assumed that all fish consumed by mink are trophic level 3 (small fish). However, Melquist *et al.* 1981 reported mink feeding on kokanee (land-locked *Oncorhynchus nerka*) after their spawning, and we have reason to believe that mink feed on piscivorous salmonids in the AOC when they are available (Haynes and Pagano, in preparation). Still, the average trophic level of 3.8 for mink in BBWMA indicates that if salmonids (trophic level 4) do contribute a significant portion of the minks' diet, they are balanced by a comparable portion of level 2 aquatic prey. Thus, we used the BAF factors provided by Sample *et al.* (1996) for prey of trophic level 3, which is slightly higher (and thus more protective than) the prey from trophic level of 2.8 implied by our results.

When our results are incorporated into the equation for exposure, the equation becomes

$$Exp = \frac{C_w [100 g + (177 g \times P_{aq} \times BAF)]}{760 g}$$

Given C_w (the concentration of the BCC in the water), P_{aq} (the aquatic proportion of the diet), and BAF (the bioaccumulation factor of the chemical of concern at trophic level 3), the results of this model are the levels of BCCs to which mink in the AOC would be exposed daily. For example, if the concentration of dieldrin in water is 1.55E-9 ng/L, and a mink's diet consists of 50% aquatic prey, the mink will be exposed to 1.02E-5 ng/g body weight per day. A 760 g mink would thus be exposed to 7.8E-3 ng, or 7.8 picograms, of dieldrin per day. In contrast, if the dieldrin concentration in water is 3.4E-09 ng/L and the mink's diet is 90% aquatic, the same mink would ingest 3.1E-2 ng, or 31 picograms, of dieldrin per day (Table 4).

Confirmation of the model awaits the results of the Pagano's and Haynes' (in preparation) analyses of BCC concentrations in mink, and will require knowing BCC concentrations in Lake Ontario water. If the sample results (Pagano and Haynes, in preparation) and model results are comparable, the model can be expanded to apply to any BCC if the concentration in the ambient water is known, along with the k_{ow} and BAF of the compound (many of which are in Sample *et al.* 1996). Comparing the results of a validated model to NOAELs (No Observed Adverse Effect Levels) or LOAELs (Lowest Observed Adverse Effect Levels) for specific compounds in mink (Wellman, in preparation) is a preferable method to assess the risk to mink exposed to Lake Ontario or other waters in the AOC because it will require only measuring BCC concentrations in water, not sacrificing mink.

SUMMARY

The first question addressed by this study was: Can stable isotope analysis be used to evaluate mink diets, at lakeshore and inland areas in and out of the AOC, in terms of trophic levels and terrestrial and aquatic food sources? Analysis of $\delta^{15}N$ allowed us to determine that mink in the study area feed on prey at trophic level 2.5 (slightly higher along the lakeshore and in the AOC than elsewhere), with the highest level (2.8) in the Lakeshore/AOC area. We were unable to use $\delta^{13}C$ values to determine % aquatic diet because we had no $\delta^{13}C$ values for carbon sources in the AOC wetlands.

The second question addressed by this study was: Can stable isotope results be used to construct a food web/bioaccumulation model for mink in the Rochester AOC that can predict body burdens of BCCs in mink in relation to their diets? Using our trophic level calculation and literature values of 50% and 90% aquatic diet, we were able to create a food web bioaccumulation model to predict the exposure of mink in the AOC to BCCs, once the BCCs' concentrations in ambient water such as Lake Ontario or Braddock Bay are known. Validation of the model awaits the results of the analyses of mink tissues for BCCs (Pagano and Haynes, in preparation).

In addition, we found that the ages of mink trapped had no effect on either body weight or body length, as they were all near or at their adult size when they were trapped. The Regional Descriptors (AOC: In vs. Out, Lakeshore vs. Inland, Wetland vs. Mixed habitat) also had no effect on the ages of mink trapped. Mean ages of trapped mink were lower in historically trapped areas. Mink less than one year old were trapped in all areas, implying reproduction in all areas including the AOC. However, indications of reproduction of mink in the AOC are not sufficient to justify delisting the wildlife reproduction impairment for the Rochester Embayment. That determination will require the comparison of BCCs in mink tissues (Pagano and Haynes, in preparation) to the NOAELs and LOAELs of the BCCs in question (Wellman, in preparation).

ACKNOWLEDGMENTS

We are very grateful to Randy Baase who for two winters spent many hours trapping mink in the worst weather, and who taught us much about mink. Thanks are also due to other trappers who contributed carcasses and taught us about mink: Matt Lochner, Dan Carroll, Dan Frey, Michael Ingham, Don Newcombe, Tom Raduns, William Schwartz, Jr., Fred Sinclair, and Chuck Tirrano; and to the landowners near the Bergen Swamp who allowed us to collect mink on their property: "Doc" Fink, Dick Sands, Mel Reber and Al Burkhart. We are grateful to Scott

Wells and Ross Abbett for dissecting the mink, and to Albert Fulton for creating Figure 1. We thank Art Kasson at COIL for doing the isotope analyses, and Matson's Lab for aging the mink teeth. Bob Gilliam of the InterLibrary Loan office of Drake Library was very helpful in acquiring requested literature. We are very grateful for Dr. Richard Aulerich's generous donation of a copy of the Handbook of Biological Data for Mink. The government personnel who graciously allowed us to work in their territories include NYS DEC: Dan Carroll, Dave Woodruff and Heidi Bogner Kennedy; NYS OPRHP: James Slusarczyk, and USFWS: Bob Lamoy and Paul Hess. Finally, we gratefully acknowledge grant C302399 from the New York State Great Lakes Protection Fund that made this study possible.

LITERATURE CITED

- Albuquerque, A.L.S. and A.A. Mozeto. 1997. C:N:P ratios and stable carbon isotope compositions as indicators of organic matter sources in a riverine wetland system (Mojí-Guaçu River, Sao Pãulo-Brazil). *Wetlands* 17(1): 1-9.
- Aulerich, R.J., D.C. Powell, and S.J. Bursian. 1999. Handbook of Biological Data for Mink. Michigan State University. East Lansing, Michigan.
- Balasse, M., A. Tresset, K. Dobney, and S.H. Ambrose. 2005. The use of isotope ratios to test for seaweed eating in sheep. *Journal of Zoology* 266:283-291.
- Booty, W.G., O. Resler, and C. McCrimmon. 2005. Mass balance modelling of priority toxic chemicals within the Great Lakes toxic chemical decision support system: RateCon model results for Lake Ontario and Lake Erie. *Environmental Modelling & Software* 20:671-688.
- Cabana, G. and J. B. Rasmussen. 1994. Modelling food chain structure and contaminant bioaccumulation using stable nitrogen isotopes. *Nature* 372:255-257.
- Chittenden, M.E., Jr. 2002. Given a significance test, how large a sample size is large enough? *Fisheries* 27(8): 25-29.
- Choi, W.J., H.-M. Ro, S.X. Chang. 2005. Carbon isotope composition of *Phragmites australis* in a constructed saline wetland. *Aquatic Botany* 82: 27-38.
- Cloern, J.E., E.A. Canuel, and D. Harris. 2002. Stable carbon and nitrogen isotope composition of aquatic and terrestrial plants of the San Francisco Bay estuarine system. *Limnology and Oceanography* 47(3): 713-729.
- Collier, K.J. and G.L. Lyon. 1991. Trophic pathways and diet of blue duck (*Hymenolaimus malacorhynchos*) on Manganuiateao River; a stable carbon isotope study. *New Zealand Journal of Marine and Freshwater Research* 25(2):181-186.
- DeNiro, M.J. and S. Epstein. 1978. Influence of diet on the distribution of carbon isotopes in animals. *Geochemica et Cosmochemica Acta* 42: 495-506.
- Doucett, R.R. 1999. Food-web relationships in Catamaran Brook, New Brunswick, as revealed by stable-isotope analysis of carbon and nitrogen. Ph.D. Thesis, Biology. University of Waterloo. Waterloo, Ontario, Canada.
- Dunstone, N. (1993). The Mink. London, T & A D Poyser Ltd.
- Eagle, T. C. and J. S. Whitman. 1987. Mink. Pages 614-625 in M. Novak, J. A. Baker, M. E. Obbard, and B. Mallock, eds. Wild furbearer management and conservation in North America. Ontario Trappers Association and Ontario Ministry of Natural Resources

- Focken, U. and K. Becker. 1998. Metabolic fractionation of stable carbon isotopes: implications of different proximate compositions for studies of the aquatic food webs using $\delta^{13}\text{C}$ data. *Oecologia* 115: 337-343.
- France, R.L. 1995. Differentiation between littoral and pelagic food webs in lakes using stable carbon isotopes. *Limnology and Oceanography* 40: 1310-1313.
- Fry, B. 1991. Stable isotope diagrams of freshwater food webs. *Ecology* 72(6): 2293-2297.
- Gerell, R. (1970). Home ranges and movements of the mink *Mustela vison* in southern Sweden. *Oikos* 21: 160-173.
- Haynes, J.M., Pagano, J.J., and Wellman, S.T. 2002. New York State Great Lakes Protection Fund Application for State Assistance—RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species.
- Heaton, T.H.E. 1999. Spatial, species, and temporal variations in the $^{13}\text{C}/^{12}\text{C}$ ratios of C_3 plants: implications for palaeodiet studies. *Journal of Archaeological Science* 26: 637-649.
- Keough, J.R., M.E. Sierszen, and C.A. Hagley. 1996. Analysis of a Lake Superior coastal food web with stable isotope techniques. *Limnology and Oceanography* 41(1): 136-146.
- Kiriluk, R.M., M.R. Servos, D.M. Whittle, G. Cabana, and J.B. Rasmussen. 1995. Using ratios of stable nitrogen and carbon isotopes to characterize the biomagnification of DDE, mirex, and PCB in a Lake Ontario pelagic food web. *Canadian Journal of Fisheries and Aquatic Science* 52: 2660-2674.
- Marvin, C., S. Painter, D. Williams, V. Richardson, R. Rossmann, and P. Van Hoof. 2004. Spatial and temporal trends in surface water and sediment contamination in the Laurentian Great Lakes. *Environmental Pollution* 129: 131-144.
- Matson, G.M. 1981. Workbook for Cementum Analysis. Matson's Laboratory, LLC. Milltown, MT. www.matsonslab.com.
- Melquist, W.E., J.S. Whitman and M.G. Hornocker. 1981. Resource partitioning and coexistence of sympatric mink and river otter populations. Pages 187-220 in J.A. Chapman and D. Pursley, eds. *Worldwide Furbearer Conference Proceedings*. Worldwide Furbearer Conference, Inc. Frostburg, MD.
- Minigawa, M. and E. Wada. 1984. Stepwise enrichment of ^{15}N along food chains: Further evidence and the relation between $\delta^{15}\text{N}$ and animal age. *Geochemica et Cosmochemica Acta* 48: 1135-1140.
- Mitchell, J.L. 1961. Mink movements and populations on a Montana river. *Journal of Wildlife Management* 25(1): 48-54.
- Peterson, B.J. and B. Fry. 1987. Stable isotopes in ecosystem studies. *Annual Review of Ecology and Systematics* 18: 293-320.
- Peterson, B.J., R.W. Howarth, and R.H. Garrett. 1985. Multiple stable isotopes used to trace the flow of organic matter in estuarine food webs. *Science* 227: 1361-1363.
- RAP. 1993. Stage I Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- RAP. 1997. Stage II Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- Sample, B.E., D.M. Opresko, and G.W. Suter II. 1996. Toxicological Benchmarks for Wildlife: 1996 Revision. Oak Ridge National Laboratory, Oak Ridge, TN.
- Sample, B.E. and G. W. Suter II. 1999. Ecological risk assessment in a large river-reservoir: 4. Piscivorous wildlife. *Environmental Toxicology and Chemistry* 18(4): 610-620.

- U.S. Environmental Protection Agency. 1993. Wildlife Exposure Factors Handbook: Appendix: Literature Review Database. Office of Research and Development, USEPA. Washington, D.C.
- U.S. Environmental Protection Agency. 1995a. Great Lakes Water Quality Initiative Technical Support Document for Wildlife Criteria. Office of Science and Technology, Office of Water, USEPA. Washington, D.C.
- U.S. Environmental Protection Agency. 1995b. Great Lakes Water Quality Criteria Documents for for the Protection of Wildlife. Office of Science and Technology, Office of Water, USEPA. Washington, D.C.
- Vander Zanden, M.J., and J.B. Rasmussen. 1999. Primary consumer $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ and the trophic position of aquatic consumers. *Ecology* 80(4):1395-1404.
- Wellman, S.T., and Haynes, J.M. 2005. Monitoring mink populations using video traps. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.

TABLES

Table 1. Table of Age versus Regional Descriptors. The General Linear Model (GLM) was run with and without Trapping Pressure (TP) as a covariate. With TP as a covariate, the Landscape Regional Descriptor was dropped due to empty cells. Significant effects are in bold. The power estimates are conservative, since Minitab’s power calculator is not built to deal with GLMs (see Methods). *Power is not relevant since effect is significant (Chittenden 2002).

Regional Descriptor	N	Mean Age (SE)	P-value w/o TP	P-value w/ TP	Power
AOC: In vs. Out	29		0.404	0.660	0.171
AOC: In	15	2.2 (0.37)			
AOC: Out	14	1.8 (0.36)			
Lakeshore vs. Inland			0.386	0.634	0.074
Inland	17	1.9 (0.37)			
Lakeshore	12	2.2 (0.35)			
Landscape: Wetlands vs. Mixed			0.304	N/A	0.806
Mixed Habitat	19	2.4 (0.33)			
Wetlands Complex	10	1.2 (0.27)			
Trapping Pressure			N/A	0.017	*
None Previous	15	2.6 (0.37)			
Historically Trapped	14	1.4 (0.26)			

Table 2. Table of $\delta^{15}\text{N}$ versus Regional Descriptors showing selected descriptive statistics, and the results of the GLM with estimated power. Significant effects are in bold. The power estimates are conservative, since Minitab's power calculator is not built to deal with GLMs (see Methods). *Power is not relevant since the effect is significant (Chittenden 2002).

Area	N	Mean $\delta^{15}\text{N}$ (SE) (‰)	Min δN (‰)	Max δN (‰)
BBWMA	10	13.2 (0.54)	11.09	16.88
AOC : In/Inland	11	11.8 (0.48)	9.20	14.55
INWR	5	11.2 (0.18)	10.49	11.56
LOSPW	10	12.2 (0.42)	10.45	14.28
TUGHL	5	9.8 (0.12)	9.40	10.14
Entire Study	41	11.9 (0.23)	9.20	16.88
Regional Descriptor	N	Mean $\delta^{15}\text{N}$ (SE) (‰)	P-value	Power
AOC: In vs. Out	41		0.025	*
AOC: In	21	12.4 (0.4)		
AOC: Out	20	11.3 (0.3)		
Lakeshore vs. Inland			0.002	*
Inland	21	11.2 (0.3)		
Lakeshore	20	12.7 (0.3)		
Landscape: Wetlands vs. Mixed			0.613	0.711
Mixed Habitat	26	11.6 (0.3)		
Wetlands Complex	15	12.5 (0.4)		

Table 3. Table of $\delta^{13}\text{C}$ versus Regional Descriptors showing selected descriptive statistics, and the results of the GLM with estimated power. The power estimates are conservative, since Minitab's power calculator is not built to deal with GLMs (see Methods).

Area	N	Mean $\delta^{13}\text{C}$ (SE) (‰)	Min δC (‰)	Max δC (‰)
BBWMA	10	-25.0 (0.7)	-28.10	-19.98
AOC : In/Inland	11	-25.3 (0.2)	-26.56	-24.36
INWR	5	-25.7 (1.0)	-28.29	-23.14
LOSPW	10	-25.3 (0.4)	-27.03	-23.15
TUGHL	5	-26.2 (0.3)	-26.95	-25.31
Entire Study	41	-25.4 (1.5)	-28.29	-19.98
Regional Descriptor	N	Mean $\delta^{13}\text{C}$ (SE) (‰)	P-value	Power
AOC: In vs. Out	41		0.333	0.292
AOC: In	21	-25.1 (0.3)		
AOC: Out	20	-25.6 (0.3)		
Lakeshore vs. Inland			0.314	0.231
Inland	21	-25.6 (0.3)		
Lakeshore	20	-25.1 (0.4)		
Landscape: Wetlands vs. Mixed			0.963	0.091
Mixed Habitat	26	-25.5 (0.2)		
Wetlands Complex	15	-25.2 (0.6)		

Table 4. Predicted daily exposure levels of mink in the AOC, based on literature values for BCC concentrations in Lake Ontario. Observed values indicated by (o); estimated values by (e). BAF values are from Sample *et al.* (1996). Constants used: water intake = 0.1 L/day, food intake = 177 g/day, body weight of mink = 760 g. The concentration of 2,3,7,8-TCDD was estimated by Booty *et al.* (2005), as the compound was not detectable in their study. The concentrations for dieldrin and mercury are the minimum and maximum values observed by Booty *et al.* (2005), while the concentration for B(a)P is the maximum they observed (minimum was zero). The concentration of lindane was reported by Williams *et al.* (2001), cited by Marvin *et al.* (2004).

Compound	K(ow)	BAF: Prey Trophic Level 3	Concentration Cw (ng/L)	Daily Exposure (ng/g bw)	
				Diet 50% Aquatic	Diet 90% Aquatic
2,3,7,8-TCDD (e)	6.53	172100	1.8E-7	3.61E-03	6.49E-03
Benzo(a)pyrene (o)	6.11	293831	1.48E-09	5.06E-05	9.12E-05
Dieldrin (o)	5.37	56523	1.55E-09	1.02E-05	1.84E-05
		56523	3.40E-09	2.24E-05	4.03E-05
Lindane (o)	3.73	454	2.4E-10	1.27E-08	2.28E-08
Mercury (o)	N/A	27900	2.60E-09	8.45E-06	1.52E-05
		27900	2.50E-08	8.12E-05	1.46E-04

FIGURES

Figure 1. Map showing the four regions referred to in the study. AOC/Lakeshore is Braddock Bay WMA, AOC/Inland is at least 3 km from Lake Ontario, Out of AOC/Lakeshore is the Lake Ontario State Parkway west of Rte.19, and Out of AOC/Inland is Iroquois NWR and the Tug Hill Plateau (not shown). RELO is the Rochester Embayment of Lake Ontario. (Map by Albert Fulton 2005.)

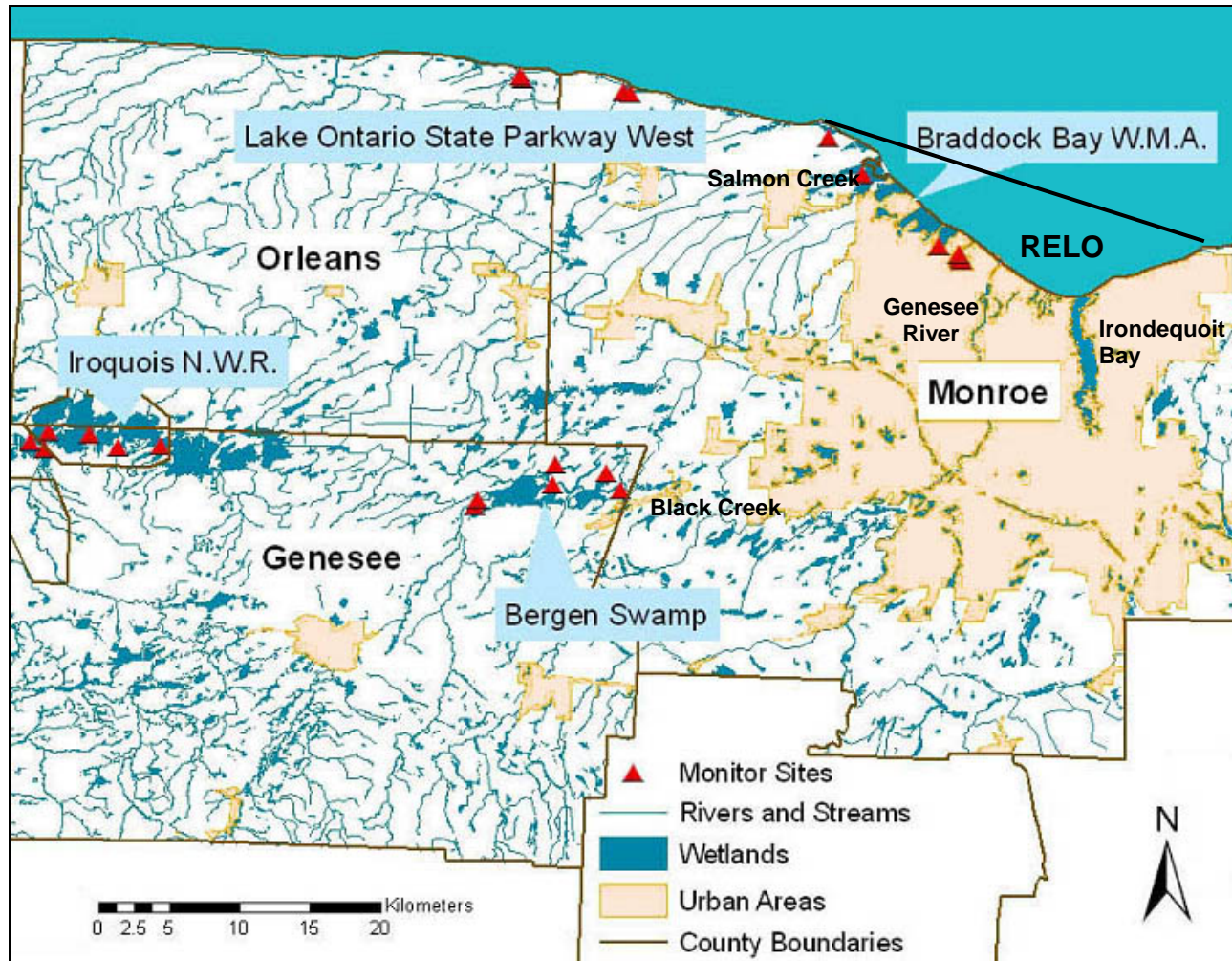


Figure 2. Ages of mink trapped.

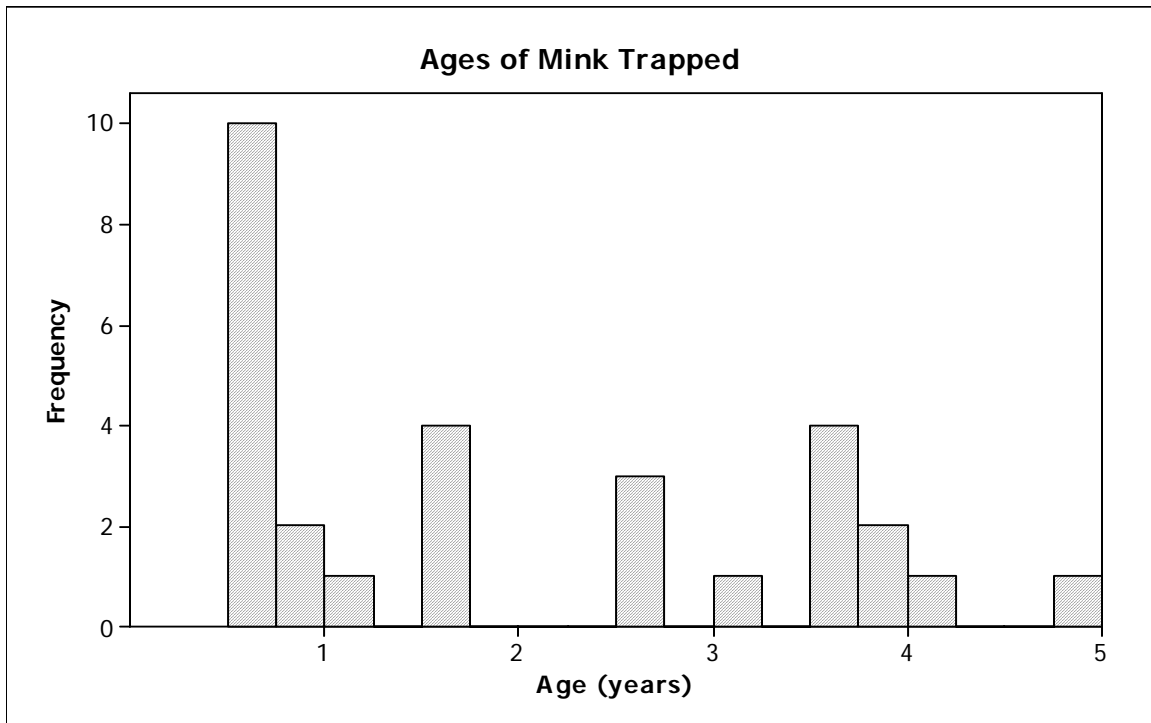


Figure 3. Plot of individual mink ages in each region.

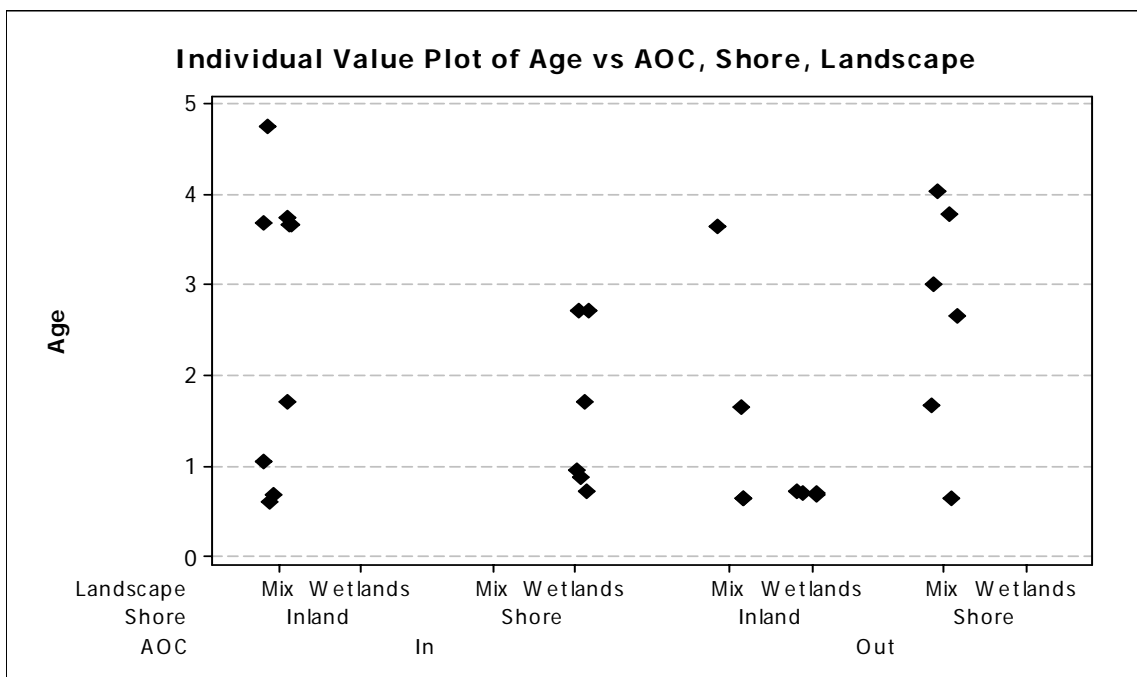
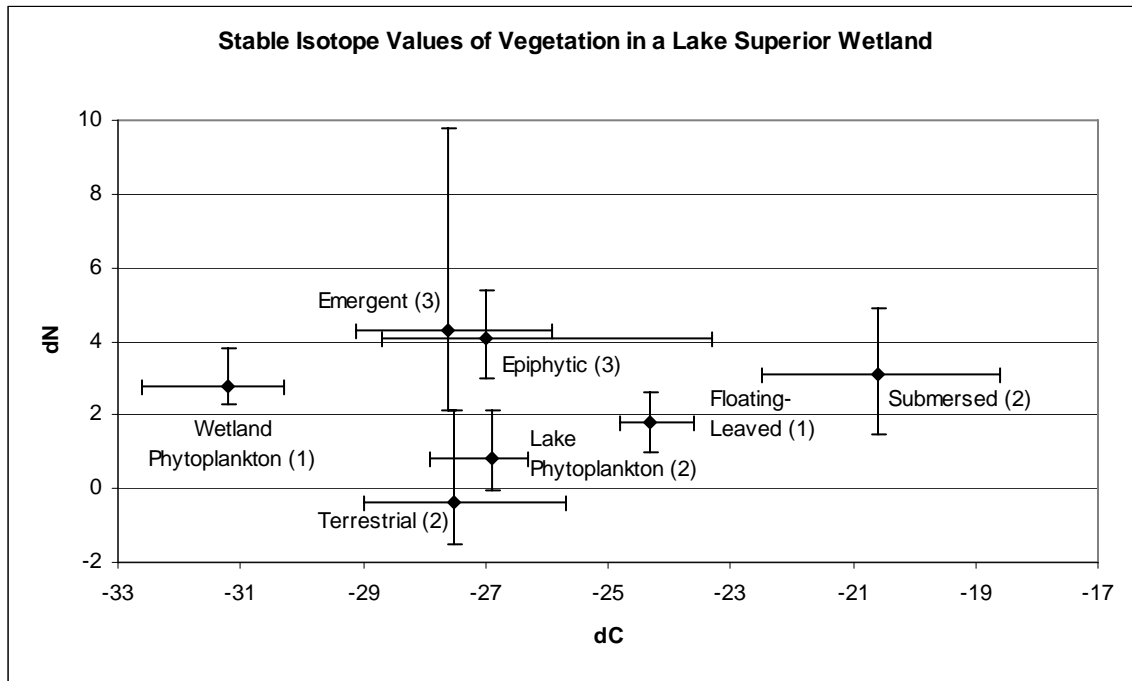


Figure 4. Stable isotope values of carbon sources in a Lake Superior wetland. (Constructed using values from Keough *et al.* 1996; numbers in parentheses are the number of species or types in each category).



APPENDICES

Appendix A: Mink age, length and weight relationships.

Figure A1: Body length versus age of mink, showing no correlation ($R^2 = 0.007$, $P = 0.655$). Tails removed for consistency (see Methods).

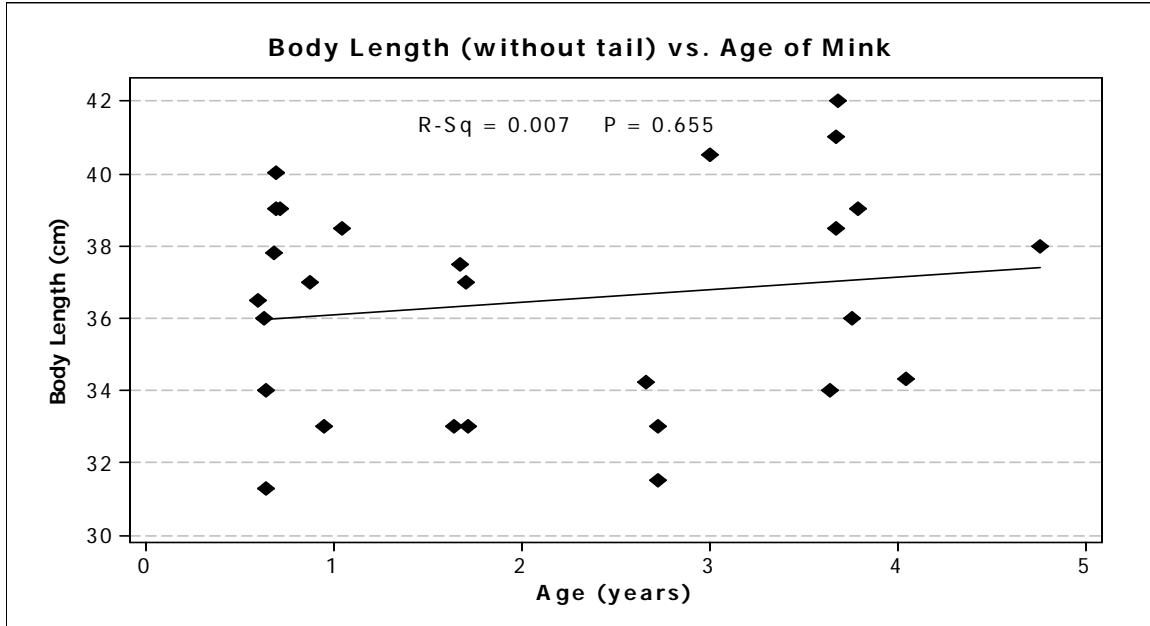
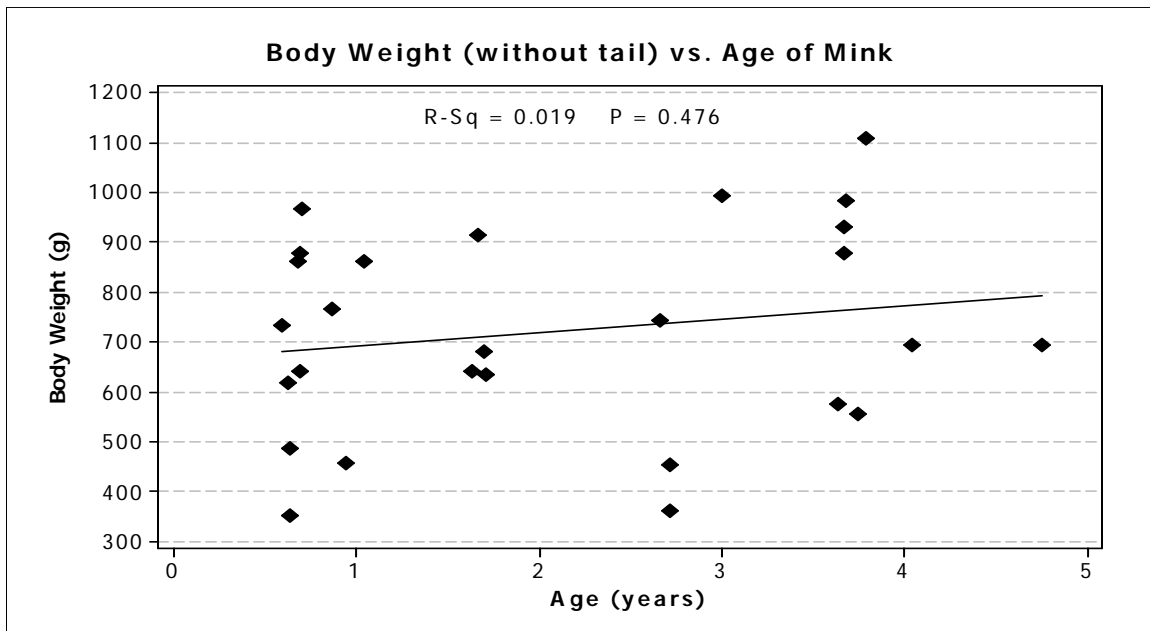


Figure A2. Body weight versus age of mink, showing no correlation ($R^2 = 0.019$, $P = 0.476$). Tails removed for consistency (see Methods).



Appendix B: Age versus Regional Descriptors

Table B1: Descriptive statistics for Age versus Regional Descriptors

Descriptive Statistics: AOC: In vs. Out, One-way ANOVA P = 0.442

Variable	AOC	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median
Age	In	15	6	2.214	0.367	1.420	0.600	0.870	1.710
	Out	14	6	1.794	0.360	1.346	0.630	0.670	1.175

Variable	AOC	Q3	Maximum
Age	In	3.670	4.750
	Out	3.160	4.040

Descriptive Statistics: Inland vs. Lakeshore, One-way ANOVA P = 0.722

Variable	Shore	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median
Age	Inland	17	4	1.933	0.369	1.522	0.600	0.675	1.040
	Shore	12	8	2.123	0.346	1.197	0.630	0.890	2.185

Variable	Shore	Q3	Maximum
Age	Inland	3.670	4.750
	Shore	2.930	4.040

Descriptive Statistics: Landscape: Wetlands vs. Mix, One-way ANOVA P = 0.027

Variable	Landscape	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Age	Mix	19	7	2.415	0.332	1.449	0.600	0.670
	Wetlands	10	5	1.245	0.265	0.837	0.680	0.690

Variable	Landscape	Median	Q3	Maximum
Age	Mix	2.660	3.680	4.750
	Wetlands	0.790	1.963	2.720

Descriptive Statistics: Trapping History, One-way ANOVA P = 0.018

Variable	Trap	N	N*	Mean	SE Mean	StDev	Minimum	Q1
Age	Pressure 0	15	6	2.621	0.370	1.433	0.600	1.040
	1	14	6	1.358	0.265	0.990	0.640	0.688

Variable	Trap	Median	Q3	Maximum
Age	Pressure 0	3.000	3.750	4.750
	1	0.790	1.963	3.640

Table B2: General Linear Model: Age versus AOC, Shore, Landscape (without Trapping Pressure as a covariate)

Factor	Type	Levels	Values
AOC	fixed	2	AOC: In, AOC: Out
Lakeshore	fixed	2	Inland, Lakeshore
Landscape	fixed	2	Mix, Wetlands

Analysis of Variance for Age, using adjusted SS for Tests

Source	DF	Seq SS	Adj SS	Adj MS	F	P
AOC	1	1.276	1.176	1.176	0.72	0.404
Lakeshore	1	0.287	1.267	1.267	0.78	0.386
AOC*Lakeshore	1	10.684	0.645	0.645	0.40	0.535
Landscape	1	1.796	1.796	1.796	1.10	0.304
Error	24	39.029	39.029	1.626		
Total	28	53.071				

S = 1.27522 R-Sq = 26.46% R-Sq(adj) = 14.20%

Figure B2a: Main effects plot.

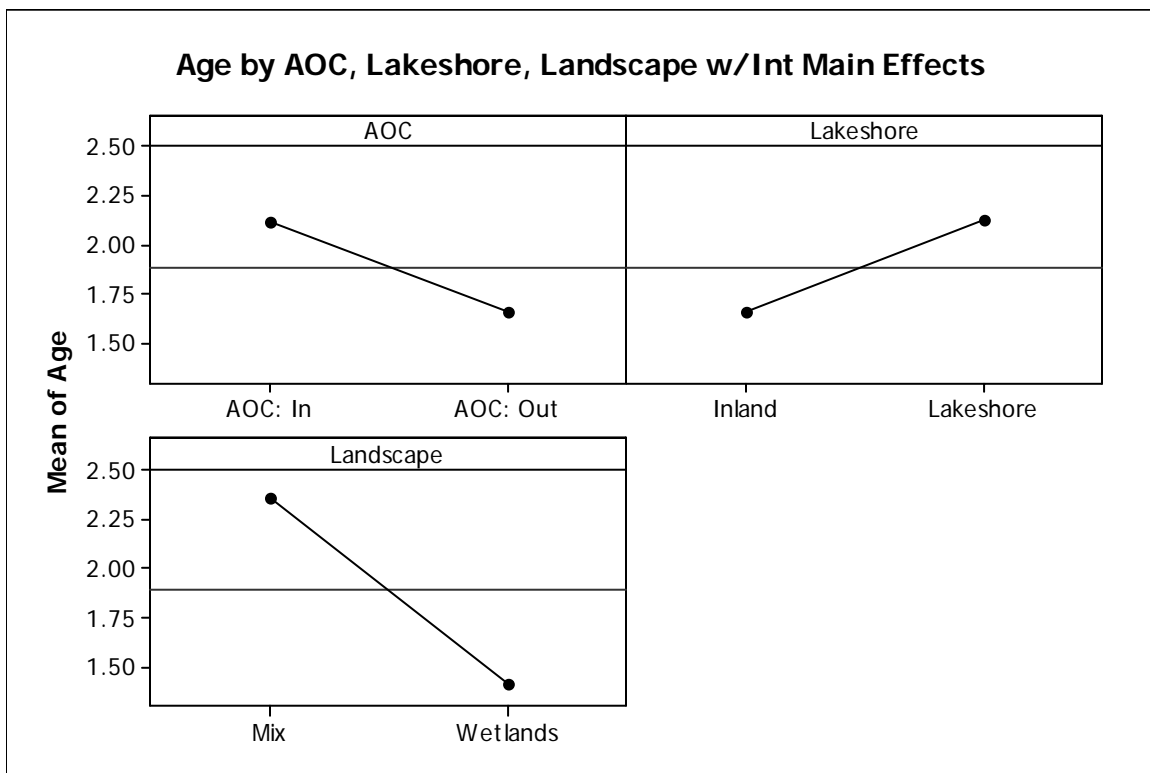


Figure B2b: Interactions plot.

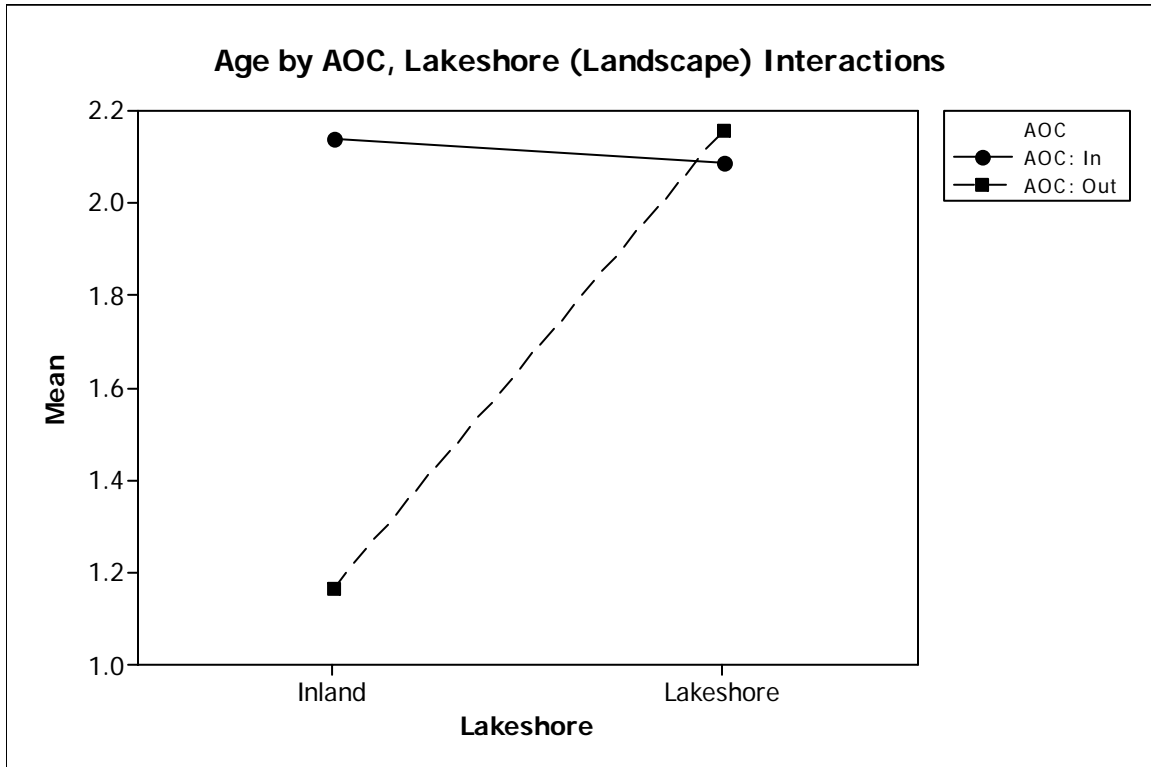


Table B3: General Linear Model: Age versus AOC, Lakeshore with Trapping Pressure as covariate.

Factor	Type	Levels	Values
AOC	fixed	2	AOC: In, AOC: Out
Lakeshore	fixed	2	Inland, Lakeshore

Analysis of Variance for Age, using Adjusted SS for Tests

Source	DF	Seq SS	Adj SS	Adj MS	F	P
Trapping	1	11.560	10.684	10.684	6.54	0.017
AOC	1	0.308	0.324	0.324	0.20	0.660
Lakeshore	1	0.379	0.379	0.379	0.23	0.634
Error	25	40.824	40.824	1.633		
Total	28	53.071				

S = 1.27788 R-Sq = 23.08% R-Sq(adj) = 13.84%

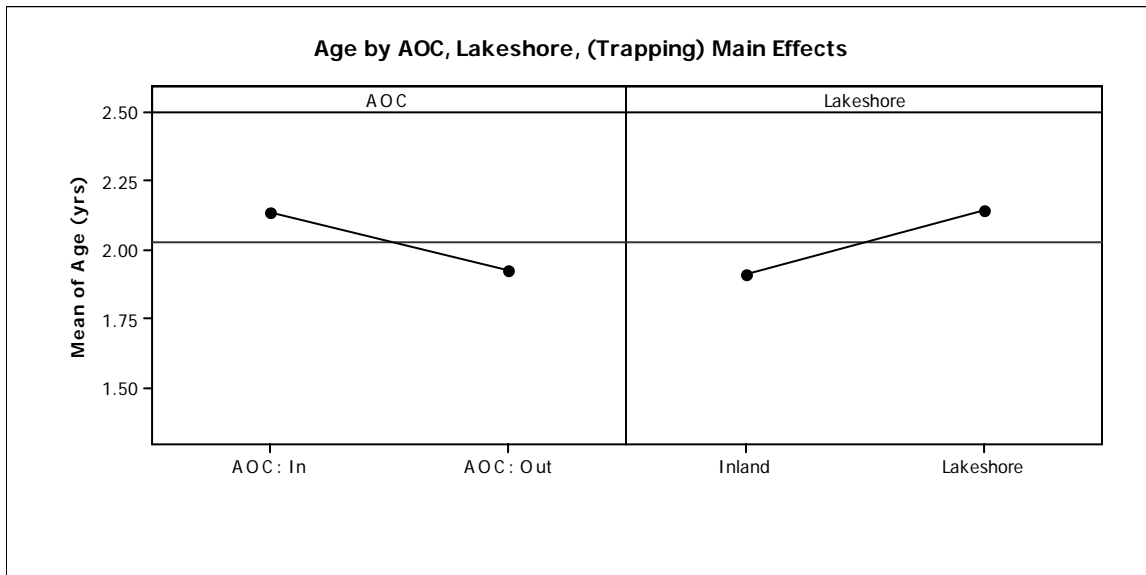
Term	Coef	SE Coef	T	P
Constant	3.8563	0.7568	5.10	0.000
Trapping	-1.2333	0.4822	-2.56	0.017

Unusual Observations for Age

Obs	Age	Fit	SE Fit	Residual	St Resid
15	3.64000	1.16625	0.45180	2.47375	2.07 R

R denotes an observation with a large standardized residual.

Figure B3a: Main effects plot.



Appendix C: Isotope data versus Age.

Appendix C1: $\delta^{15}\text{N}$ versus Age.

Figure C1. Scatterplot of $\delta^{15}\text{N}$ versus Age showing very weak correlation ($R^2 = 0.144$, $P = 0.043$).

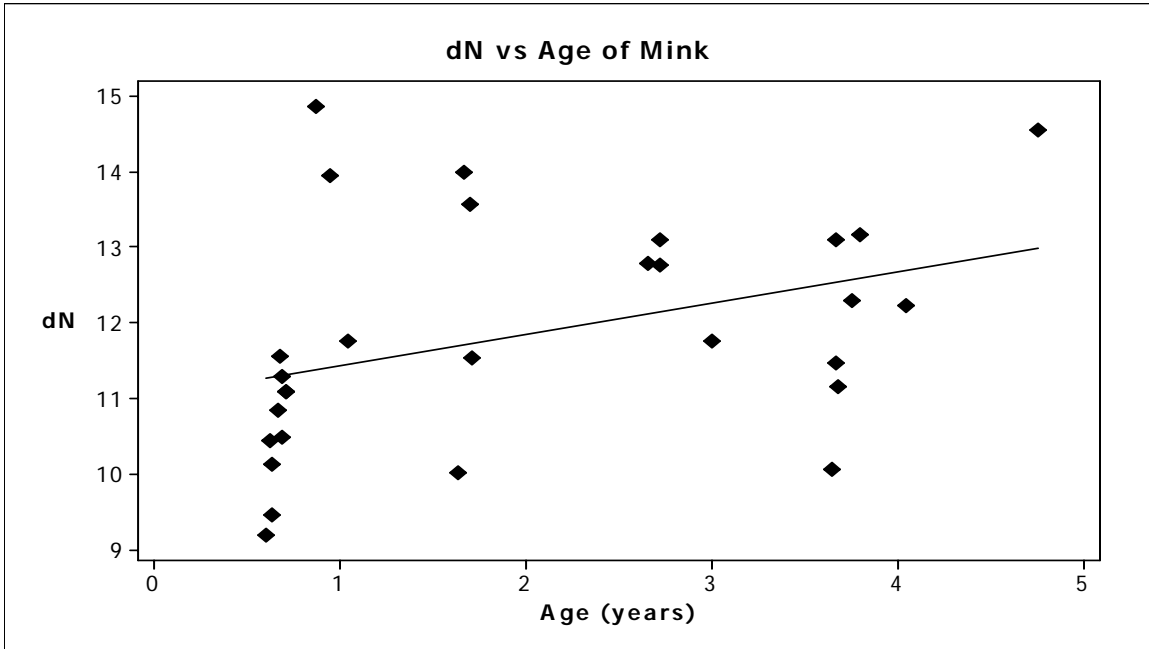


Table C1. Regression analysis: $\delta^{15}\text{N}$ versus Age (29 mink)

The regression equation is

$$\delta^{15}\text{N} = 11.0 + 0.417 \text{ Age}$$

29 cases used, 12 cases contain missing values

Predictor	Coef	SE Coef	T	P
Constant	11.0110	0.4752	23.17	0.000
Age	0.4173	0.1960	2.13	0.043

S = 1.42811 R-Sq = 14.4% R-Sq(adj) = 11.2%

Analysis of Variance

Source	DF	SS	MS	F	P
Regression	1	9.241	9.241	4.53	0.043
Residual Error	27	55.066	2.039		
Total	28	64.307			

Unusual Observations

Obs	Age	δN	Fit	SE Fit	Residual	St Resid	
36	0.87	14.850	11.374	0.347	3.476	2.51R	Mink #58, Bergen

R denotes an observation with a large standardized residual.

Appendix C2: $\delta^{13}\text{C}$ versus Age

Figure C2. Scatterplot of $\delta^{13}\text{C}$ versus Age showing no correlation ($R^2 = 0.008$, $P = 0.641$).

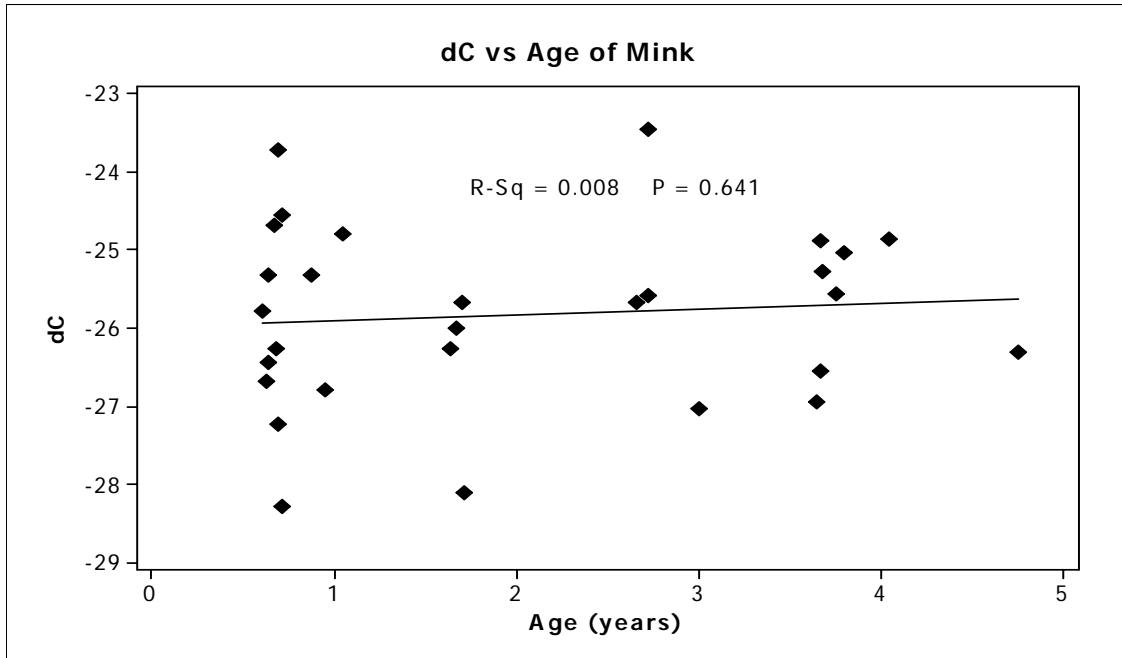


Table C2. Regression Analysis: $\delta^{13}\text{C}$ versus Age (29 mink)

The regression equation is
 $\delta^{13}\text{C} = -26.0 + 0.075 \text{ Age}$

29 cases used, 12 cases contain missing values

Predictor	Coef	SE Coef	T	P
Constant	-25.9799	0.3862	-67.27	0.000
Age	0.0752	0.1593	0.47	0.641

S = 1.16071 R-Sq = 0.8% R-Sq(adj) = 0.0%

<u>Analysis of Variance</u>					
Source	DF	SS	MS	F	P
Regression	1	0.300	0.300	0.22	0.641
Residual Error	27	36.376	1.347		
Total	28	36.676			

Unusual Observations

Obs	Age	dC	Fit	SE Fit	Residual	St Resid
3	0.71	-28.290	-25.927	0.299	-2.363	-2.11R
9	2.72	-23.450	-25.775	0.243	2.325	2.05R

R denotes an observation with a large standardized residual.

Appendix D: Isotope Data versus Regional Descriptors
Appendix D1: $\delta^{15}\text{N}$ vs. Regional Descriptors

Table D1a. Descriptive statistics: $\delta^{15}\text{N}$ vs. Regional Factors

Descriptive Statistics: AOC: In vs. Out, One-way ANOVA P = 0.035

Variable	AOC	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median	Q3
$\delta^{15}\text{N}$	In	21	0	12.429	0.382	1.753	9.200	11.305	12.420	13.325
	Out	20	0	11.349	0.310	1.385	9.400	10.218	11.310	12.110

Variable	AOC	Maximum
$\delta^{15}\text{N}$	In	16.880
	Out	14.280

Descriptive Statistics: Inland vs. Lakeshore, One-way ANOVA P = 0.002

Variable	Shore	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median
$\delta^{15}\text{N}$	Inland	21	0	11.158	0.307	1.409	9.200	10.045	11.150
	Shore	20	0	12.684	0.349	1.561	10.450	11.563	12.490

Variable	Shore	Q3	Maximum
$\delta^{15}\text{N}$	Inland	11.835	14.550
	Shore	13.755	16.880

Descriptive Statistics: Landscape: Wetlands vs. Mix, One-way ANOVA P = 0.085

Variable	Landscape	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median
$\delta^{15}\text{N}$	Mix	26	0	11.563	0.308	1.573	9.200	10.120	11.545
	Wetlands	15	0	12.489	0.436	1.688	10.490	11.280	12.420

Variable	Landscape	Q3	Maximum
$\delta^{15}\text{N}$	Mix	12.858	14.550
	Wetlands	13.090	16.880

Descriptive Statistics: BBWMA (AOC: In, Lakeshore, Wetlands)

Variable	Landscape	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median
$\delta^{15}\text{N}$	Wetlands	10	0	13.157	0.536	1.694	11.090	12.200	12.650

Variable	Landscape	Q3	Maximum
$\delta^{15}\text{N}$	Wetlands	14.175	16.880

Table D1b. General Linear Model: $\delta^{15}\text{N}$ versus AOC, Shore, Landscape

Factor	Type	Levels	Values
AOC	fixed	2	In, Out
Shore	fixed	2	Inland, Shore
Landscape	fixed	2	Mix, Wetlands

Analysis of Variance for $\delta^{15}\text{N}$, using Adjusted SS for Tests

Source	DF	Seq SS	Adj SS	Adj MS	F	P	
AOC	1	11.950	10.764	10.764	5.48	0.025	Significant effect.
Shore	1	24.692	20.780	20.780	10.57	0.002	Significant effect.
Landscape	1	0.510	0.510	0.510	0.26	0.613	No effect.
Error	37	72.714	72.714	1.965			
Total	40	109.866					

S = 1.40187 R-Sq = 33.82% R-Sq(adj) = 28.45%

Unusual Observations for dN

Obs	dN	Fit	SE Fit	Residual	St Resid	
7	16.8800	13.3379	0.4258	3.5421	2.65 R	(Mink #17, BBWMA)
39	14.5500	11.6028	0.4075	2.9472	2.20 R	(Mink #61, AOC: In/Inland)

R denotes an observation with a large standardized residual.

Appendix D2: $\delta^{13}\text{C}$ vs. Regional Factors

Table D2a. Descriptive statistics: $\delta^{13}\text{C}$ vs. Regional Factors

Descriptive Statistics: AOC: In vs. Out, One-way ANOVA P = 0.330

Variable	AOC	N	N*	Mean	SE Mean	StDev	Minimum	Q1	Median
$\delta^{13}\text{C}$	In	21	0	-25.155	0.344	1.578	-28.100	-25.990	-25.320
	Out	20	0	-25.617	0.315	1.409	-28.290	-26.615	-25.810
Variable	AOC			Q3	Maximum				
$\delta^{13}\text{C}$	In			-24.555	-19.890				
	Out			-24.903	-23.140				

Descriptive Statistics: Inland vs. Lakeshore, One-way ANOVA P = 0.304

Variable	Shore	N	N*	Mean	SE Mean	StDev	Minimum	Q1
$\delta^{13}\text{C}$	Inland	21	0	-25.618	0.264	1.211	-28.290	-26.375
	Shore	20	0	-25.131	0.391	1.747	-28.100	-26.143

Variable	Shore	Median	Q3	Maximum
$\delta^{13}\text{C}$	Inland	-25.670	-24.740	-23.140
	Shore	-25.430	-24.455	-19.890

Descriptive Statistics: Landscape: Wetlands vs. Mix, One-way ANOVA P = 0.635

Variable	Landscape	N	N*	Mean	SE Mean	StDev	Minimum	Q1
$\delta^{13}\text{C}$	Mix	26	0	-25.466	0.190	0.971	-27.030	-26.278
	Wetlands	15	0	-25.231	0.559	2.166	-28.290	-26.780

Variable	Landscape	Median	Q3	Maximum
$\delta^{13}\text{C}$	Mix	-25.610	-24.843	-23.150
	Wetlands	-25.540	-23.710	-19.890

Descriptive Statistics: In AOC, Lakeshore (BBWMA)

Variable	Landscape	N	N*	Mean	SE Mean	StDev	Minimum	Q1
$\delta^{13}\text{C}$	Wetlands	10	0	-24.984	0.699	2.211	-28.100	-26.338

Variable	Landscape	Median	Q3	Maximum
$\delta^{13}\text{C}$	Wetlands	-25.430	-24.178	-19.890

Table D2a. General Linear Model: $\delta^{13}\text{C}$ versus AOC, Shore, Landscape

Factor	Type	Levels	Values
AOC	fixed	2	In, Out
Shore	fixed	2	Inland, Shore
Landscape	fixed	2	Mix, Wetlands

Analysis of Variance for $\delta^{13}\text{C}$, using Adjusted SS for Tests

Source	DF	Seq SS	Adj SS	Adj MS	F	P
AOC	1	2.189	2.208	2.208	0.96	0.333
Shore	1	2.538	2.393	2.393	1.04	0.314
Landscape	1	0.005	0.005	0.005	0.00	0.963
Error	37	85.013	85.013	2.298		
Total	40	89.744				

S = 1.51580 R-Sq = 5.27% R-Sq(adj) = 0.00%

Unusual Observations for dC

Obs	$\delta^{13}\text{C}$	Fit	SE Fit	Residual	St Resid	
7	-19.8900	-24.9033	0.4604	5.0133	3.47	R Mink #17, BBWMA
18	-28.1000	-24.9033	0.4604	-3.1967	-2.21	R Mink #38, BBWMA

R denotes an observation with a large standardized residual.

Appendix 3

Levels of Bioaccumulative Chemicals of Concern in Mink In and Out of the Rochester Embayment Area of Concern and Along and Inland from the Shore of Lake Ontario

James J. Pagano¹ and James M. Haynes²

¹Environmental Research Center and Department of Chemistry
State University of New York
College at Oswego
Oswego, NY 13126

²Department of Environmental Science and Biology
State University of New York
College at Brockport
350 New Campus Drive
Brockport, NY 14420-2973

March 2007

OVERVIEW

This report is the fourth of four resulting from project C302399 funded by the New York Great Lakes Protection Fund in 2004 to address use impairments related to water quality identified in the Remedial Action Plan for the Rochester Embayment of Lake Ontario (RELO RAP). It deals with the concentrations of bioaccumulative chemicals of concern (BCCs) in mink (*Mustela vison*) in and out of the Rochester Embayment Area of Concern (AOC) and along and inland from the shore of Lake Ontario. The previous reports addressed 1) development and use of videocapture (MustelaVision) systems that established the presence and reproduction of mink in and out of the AOC (Wellman and Haynes 2006a); 2) ages, sizes and trophic positions (stable isotope analysis) of mink in the study areas and a predictive model for bioaccumulation of BCCs by mink (Wellman and Haynes 2006b); and 3) literature review of the effects of BCCs on mink and testing of the bioaccumulation model (Wellman and Haynes 2007) against actual tissue concentrations found in this part of the study. Because mink are the most sensitive species to BCCs known, the results of this four-part project (to be integrated and summarized in a final report in early 2007) will determine whether or not delisting can be recommended for the fish and wildlife population, reproduction and deformities use impairments identified in the RELO AOC.

Table of Contents

INTRODUCTION	90
MATERIALS AND METHODS	90
Collection	90
Processing	91
Handling and Shipping	91
Analytical Procedures: PCBs, Organochlorine Pesticides and PBDEs	91
Sample Extraction and Clean-Up	91
Chemical Analysis	91
Laboratory Quality Assurance/Quality Control	93
Analytical Procedures: Dioxins/Furans and Mercury	93
Data Analysis	94
RESULTS	94
Relationships of BCC Concentrations	94
BCC Concentrations vs. Regional Descriptors	94
Total Mercury in Brain Tissue	94
BCC Concentrations in Adipose Tissue	94
BCC Concentrations in Liver Tissue	95
DISCUSSION	96
Statistical and Other Issues	96
Outliers	96
Statistical Power	97
Lack of Co-planar PCB Concentration Data	97
ACKNOWLEDGMENTS	98
LITERATURE CITED	98
TABLES	100
Table 1. Primary column: PCB congeners analyzed with Agilent 25m UltraII	100
Table 2. Confirmation column: PCB congeners analyzed with Agilent 60m DB-XLB	101
Table 3. Primary analytical column: organochlorine pesticide components and polybrominated diphenyl ether congeners analyzed with 60m DB-XLB	102
Table 4. GLPF mink project surrogate recoveries (SR) and surrogate spike checks for various sample matrices.	103
Table 5. Correlations of concentrations of selected bioaccumulative chemicals of concern (BCC) in adipose and brain (total Hg only) tissue of mink	104
Table 6. Correlations of concentrations of selected bioaccumulative chemicals of concern (BCC) in liver and brain (Total Hg only) tissue of mink.	105
Table 7. Total mercury concentrations (ng/g) in the brains of mink	106
Table 8. Concentrations of selected BCCs in mink adipose tissue	107
Table 9. Concentrations of selected BCCs in mink liver tissue	108
FIGURE 1. Map showing the four regions referred to in the study.	110

APPENDICES111

Appendix A. Chemicals detected in this study. Names in bold were analyzed statistically and are discussed in the text. Raw data for all chemical are in Appendix B (CD).....111

Appendix B. Separate spreadsheet on a CD with all BCC data collected in this study.....112

Levels of Bioaccumulative Chemicals of Concern in Mink In and Out of the Rochester Embayment Area of Concern and Along and Inland from the Shore of Lake Ontario

INTRODUCTION

In the 1980s the binational (Canada, U.S.) International Joint Commission (IJC) began the process of creating and implementing remedial action plans (RAPs) in 43 contaminated areas of concern (AOCs) throughout the Great Lakes Basin. The IJC established 14 “use impairments” that could cause a local area to be “listed” as an AOC, including “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems.” In the Rochester AOC, both uses were defined as impaired because “very few” mink were then being trapped or observed within 2 miles of the lake (RAP 1993, 1997). This study was part of a project (Haynes et al. 2002) to determine if populations of mink on the shore of the Rochester Embayment of Lake Ontario (RELO) are negatively impacted by bioaccumulative chemicals of concern (BCCs) and, if so, whether the BCCs are originating in the embayment watershed or elsewhere.

The RELO AOC includes the Embayment, a 35 square mile portion of Lake Ontario south of a line between Bogus Point in the town of Parma and Nine Mile Point in the town of Webster (both in Monroe County, New York); adjacent wetlands and bays; and the six mile reach of the Genesee River, from the Lower Falls to the mouth at Lake Ontario. The RAP also includes the subwatersheds of Salmon Creek, the Genesee River and Irondequoit Creek (RAP 1993, 1997; **Figure 1**).

The question addressed by this portion of the study was: What are the current levels of BCCs in lakeshore and inland populations of mink in and out of the AOC, and how do the levels compare between the four regions? These data are needed for comparison to levels of BCCs known to affect mink reproduction (Wellman and Haynes 2007) in order to determine if mink in the RELO AOC are potentially suffering from the “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems” listed as use impairments in the RAP (1993, 1997).

MATERIALS AND METHODS

Specimen Collection, Processing and Handling **Collection**

Mink carcasses were collected from trappers (after skinning) in five areas. We divided the study area into four Regions: Inland/AOC, Lakeshore/AOC, Inland/Out of AOC, and Lakeshore/Out of AOC. Both Lakeshore regions were identical to those defined by Wellman and Haynes (2006a, 2006b)—Lakeshore/AOC was the Braddock Bay Wildlife Management Area (BBWMA), and Lakeshore/Out of AOC was along the Lake Ontario State Parkway west of Route 19 (LOSPW). Inland/AOC included any animals taken in the AOC watershed more than 5 km from the lakeshore (primarily from areas near the Bergen Swamp), and Inland/Out of AOC included animals taken from the Tug Hill Plateau and the Iroquois National Wildlife Refuge (to provide two presumably “clean” control areas). For the purpose of analysis, we described each collection area using the Regional Descriptors AOC: In vs. Out and Lakeshore vs. Inland.

Carcasses were put in plastic bags and frozen by the trappers as soon as possible. The trappers completed log sheets indicating the date and location of capture for each animal, as well as the trapper’s name and contact information. Carcasses were assigned specimen numbers in the

order in which they were collected, and the specimen number, date and location of capture were written on the plastic bags with a permanent marker.

Processing

We thawed the frozen mink carcasses overnight in a refrigerator before processing them. Because some trappers removed the tails when skinning the carcasses, we removed all other tails to obtain comparable measures of body weight and length. We recorded the body weight, tail-less body length, and weight of each tissue sample on a separate sheet for each mink, along with its specimen number and collection record. We placed carcasses in hexane-rinsed aluminum pans or aluminum foil for resection, and all utensils used were rinsed with hexane before each use. Tissues collected for analyses were adipose, liver, brain, testis, kidney and thigh muscle.

Handling and Shipping

Liver and adipose samples were divided into halves or thirds and shipped frozen in dry ice to Columbia Analytical Services (CAS), Inc.'s laboratory in Houston, TX for dioxin/furan analyses, and to the Environmental Research Center (ERC) at SUNY Oswego (co-PI Pagano's lab) for PCB-pesticide-PBDE analyses. If liver or adipose tissue was sufficiently large to divide into thirds, the third sample was frozen and kept at SUNY Brockport. Brain tissues for total mercury analyses were shipped to CAS's laboratory in Kelso, WA. Other tissues (testes, kidney and thigh muscle) were transferred frozen to the ERC for PCB-pesticide-PBDE analyses.

Analytical Procedures: PCBs, Organochlorine Pesticides and PBDEs

Sample Extraction and Clean-Up

The chemicals examined in this study are listed in Appendix A. All tissue samples were extracted for gas chromatographic analysis after methods developed at the SUNY Oswego ERC (Pagano et al. 1999). Pre-cleaned anhydrous sodium sulfate (approximately 10 times sample weight) was added, and the sample extracted three times each with 50 mL hexane using a Brinkman Polytron homogenizer (Model PT 10/35) with small generator (PTA-10S). After each extraction, the hexane extracts were transferred into a volumetric flask and brought to volume. Lipid analysis was conducted by gravimetric procedures utilizing an aliquot (subsample) of the extracted sample. The remaining sample was used for congener-specific PCB, OC pesticide and PBDE analyses. Sample cleanup for OC pesticides followed USEPA Method 3640A (1997) using a Waters Gel Permeation Chromatography (GPC) system (binary pump, Envirolgel column, UV detector and fraction collector) followed by specialized silica gel column for separation of PCBs/OCs/PBDEs from other interferences. The analytical methods used to separate PCBs, OCs and PBDEs were based on methods and standard operating procedures (SOPs) developed at the SUNY Oswego ERC and adapted from Method 3630C-USEPA (1997), Basu (1995a,b), and Harlin and Surratt (1995). In general, silica gel adsorption column cleanup utilized 5.5 grams of 4% deactivated silica gel (100-200 mesh) placed in a 10.5x250 mm chromatography column (VWR-Labglass Wilmad) with an upper layer (0.5 g) of anhydrous sodium sulfate. The sample extract was added to the silica gel column and sequentially eluted with hexane and DCM into PCB (F1) and OC/PBDE (F2) fractions, which were concentrated to 1 mL with a Kuderna-Danish (KD) apparatus using a three ball Snyder Column on a steam bath for gas chromatographic analysis.

Chemical Analysis

Congener-specific PCB, hexachlorobenzene, p-p' DDE, and mirex analyses were conducted based on capillary column procedures previously described (Pagano et al. 1995,

Pagano et al. 1998, Pagano et al. 1999, Chiarenzelli et al. 2001). Briefly, analytical instruments were recalibrated every five samples, with a system blank, instrument blank, and mid-level calibration check solution analyzed during each analytical run. A Hewlett-Packard (HP) Model 5890II GC with an electron capture detector (ECD - Ni⁶³) and autosampler was used for primary data acquisition. The capillary column utilized was a HP Ultra II, 25 meter with 0.22 mm id and 0.33 μ m film thickness. The calibration standard used was a 1:1:1:1 mixture of Aroclors 1221, 1016, 1254, and 1260 each at 200 pg/uL, hexachlorobenzene (HCB) at 5 pg/uL, and p-p' DDE (dichlorodipenyldichloroethylene) and Mirex each at 10 pg/uL (Custom Mixed Fraction #3, AccuStandard, Inc.), which allowed for the analysis of 99 chromatographic zones of 132 congeners/co-eluters (Table 1). PCB analyses were confirmed with a HP Model 5890 II gas chromatograph with an electron capture detector (Ni⁶³) and autosampler using a 60 meter DB-XLB capillary column with 0.25 mm id and 0.25 μ m film thickness. The calibration standard used was a 1:1:1:1:1 mixture of congener mixture sets (C-CSQ-SET 1-5; 10 pg/uL per individual congener, AccuStandard, Inc., New Haven, CT) based on the work of George Frame and co-workers (1996). This analytical setup allowed for analysis of 122 chromatographic zones of 155 congeners/co-eluters (Table 2).

PCB congener nomenclature is based on the arrangement and number of chlorines (1-10) substituted per biphenyl molecule. Congener determination, assignments and accuracy of quantitation were verified for both GC-ECD analytical systems utilizing nine PCB congener mixtures (C-CSQ-SET; AccuStandard, Inc., New Haven, CT) (Frame et al. 1996). Chromatographic data were collected and processed by use of the HP ChemStation software and Microsoft Excel spreadsheet procedures developed at the SUNY Oswego Environmental Research Center (Pagano et al. 1995). The HP software system generated the identity and amount of each PCB congener, confirmed by operator reprocessing of each chromatographic run.

The complex, congener-specific PCB patterns (data) found in mink adipose and liver samples were further processed to a single number representing the average overall chlorination of all congeners in the sample based either on PCB mass or moles of PCBs measured. The unit used was average number of chlorines per biphenyl (Avg Cl/BP). The manipulation of congener-specific data allows for a direct comparison of different types of tissue samples or widely different concentration levels. In addition, congener-specific data provided by other researchers can be transformed to provide a direct comparison of PCB chlorination (qualitative assessment) and concentration (quantitative assessment) between this and previous studies.

Selected organochlorine (OC) pesticides were measured based on Methods 8081A (USEPA 1996). Single instrument/column detection was used for quantitation (DB-XLB, see conditions above). The calibration standard was a 100 pg composite mixture (Single-Column Analytes Mix, M-8081-SC, AccuStandard, Inc.) of USEPA 8081A standard analytes (Table 3). Polybrominated diethyl ethers (PBDEs) were co-analyzed with the OCs on the DB-XLB column setup. The PBDE calibration standard used was an 800 pg/uL (total PBDE - 12 components) solution using the original Great Lakes Chemical Corporation (Great Lakes DE-71, CAS 32534-81-9) technical formulation. The DE-71 technical formulation mass fractions and congener identifications were confirmed with pure PBDE congener standards (BDE-MXE) purchased from Wellington Laboratories (Guelph, ON, Canada) and by mass spectrometric confirmation.

As needed, confirmation of PCBs, OCs, PBDEs, and any co-eluting contaminants were determined utilizing a HP Model 5890II GC with a Model 5971 mass selective detector (GC/EI-MS, electron impact mode) and autosampler. The GC/MS system was set-up to complement the

GC-ECD system, utilizing the same column and temperature programming. Helium was used as the carrier gas at 55 kPa. The injection port and mass selective detector interface were maintained at 270 °C and 300 °C, respectively. Selective ion monitoring (SIM) for PCB (SIM-PCB, Cl homologs 1-10) included ions (m/z) 152, 186, 188, 190, 220, 222, 224, 254, 256, 258, 290, 292, 324, 326, 360, 362, 394, 396, 428, 430, 432, 462 and 464 (Pagano et al. 1998).

Laboratory Quality Assurance/Quality Control

The QA/QC program at the SUNY Oswego ERC is based on a program developed from USEPA protocols (USEPA 1997). The program consists of replicate analyses, surrogate analyte recoveries (IUPAC 14, 30 IS (F1+F2), 65, 166, and PCT3-F2), matrix spikes/matrix spike duplicates, and method, reagent and system blanks at prescribed intervals. Surrogate recoveries and surrogate spike checks for the various mink tissues analyzed for this project are reported in Table 4. Instrument detection limits (IDL) and detector linearity were established at the start of the project by replicate analyses (N=7) of progressively smaller serial dilutions from the quantitation standards utilized for each analytical system (acceptance criteria > 10% Relative Standard Deviation, RSD). Analytical detection limits (IDL) and practical detection limits (PDL) are provided in the PDL+IDL worksheet of Appendix B (a spreadsheet on a CD that includes all chemical data collected for this project).

Limitations associated with the accurate and bias-free measurement of low-level PCBs, OCs and PBDEs are generally not due to IDL, but are attributed to the ubiquitous background contamination found in the analytical manipulations necessary to prepare and extract samples. The qualitative nature and quantitative amount of the analytical background is of critical importance when analyzing environmental samples at part per billion levels for individual congeners (Stewart et al. 2000). The SUNY Oswego ERC has developed a methodology to determine practical detection limits (PDL) based on the assessment of the extent and congener-specific distribution of background contamination using method (procedural) blanks analyzed with every sample batch. Method blanks are used to document contamination resulting from the analytical process. Method blanks encompass all Standard Operating Procedure (SOP) sample preparation and analytical manipulations within an analyte-free matrix. PDLs are calculated by multiplying the standard deviation of a series of method blanks by the associated Student t variate (usually N=7, df=6, t=3.134) to provide a known confidence interval ($t_{.99}$) for the PDL estimate. PDLs are reported in the PDL+IDL worksheet of Appendix B.

During the project, general laboratory quality assurance and silica gel method validation was determined by analysis (N=16) of National Institute of Standards and Technology (NIST) Standard Reference Material 1946 (Lake Superior Fish Tissue). Results are provided in the NIST SRM 1946 worksheet of Appendix B. Average recoveries of certified concentration values for PCBs were 92.9% and average recoveries of various organochlorine pesticides was 77.6%.

Analytical Procedures: Dioxins/Furans and Mercury

Dioxin-furan analyses were done using Method 8290A (USEPA 1998) for extractable organics in solid and chemical materials by Columbia Analytical Services, Inc., Houston, TX. Because of analytical uncertainties, because virtually all mercury in biota is methyl mercury (Me-Hg), and because the highest mercury concentrations are found in brain tissue (J. Freemyer, CAS, Houston, TX, pers. comm.), CAS (Kelso, WA) used Method 7471A (USEPA 1994) for total mercury to estimate the Me-Hg concentrations in mink brains. CAS is NELAC-certified (E87611, FAC Rule 64E-1 regulations) by the state of Florida.

Data Analysis

We used Microsoft® Excel 2000 for data management and non-statistical calculations. For statistical analyses, we used SPSS® 13.0 for Windows (SPSS Inc., Chicago, IL). We conducted regression analyses to evaluate the relationships between BCC concentrations. We computed descriptive statistics for selected chemicals in selected tissues: 1) total mercury in brain, and 2) total PCBs and dioxin-furan TEQs; average number of chlorine atoms per biphenyl; dieldrin; PBDEs, mirex and DDE in liver and adipose. We used General Linear Models (GLM, a 2-way ANOVA followed by Tukey's pair-wise comparisons and estimates of statistical power) to analyze the relationships between BCC concentrations and the Regional Descriptors—AOC: In vs. Out and Lakeshore vs. Inland. Results for a number of chemicals, detected infrequently or in minute quantities in mink tissues, are not presented here, but all analytical results and calculations are presented in the NYS GLPF MINK worksheet of Appendix B.

Excluding a statistical outlier (mink 17) and results below detection limits, 36 of 40 mink had mercury in brain tissue. Excluding a statistical outlier (mink 17), one animal with insufficient adipose tissue (mink 30), and a procedural error by CAS (addition of the wrong chemical during the preparation of two samples), 33 of 40 mink had total dioxin-furan TEFs above detection limits in adipose tissue. In liver tissue, excluding statistical outliers (17, 22), 17 of 40 mink had total dioxin-furan TEQs above detection limits. For total PCBs, average chlorine number per biphenyl, mirex, dieldrin and DDE, 37 of 41 mink were used in the statistical analyses of liver. Two mink were statistical outliers for total PCBs (17, 22), one was an outlier for mirex (49), and one had insufficient adipose tissue (29). Outliers ($> \pm 3$ SE beyond the mean) and values below detection limits (BDL) were excluded from the analyses to avoid skewing general trends high (outliers) or low (BDL \neq no chemical present in a tissue), respectively. Relationships of the highest and lowest levels of BCCs found in the tissues of mink to their lowest observed adverse effect levels (LOAEL) are addressed by Wellman and Haynes (2006a).

RESULTS

Relationships of BCC Concentrations

Correlations between chemical concentrations were high in adipose and liver tissue. Among the 28 comparisons of seven chemicals and average chlorine per biphenyl in adipose, 25 were highly significant ($P < 0.01$; $r = 0.441-0.905$), two were significant ($P < 0.05$; $r = 0.367-0.424$) and one was suggestive of significance ($P = 0.088$, dieldrin vs. average chlorine per biphenyl; $r = 0.284$) (Table 5). Among the 28 comparisons of seven chemicals and average chlorine per biphenyl in liver, 19 were highly significant ($P < 0.01$; $r = 0.430-0.899$), four were significant ($P < 0.05$; $r = 0.358-0.408$) and two were suggestive of significance ($P < 0.100$; $r = 0.274-0.278$) (Table 6).

BCC Concentrations vs. Regional Descriptors

Total Mercury in Brain Tissue

Total mercury concentrations (Table 7) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.609$; power = 0.079) but concentrations in mink from the Lake Ontario shore were higher than those of inland mink ($P = 0.032$; power = 0.587). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.835$; power = 0.055).

BCC Concentrations in Adipose Tissue

Total PCB concentrations (Table 8a) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.632$; power = 0.076) but concentrations in mink captured near Lake Ontario

were higher than those of inland mink ($P = 0.014$; power = 0.708). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.601$; power = 0.081).

The average number of chlorine atoms per biphenyl molecule (Table 8b) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.475$; power = 0.108) but chlorination in mink captured near the Lake Ontario shore was higher than that of inland mink ($P = 0.006$; power = 0.817). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.375$; power = 0.141).

Total dioxin-furan TEQs (Table 8c) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.354$; power = 0.149) but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.010$; power = 0.763). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.405$; power = 0.129).

DDE concentrations (Table 8d) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.357$; power = 0.148) but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.002$; power = 0.916). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.339$; power = 0.156).

Dieldrin concentrations (Table 8e) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.241$; power = 0.212) but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.007$; power = 0.800). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.614$; power = 0.079).

Mirex concentrations (Table 8f) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.259$; power = 0.200). The data suggested that concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.073$; power = 0.536). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.136$; power = 0.317).

Polybrominated diphenyl ether concentrations (Table 8g) did not differ (excluding outlier #17) in and out of the AOC ($P = 0.937$; power = 0.051), but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.005$; power = 0.838). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.266$; power = 0.196).

BCC Concentrations in Liver Tissue

Total PCB concentrations (Table 9a) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.960$; power = 0.050) but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.018$; power = 0.673). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.232$; power = 0.219).

The average number of chlorine atoms per biphenyl molecule (Table 9b) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.449$; power = 0.116) but chlorination in mink captured near Lake Ontario was higher than that of inland mink ($P = 0.026$; power = 0.618). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.145$; power = 0.306).

Total dioxin-furan TEQs (Table 9c) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.547$; power = 0.089) or between the Lake Ontario shoreline and inland ($P = 0.337$; power = 0.152), nor was there an interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.423$; power = 0.120).

DDE concentrations (Table 9d) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.193$; power = 0.252) but concentrations in mink captured near Lake Ontario

were higher than those of inland mink ($P = 0.005$; power = 0.836). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.667$; power = 0.071).

Dieldrin concentrations (Table 9e) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.363$; power = 0.146) but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.037$; power = 0.559). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.260$; power = 0.200).

Mirex concentrations (Table 9e) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.153$; power = 0.295). The data suggested differences between the Lake Ontario shore and inland ($P = 0.066$; power = 0.455) and an interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.068$; power = 0.449).

Polybrominated diethyl ether concentrations (Table 9g) did not differ (excluding outliers #17, 22) in and out of the AOC ($P = 0.811$; power = 0.056) but concentrations in mink captured near Lake Ontario were higher than those of inland mink ($P = 0.002$; power = 0.899). There was no interaction between the factors AOC: In vs. Out and Lakeshore vs. Inland ($P = 0.143$; power = 0.308).

DISCUSSION

The question addressed by this study was: What are the current levels of BCCs in lakeshore and inland populations of mink in and out of the AOC, and how do levels compare between the four regions? Highly consistent patterns were observed across tissues and chemicals.

- Correlations among concentrations of the seven most notable chemicals analyzed were mostly high and significant in adipose and liver tissue.
- There were no significant differences in BCC concentrations in and out of the Rochester Embayment AOC, although mean values were almost always higher (mostly by factors > 3) in the AOC.
- BCC concentrations in mink captured near the Lake Ontario shore were almost always significantly ($P < 0.05$) or suggestively ($0.05 < P < 0.1$) greater than concentrations in mink captured inland.
- After removing statistical outliers, there were no significant statistical interactions between the two factors analyzed—AOC: In vs. Out and Lakeshore vs. Inland.

The clear signal in these chemical data are that mink captured near Lake Ontario, and presumably eating organisms exposed to Lake Ontario water and its food web, have significantly higher BCC concentrations in their tissues than mink captured inland. Therefore, it is unlikely that BCC sources in the AOC are contributing to the “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems” use impairments identified in the RAP (1993, 1997). Whether the data collected in the four parts of this study (Wellman and Haynes 2006a, 2006b, 2007; this report) support delisting of these two use impairments will be addressed in detail in the final report for this project (Haynes et al., in prep.).

Statistical and Other Issues

Outliers

Mink #17 was excluded from statistical analyses of adipose and liver tissue, and mink #22 was excluded from analysis of liver tissue. The BCC concentrations reported for these two lakeshore-AOC mink are accurate but very high compared to the other lakeshore mink. When

statistical analyses were run including these animals, the resulting significant interaction effects made the results difficult to interpret. Excluding the data for these animals, there were no interaction effects between the treatments AOC: In vs. Out and Lakeshore vs. Inland.

Several factors may account for the high levels of BCCs in minks #17 and #22. Haynes et al. (2004) reported that one sediment sample from Salmon Creek near where mink #17 was captured had a concentration of 1.5 ppm total PCBs. Mink #17 was caught in the large Braddock Bay wetlands complex with broad access to Lake Ontario water and its food web, particularly the carcasses of migrating salmonines. The stable isotope analysis indicated that it fed on organisms about one-half trophic level higher than other mink captured in the Braddock Bay area (Wellman and Haynes 2006b), suggesting that salmonines may have been in its diet. In contrast, lakeshore mink out of the AOC were captured in the upland portion of the Yanty Creek basin south of the Lake Ontario State Parkway, an area with less direct contact to Lake Ontario.

Mink #22 was captured where Round Pond Creek crosses under the Lake Ontario State Parkway (Figure 1), upstream from an area anecdotally reported to have been a munitions factory long ago. Although not previously suspected to exist (RAP 1993, 1997), it is possible that small toxic hotspots exist in the Braddock Bay area to which mink are exposed. Alternatively, it may be that some mink store more BCCs than others due to individual physiological differences in uptake, biotransformation and excretion rates. For example, before beginning the analyses of the 40 mink discussed here, tissues of six mink from the Tug Hill Plateau (presumably a “clean” area) were analyzed to test analytical procedures. For no apparent reason, mink #28 had very high levels of BCCs (Appendix B).

Statistical Power

In each analysis in this report showing no significant difference between the AOC: In vs. Out treatments and their interactions with the Lakeshore vs. Inland treatments, statistical power was very low despite sample sizes of 8-11 mink per treatment. This result was a consequence of high variation in BCC concentrations among animals, even after eliminating statistical outliers (see above). It is notable that the non-significant differences in tissue concentrations reported for inland mink are always greater than out of the AOC (e.g., TPCB: 1552 vs. 387 ng/g wet, Table 8a; PBDE: 128 vs. 8 ng/g wet, Table 9g). Thus, it is possible that there are differences in the concentrations of BCCs in the tissues of mink in and out of the AOC not detected in our study, but a large number of additional mink would have to be captured and their tissues analyzed to test this hypothesis. Given the large differences in BCC concentrations between lakeshore and inland mink, even if enough additional mink were analyzed it is unlikely that any differences that might be found in concentrations in and out of the AOC would be biologically meaningful in comparison. Therefore, despite low power for the AOC: In vs. Out treatments, the most reasonable conclusion without much greater expense is that there are no biologically meaningful differences in the BCC concentrations of mink in and out of the AOC.

Lack of Co-planar PCB Concentration Data

Due to expense, no co-planar PCB data were collected for this project; therefore, total TEQ values reported here are low. However, it is well established that co-planar PCBs account for 50-90% of dioxin-furan TEQs in tissues (Wellman and Haynes, 2006b), and this issue was addressed in their modeling predictions for total TEQ levels in Rochester Embayment mink.

ACKNOWLEDGMENTS

We are very grateful to Randy Baase who for two winters spent many hours trapping mink in the worst weather, and who taught us much about mink. Thanks are also due to other trappers who contributed carcasses and taught us about mink: Matt Lochner, Dan Carroll, Dan Frey, Michael Ingham, Don Newcombe, Tom Raduns, William Schwartz, Jr., Fred Sinclair and Chuck Tirrano; and to the landowners near the Bergen Swamp who allowed us to collect mink on their property: “Doc” Fink, Dick Sands, Mel Reber and Al Burkhardt. We are grateful to Scott Wells and Ross Abbett for dissecting the mink, and to Albert Fulton for creating Figure 1. The government personnel who graciously allowed us to work in their territories include NYS DEC: Dan Carroll, Dave Woodruff and Heidi Bogner Kennedy; NYS OPRHP: James Slusarczyk; and USFWS: Bob Lamoy and Paul Hess. Sara T. Wellman critically reviewed an earlier version of this document. Finally, we gratefully acknowledge grant C302399 from the New York State Great Lakes Protection Fund that made this study possible.

LITERATURE CITED

- Basu, I. Analysis of PCBs, Pesticides, and PAHs in Air and Precipitation Samples: IADN Project: Sample Preparation Procedures, School of Public and Environmental Affairs, Indiana University, Bloomington, IN, Version 1, June 1995a.
- Basu, I. Analysis of PCBs, Pesticides, and PAHs in Air and Precipitation Samples: IADN Project: Gas Chromatography Procedure, School of Public and Environmental Affairs, Indiana University, Bloomington, IN, Version 1, June 1995b.
- Chiarenzelli, J., Pagano, J., Milligan, M., Holsen, T., Hopke, P., Scudato, R., Falanga, L., Battalagia, T., Migdal, K., Hsu, Y-K, and Hartwell, A. 2001. Enhanced airborne PCB concentrations and chlorination downwind of Lake Ontario. *Environmental Science and Technology* 35(16): 3280-3286.
- Frame, G.M., Cochran, J.W., and Bowadt, S.S. 1996. Complete PCB congener distributions for 17 Aroclor mixture determined by 3 HRGC systems optimized for comprehensive, quantitative, congener-specific analysis. *Journal High Resolution Chromatography* 19(12): 657-668.
- Harlin, K., and Surratt, K. Analysis of PCBs, Pesticides, and PAHs in Air and Precipitation Samples: Sample Preparation Procedures, Illinois State Water Survey, Office of Atmospheric Chemistry, Champaign, IL, SOP #CH-PR-001.3, Version 3, June 1995.
- Haynes, J.M., J.C. Makarewicz, and T.C. Young. 2004. Levels of Bioaccumulative Chemicals of Concern in Air, Water, Sediment and Sentinel Species in the Rochester Embayment of Lake Ontario. Final Report to the NY Great Lakes Protection Fund. SUNY College at Brockport.
- Haynes, J.M., Pagano, J.J., and Wellman, S.T. 2002. New York State Great Lakes Protection Fund Application for State Assistance—RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species.
- Pagano, J.J., Scudato, R.J., Roberts, R.N., and Bemis, J.C. 1995. Reductive dechlorination of PCB-contaminated sediments in an anaerobic bioreactor system. *Environmental Science &*

Technology 29(10):2584-2589.

- Pagano, J.J., Roberts, R.N., Sumner, G.S. 1998. An improved method for the separation of PCB and PCT utilizing alumina adsorption column chromatography. *In* Proceedings of the 216th National Meeting of the American Chemical Society, Boston, MA, USA. 38(2):13-16.
- Pagano, J.J., Rosenbaum, P., Roberts, R., Sumner, G., and Williamson, L. 1999. Assessment of maternal contaminant burden by analysis of snapping turtle eggs. *Journal of Great Lakes Research* 25(4):950-962.
- RAP. 1993. Stage I Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- RAP. 1997. Stage II Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- Stewart, P., Pagano, J., Darvill, T., Lonky, E., and Reihman, J. 2000. Effects of great lakes fish consumption on brain PCB pattern, concentration and progressive ratio performance. *Environmental Research, Section A* 82:18-32.
- USEPA. 1994. Method 7471A. Mercury in Solid or Semisolid Waste (Manual Cold Vapor Technique). U.S. Environmental Protection Agency. Washington, DC.
- USEPA. 1996. Methods 8081A. Organochlorine Pesticides by Gas Chromatography. U.S. Environmental Protection Agency. Washington, DC.
- USEPA 1997. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods (SW-846). U.S. Environmental Protection Agency. CD-ROM Version 2, National Tech. Info. Serv.
- USEPA. 1998. Method 8290A. Polychlorinated Dibenzodioxins (PCDDs) and Polychlorinated Dibenzofurans (PCDFs) by High Resolution Gas Chromatography/High Resolution Mass Spectrometry (HRGS/HRMS). U.S. Environmental Protection Agency. Washington, DC.
- Wellman, S.T., and Haynes, J.M. 2006a. Monitoring Mink Populations using Video Traps. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.
- Wellman, S.T., and Haynes, J.M. 2006b. Age, Size, and Stable Isotope Data of Mink Populations, and a Predictive Model of Bioaccumulation of Chemicals of Concern in the Rochester Embayment of Lake Ontario. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.
- Wellman, S.T., and Haynes, J.M. 2007. Bioaccumulative Chemicals of Concern in Mink: Adverse Effects Levels and Results of a Predictive Model for the Rochester Embayment of Lake Ontario. Department of Environmental Science and Biology, State University of New York at Brockport. Brockport, N.Y.

TABLES

Table 1. Primary column: PCB congeners analyzed with Agilent 25m UltraII.

Peak #	# Cl	IUPAC #	Peak #	# Cl	IUPAC #	Peak #	# Cl	IUPAC #
1	1	1	34	4+4+4	41+64+71	67	7	179
2	1	3	35	5	96	68	6	137
3	2+2	4+10	36	4	40	69	6+7	130+176
4	2+2	7+9	37	4	67	70	6+6+6	138+163+164
5	2	6	38	4	63	71	6	158
6	2+2	5+8	39	4	74	72	6+7	129+178
7		HCB	40	4	70	73	6	166
8	2	14	41	4+5	66+95	74	7+7	182+187
9	3	19	42	5	91	75	7	183
10	3	30	43	4+4+5	56+60+92	76	6+6	128+167
11	2+2	12+13	44	5	84	77	7	185
12	3	18	45	5+5	89+101	78	7	174
13	2+3	15+17	46	5	99	79	7	177
14	3+3	24+27	47	5	119	80	6+7+8	156+171+202
15	3+3	16+32	48	5	83	81	6+7+8	157+173+201
16	3	34	49	5	97	82	7	172
17	3+4	29+54	50	5+5+5	87+115+117	83	8	197
18	3	26	51		p-p'-DDE	84	7	180
19	3	25	52	5	85	85	7	193
20	3	31	53	6	136	86	7	191
21	3	28	54	4+5	77+110+154	87	8	200
22	3+3+4	20+33+53	55	5+6	82+151	88		mirex
23	4	51	56	5+6+6	124+135+144	89	7+7	170+190
24	3	22	57	5+6	109+147	90	8	198
25	4	45	58	5+6	123+149	91	8	199
26	4	46	59	5	118	92	8+8	196+203
27	4	52	60	6	134	93	7	189
28	4+4	43+49	61	5+6	114+133	94	8+9	195+208
29	4+4+4	47+48+75	62	5+6	122+131	95	9	207
30	4	65	63	6	146	96	8	194
31	3	35	64	6	153	97	8	205
32	4	44	65	5+6	105+132	98	9	206
33	3+4+4	37+42+59	66	6	141	99	10	209

Table 2. Confirmation column: PCB congeners analyzed with Agilent 60m DB-XLB.

Peak #	# Cl	IUPAC #	Peak #	# Cl	IUPAC #	Peak #	# Cl	IUPAC #
1	1	1	42	4	42	83	7	179
2	1	2	43	3	35	84	5+6	105+141
3	1	3	44	4	71	85	7	176
4	2+2	4+10	45	3+4	37+41	86	6	137
5	2	9	46	4	64	87	6	130
6	2	7	47	4+5	40+103	88	6	164
7	2	6	48	5	100	89	6	138
8	2	5	49	4	67	90	6	163
		HCB	50	4+5	63+93	91	6+7	129+178
9	2	8	51	5	95	92	6	158
10	3	19	52	4	74	93	7	175
11	2	14	53	4	70	94	7	187
12	3	30	54	4+5	66+91	95	7	183
13	3	18	55	5	92	96	6+7	128+185
14	3	17	56	4+5	56+84	97	7	174
15	2	12	57	5+5	90+101	98	6	167
16	2+3	13+27	58	4	60	99	8	202
17	3	24	59	5	99	100	7	177
18	3	16	60	5+5	83+119	101	7+8	171+201
19	2	15	61	5	97	102	7	173
20	3	32	62	5	87	103	8	197
21	3+4	34+54			p-p' DDE	104	6	156
22	3	29	63	5+6	117+136	105	7	172
23	3	26	64	5+5+6	85+115+154	106	6	157
24	3	25	65	5	110	107	7	180
25	3	31	66	4	81	108	7	193
26	4	53	67	6	151	109	8	200
27	3	28	68	5	82	110	7	191
28	3+3	20+33	69	6	135	111	7	170
29	4	51	70	4+6	77+144	112	8	199
30	4	45	71	6	147	113	7	190
31	3	22	72	6	149			Mirex
32	4	46	73	5	124	114	8	196
33	4	73	74	5+5	109+123	115	8	203
34	4	69	75	6	134	116	9	208
35	4	52	76	5	118	117	7	189
36	4	48	77	6	131	118	8+9	195+207
37	4	49	78	5+6	122+165	119	8	194
38	4+5	47+104	79	6	146	120	8	205
39	4	75	80	5	114	121	9	206
40	4	44	81	6	153	122	10	209
41	4	59	82	6	132			

Table 3. Primary analytical column: organochlorine pesticide components and polybrominated diphenyl ether congeners analyzed with 60m DB-XLB.

Organochlorine Pesticides	PBDEs
cis-chlordane	BDE-17
trans-chlordane	BDE-28
alpha-BHC (HCH)	BDE-47
beta-BHC	BDE-66
delta-BHC	BDE-85
gamma-BHC (Lindane)	BDE-99
dieldrin	BDE-100
endosulfan I	BDE-119
endosulfan II	BDE-138
endosulfan sulfate	BDE-153
endrin	BDE-154
hexachlorobenzene (HCB)	
heptachlor epoxide	Polychlorinated biphenyls (PCBs)
mirex	(Dual column confirmational analysis)
p-p'-DDT	
p-p'-DDD	HP UltraII (25m) 100 zones of 132 congeners/co-eluters
p-p'-DDE	DB-XLB (60m) 122 zones of 155 congeners/co-eluters

Table 4. GLPF mink project surrogate recoveries (SR) and surrogate spike checks for various sample matrices. F1 and F2 denote silica separation fractions.

		SR14-F1	IS30-F1	IS30-F2	SR65-F1	SR166-F1	SRPCT3-F2
Adipose	N=42	90.4%	111.6%	110.1%	89.6%	97.9%	94.2%
	STDEV	17.2%	6.4%	8.7%	20.8%	16.1%	15.8%
Kidney	N=26	78.9%	115.5%	97.8%	80.7%	80.8%	67.0%
	STDEV	11.0%	11.1%	6.5%	20.5%	12.3%	10.9%
Testes	N=33	86.6%	119.6%	110.5%	85.2%	99.2%	89.6%
	STDEV	14.5%	9.4%	6.0%	13.8%	20.0%	14.1%
Liver	N=45	92.0%	118.6%	106.3%	90.2%	82.4%	81.8%
	STDEV	10.4%	7.5%	8.6%	13.8%	26.6%	26.5%
Surrogate	N=33	107.9%	114.2%	98.4%	93.2%	97.8%	95.1%
Checks	STDEV	6.0%	7.1%	3.2%	6.4%	5.8%	3.3%

Table 5. Correlations of concentrations of selected bioaccumulative chemicals of concern (BCC) in adipose and brain (total Hg only) tissue of mink. PCB = polychlorinated biphenyl; DDE = dichlorodiphenyldichloroethylene; TEQ = 2,3,7,8-tetrachlorodibenzo-p-dioxin toxic equivalents; BDE = polybrominated diethyl ether. *P < 0.05, **P < 0.01; 2-tailed significance level.

BCC	Total PCB	Avg. Cl/ Biphenyl	Dieldrin	DDE	Total TEQ	Mirex	Total BDE
Total Hg							
r =	0.484**	0.424*	0.484**	0.530**	0.536**	0.554**	0.561**
P =	0.004	0.012	0.004	0.001	0.002	0.001	0.001
n =	34	34	34	34	31	34	34
Total PCB							
r =		0.468**	0.460**	0.507**	0.654**	0.809**	0.894**
P =		0.0003	0.004	0.001	<0.001	<0.001	<0.001
n =		37	37	37	30	37	37
Avg. Cl/ Biphenyl							
r =			0.284	0.447**	0.367*	0.464**	0.509**
P =			0.088	0.006	0.046	0.004	0.001
n =			37	37	30	37	37
Dieldrin							
r =				0.905**	0.529**	0.441**	0.501**
P =				<0.001	0.003	0.006	0.002
n =				37	30	37	37
DDE							
r =					0.607**	0.541**	0.525**
P =					<0.001	0.001	0.001
n =					30	37	37
Total TEQ							
r =						0.764**	0.737**
P =						<0.001	<0.001
n =						30	30
Mirex							
r =							0.825**
P =							<0.001
n =							37

Table 6. Correlations of concentrations of selected bioaccumulative chemicals of concern (BCC) in liver and brain (Total Hg only) tissue of mink. PCB = polychlorinated biphenyl; DDE = dichlorodiphenyldichloroethylene; TEQ = 2,3,7,8-tetrachlorodibenzo-p-dioxin toxic equivalents; BDE = polybrominated diethyl ether. *P < 0.05, **P < 0.01; 2-tailed significance level.

BCC	Total PCB	Avg. Cl/ Biphenyl	Dieldrin	DDE	Total TEQ	Mirex	Total BDE
Total Hg							
r =	0.474**	0.430**	0.480**	0.533**	0.473**	0.248	0.467**
P =	0.004	0.010	0.004	0.001	0.006	0.150	0.005
n =	35	35	35	35	32	35	35
Total PCB							
r =							
P =		0.479**	0.408*	0.496**	0.722**	0.621*	0.833**
n =		0.002	0.011	0.002	<0.001	*	<0.001
		38	38	33	31	<0.001	38
						38	
Avg. Cl/ Biphenyl							
r =			0.278	0.453**	0.379*	0.274	0.468**
P =			0.091	0.004	0.036	0.096	0.003
n =			38	38	31	38	38
Dieldrin							
r =				0.899**	0.399*	0.113	0.358*
P =				<0.001	0.026	0.499	0.027
n =				38	31	38	38
DDE							
r =					0.532**	0.251	0.450**
P =					0.002	0.129	0.005
n =					31	38	38
Total TEQ							
r =							
P =						0.759*	0.841**
n =						*	<0.001
						<0.001	31
						31	
Mirex							
r =							0.822**
P =							<0.001
n =							38

Table 7. Total mercury concentrations (ng/g) in the brains of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In					
Lakeshore	8	0.281	0.143	AOC: In vs. Out 0.609	0.079
Inland	10	0.158	0.154		
AOC: Out					
Lakeshore vs. Inland					
Lakeshore	9	0.296	0.155	0.032	0.587
Inland	9	0.194	0.145		
Interaction					
				0.835	0.055

Table 8a. Total polychlorinated biphenyl (TCPB) concentrations (ng/g wet) in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In					
Lakeshore	9	3918.3	5780.1	AOC: In vs. Out 0.632	0.076
Inland	11	1552.4	2410.9		
AOC: Out					
Lakeshore vs. Inland					
Lakeshore	10	3970.4	3280.8	0.014	0.708
Inland	8	387.3	226.2		
Interaction					
				0.601	0.081

Table 8b. Average number of chlorine atoms per biphenyl in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In					
Lakeshore	9	6.35	0.28	AOC: In vs. Out 0.475	0.108
Inland	11	6.16	0.24		
AOC: Out					
Lakeshore vs. Inland					
Lakeshore	10	6.37	0.27	0.006	0.817
Inland	8	6.10	0.37		
Interaction					
				0.808	0.141

Table 8c. Total TEQ values for dioxins and furans (ng/Kg) in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	7	10.281	7.229	0.354	0.149
Inland	10	4.723	4.143		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	9	15.436	12.159	0.010	0.763
Inland	6	5.005	5.252		
				Interaction	
				0.405	0.129

Table 8d. Dichlorodiphenyldichloroethylene (DDE) concentrations (ng/g wet) in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	9	5612.2	6118.9	0.357	0.148
Inland	11	2600.5	2967.3		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	5656.1	3081.7	0.002	0.916
Inland	8	276.9	452.8		
				Interaction	
				0.339	0.156

Table 8e. Dieldrin concentrations (ng/g wet) in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	9	40.1	51.4	0.241	0.212
Inland	11	18.6	15.0		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	33.9	17.1	0.007	0.800
Inland	8	3.2	1.7		
				Interaction	
				0.614	0.079

Table 8f. Mirex concentrations (ng/g wet) in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In					
Lakeshore	9	16.2	22.1	AOC: In vs. Out 0.259	0.200
Inland	11	9.3	13.8		
AOC: Out					
Lakeshore	10	72.6	123.0	Lakeshore vs. Inland 0.073	0.436
Inland	8	1.3	1.2		
Interaction					
				0.136	0.317

Table 8g. Polybrominated diphenyl ether (PBDE) concentrations (ng/g wet) in the adipose tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In					
Lakeshore	9	177.9	167.3	AOC: In vs. Out 0.937	0.051
Inland	11	87.6	99.0		
AOC: Out					
Lakeshore	10	227.7	208.5	Lakeshore vs. Inland 0.005	0.838
Inland	8	30.4	11.9		
Interaction					
				0.266	0.196

Table 9a. Total polychlorinated biphenyl (PCB) concentrations (ng/g wet) in the liver tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In					
Lakeshore	8	164.3	219.7	AOC: In vs. Out 0.960	0.050
Inland	11	92.6	168.3		
AOC: Out					
Lakeshore	10	230.5	227.3	Lakeshore vs. Inland 0.018	0.673
Inland	10	20.6	12.7		
Interaction					
				0.232	0.219

Table 9b. Average number of chlorine atoms per biphenyl in the liver tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	8	6.14	0.31	0.449	0.116
Inland	11	6.05	0.36		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	6.37	0.20	0.026	0.618
Inland	10	5.97	0.37		
				Interaction	
				0.145	0.306

Table 9c. Total TEQ values for dioxins and furans (ng/Kg) in the liver tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	3	1.655	1.656	0.547	0.089
Inland	5	1.009	1.778		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	5	7.201	13.772	0.337	0.152
Inland	4	0.206	0.390		
				Interaction	
				0.423	0.120

Table 9d. Dichlorodiphenyldichloroethylene (DDE) concentrations (ng/g wet) in the liver tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	8	305.0	354.0	0.193	0.252
Inland	11	128.2	216.4		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	244.0	157.1	0.005	0.836
Inland	10	8.0	11.0		
				Interaction	
				0.667	0.071

Table 9e. Dieldrin concentrations (ng/g wet) in the liver tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	9	10.7	11.6	0.363	0.146
Inland	11	7.2	14.4		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	11.4	9.3	0.037	0.559
Inland	8	0.3	0.2		
				Interaction	
				0.260	0.200

Table 9f. Mirex concentrations (ng/g wet) in the liver tissue of mink.

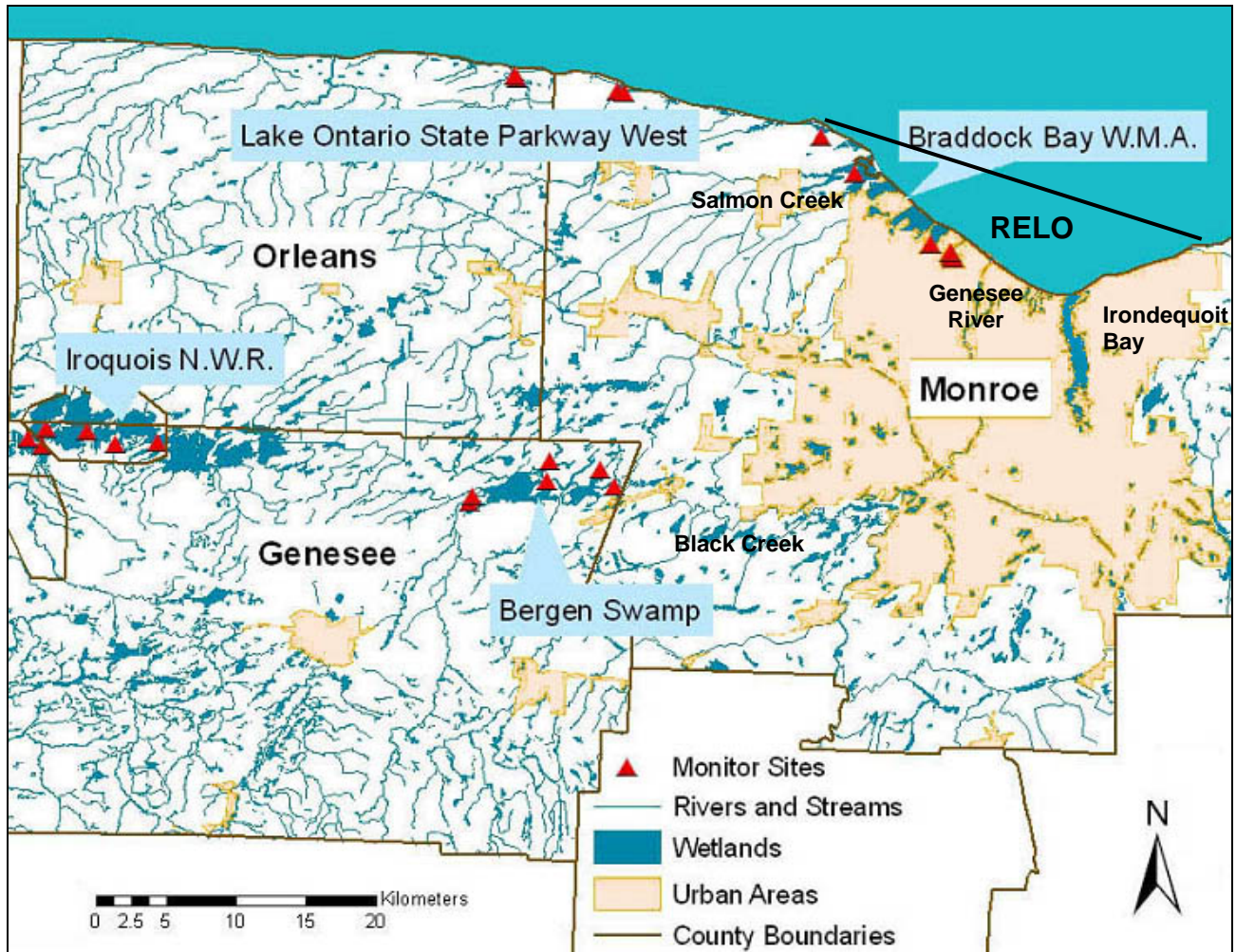
Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	8	3.2	5.4	0.153	0.295
Inland	11	3.1	6.4		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	25.9	40.7	0.066	0.455
Inland	10	0.2	0.2		
				Interaction	
				0.068	0.449

Table 9g. Polybrominated diphenyl ether (PBDE) concentrations (ng/g wet) in the liver tissue of mink.

Location	N	Mean	Std. Dev.	P-value	Power
AOC: In				AOC: In vs. Out	
Lakeshore	8	6.1	7.5	0.811	0.056
Inland	11	3.0	3.7		
AOC: Out				Lakeshore vs. Inland	
Lakeshore	10	9.1	7.1	0.002	0.899
Inland	10	0.8	0.6		
				Interaction	
				0.143	0.308

FIGURES

Figure 4. Map showing the four regions referred to in the study. AOC/Lakeshore is Braddock Bay WMA, AOC/Inland is at least 3 km from Lake Ontario, Out of AOC/Lakeshore is the Lake Ontario State Parkway west of Rte.19, and Out of AOC/Inland is Iroquois NWR and the Tug Hill Plateau (not shown). RELO is the Rochester Embayment of Lake Ontario. (Map by Albert Fulton 2005.)



APPENDICES

Appendix A. Chemicals detected in this study. Names in bold were analyzed statistically and are discussed in the text. Raw data for all chemical are in Appendix B (CD).

Chemical	Abbreviation
Total Polychlorinated Biphenyls ^{1,2}	TCPB
Cl-1	
Cl-2	
Cl-3	
Cl-4	
Cl-5	
Cl-6	
Cl-7	
Cl-8	
Cl-9	
Cl-10	
Pesticides	
(cis) alpha-chlordane	ACHLOR
(trans) gamma-chlordane	GCHLOR
Aldrin	ALDRIN
alpha-BHC (HCH)	ABHC
beta-BHC	BBHC
delta-BHC	DBHC
Dieldrin	DIELDRIN
endosulfan I	ENDO1
endosulfan II	ENDO2
endosulfan sulfate	ENDOSUL
Endrin	ENDRIN
endrin aldehyde	ENDA
endrin ketone	ENDK
gamma-BHC	GBHC
Heptachlor	HEP
heptachlor epoxide	HEPEPO
Hexachlorobenzene	HCB
Methoxychlor	METH
Mirex	MIREX
p-p'-DDD	DDD
p-p'-DDE	DDE
p-p'-DDT	DDT
Total Brominated Diethyl Ethers	TBDE
BDE-17	BDE17
BDE-28	BDE28
BDE-47	BDE47

BDE-66	BDE66
BDE-85	BDE85
BDE-99	BDE99
BDE-100	BDE100
BDE-119	BDE119
BDE-138	BDE138
BDE-153	BDE153
BDE-154	BDE154

Dibenzo-p-dioxins

2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD)	TCDD
1,2,3,7,8-Pentachlorodibenzo-p-dioxin (PeCDD)	PeCDD
1,2,3,4,7,8-Hexachlorodibenzo-p-dioxin (HxCDD)	HxCDD
1,2,3,6,7,8-Hexachlorodibenzo-p-dioxin (HxCDD)	HxCDD2
1,2,3,7,8,9-Hexachlorodibenzo-p-dioxin (HxCDD)	HxCDD3
1,2,3,4,6,7,8-Heptachlorodibenzo-p-dioxin (HpCDD)	HpCDD
Octachlorodibenzo-p-dioxin (OCDD)	OCDD

Tetrachlorodibenzofurans

2,3,7,8-Tetrachlorodibenzofuran (TCDF)	TCDF
1,2,3,7,8-Pentachlorodibenzofuran (PeCDF)	PeCDF
2,3,4,7,8-Pentachlorodibenzofuran (PeCDF)	PeCDF2
1,2,3,4,7,8-Hexachlorodibenzofuran (HxCDF)	HxCDF
1,2,3,6,7,8-Hexachlorodibenzofuran (HxCDF)	HxCDF2
1,2,3,7,8,9-Hexachlorodibenzofuran (HxCDF)	HxCDF3
2,3,4,6,7,8-Hexachlorodibenzofuran (HxCDF)	HxCDF4
1,2,3,4,6,7,8-Heptachlorodibenzofuran (HpCDF)	HpCDF
1,2,3,4,7,8,9-Heptachlorodibenzofuran (HpCDF)	HpCDF2
Octachlorodibenzofuran (OCDF)	OCDF

Total Toxic Equivalents¹ (TEQ) **Total TEQ**

Total Mercury **Total Hg**

¹Excluding co-planar PCBs. See Discussion for explanation.

²PCB homologues were determined by spreadsheet manipulations (see NYS GLPF MINK worksheet in Appendix B) based on congener-specific PCB measurements. PCB data were further processed such that the mole percent (congener specific and homologue) and average chlorine/biphenyl (Cl/BP) values were generated. Coeluting congeners were assumed to be in equal proportions for all spreadsheet calculations (Pagano et al., 1995).

Appendix B. Separate spreadsheet on a CD with all BCC data collected in this study.

Appendix 4

Bioaccumulative Chemicals of Concern in Mink: Adverse Effects Levels and Results of a Predictive Model for the Rochester Embayment of Lake Ontario

Sara T. Wellman and James M. Haynes

Department of Environmental Science and Biology
State University of New York
College at Brockport
350 New Campus Drive
Brockport, NY 14420-2973

March 2007

OVERVIEW

This report is the fourth of four resulting from project C302399 funded by the New York Great Lakes Protection Fund in 2004 to address use impairments related to water quality identified in the Remedial Action Plan for the Rochester Embayment of Lake Ontario (RELO RAP). It gives a brief review of pertinent literature and reports the results of a model for bioaccumulation of selected chemicals in mink in the Rochester Embayment. Previous reports addressed the 1) development and use of videocapture (Mustelavision) systems that established the presence and reproduction of mink in and out of the RELO RAP Area of Concern (AOC); ages, sizes and trophic positions (stable isotope analysis) of mink (*Mustela vison*) in the study area; and 3) levels of BCCs in mink tissues. Because mink are the most sensitive known species to BCCs, the results of this four-part project will determine whether delisting the fish and wildlife reproduction impairment for the RELO AOC can be recommended.

TABLE OF CONTENTS

INTRODUCTION	115
ADVERSE EFFECTS LEVELS.....	115
Method	115
Results And Discussion	116
Dioxins And Furans	116
Polychlorinated Biphenyls.....	116
Mercury.....	117
Overlaps Of PCB And Dioxin/Furan Toxicities.....	117
Conclusion	117
BIOACCUMULATION MODEL	118
Method	118
Results And Discussion	119
SUMMARY	120
ACKNOWLEDGMENTS	120
LITERATURE CITED	1203
Table 1. Selected endpoints and effects levels reported for mercury, PCBs and TEQs in mink diets and tissues	123
Table 2. TEQ values from dioxins and furans for Lakeshore and Inland mink livers, showing high, low and average values for each category	124
Table 3. Predicted versus measured values for tissue residues of dioxin/furans (TEQs), methylmercury, and PCBs, based on water concentrations in Lake Ontario	124
Table A-1. Adverse Effects Levels of Dioxin	125
Table A-2. Adverse Effects Levels of Polychlorinated Biphenyls.....	127
Table A-3. Adverse Effects Levels of Mercury.....	130
Table A-4. Adverse Effects Levels of Organochlorine Pesticides	132
Table A-5. Adverse Effects Levels of Multiple Polyhalogenated Hydrocarbons	133
References Used in Appendix	137

INTRODUCTION

In the 1980s the binational (Canada, U.S.) International Joint Commission (IJC) began the process of creating and implementing remedial action plans (RAPs) in 43 contaminated areas of concern (AOCs) throughout the Great Lakes Basin. The IJC established 14 “use impairments” that could cause a local area to be “listed” as an AOC, including “degradation of fish and wildlife populations” and “bird or animal deformities or reproductive problems.” In the Rochester AOC, both uses were defined as impaired because “very few” mink (*Mustela vison*) were then being trapped or observed within 2 miles of the lake (RAP 1993, 1997). This study was part of a project (Haynes *et al.* 2002) to determine if populations of mink on the shore of the Rochester Embayment of Lake Ontario (RELO) are negatively impacted by bioaccumulative chemicals of concern (BCCs) and, if so, whether the BCCs originate in the embayment watershed or elsewhere.

The RELO AOC includes the Embayment, a 35 square mile portion of Lake Ontario south of a line between Bogus Point in the town of Parma and Nine Mile Point in the town of Webster (both in Monroe County, New York); adjacent wetlands and bays; and the six mile reach of the Genesee River from the Lower Falls to the mouth at Lake Ontario. The RAP also includes the subwatersheds of Salmon Creek, the Genesee River and Irondequoit Creek (RAP 1993, 1997).

The initial questions addressed by this portion of the study were: 1) Which BCCs, and at what levels, are known to cause adverse effects on populations or reproduction, or to cause deformities, in mink? 2) How do predicted levels of BCCs in mink tissues (based on concentrations in Lake Ontario water) compare with measured tissue residues in our lakeshore mink specimens?

Our approach to the first question was to do a literature search, looking for reports on the levels of BCCs in mink tissues corresponding to adverse effects. This gave us values to compare to the tissue residues we found in mink (Pagano and Haynes, in preparation). The results of the comparisons will be described in the final report for this project (Haynes *et al.*, in preparation), along with an assessment of risk to AOC and Lake Ontario shoreline mink.

To answer the second question, our approach was to provide a predictive model for the bioaccumulation and biomagnification of BCCs in AOC mink tissues based on concentrations of BCCs in the waters of Lake Ontario (Wellman and Haynes 2006). If model predictions are well correlated with actual tissue concentrations in mink, by knowing tissue residue levels that cause adverse effects we will have created a risk assessment tool for mink without the expense of tissue analyses.

ADVERSE EFFECTS LEVELS

Method

Basu *et al.* (2006) wrote an extensive review of the literature on the toxic effects of BCCs in mink, covering field and lab studies and the use of mink in hazard assessments. Leonards *et al.* (1995) and Kannan *et al.* (2000) reviewed the available literature on the toxicity of PCBs to mink. Rather than duplicate those efforts, we focused on studies which linked dietary levels of BCCs to tissue residues as well as to reproductive or other effects. Such studies allow direct comparisons of our tissue residue results (Pagano and Haynes, in preparation) to levels associated with adverse effects. We also concentrated on reports of

chronic exposures, as they would best represent the exposures of mink to BCCs in the environment. Search engines used included Academic Search Premier, BioOne, BasicBIOSIS, InfoTrac OneFile, JSTOR, and ScienceDirect.

Results and Discussion

We found studies linking dietary concentrations to tissue residues for dioxins and furans, PCBs, and mercury. No studies of this type were found for any of the organo-chlorine (OC) pesticides except hexachlorobenzene (HCB). Rush *et al.* (1983) reported increased kit mortality at 1 ppm HCB, resulting in adipose residues of 95 ppb and undetectable liver residues. According to Giesy *et al.* (1994), studies in the 1970s and 1980s determined that OC pesticides were not the cause of effects seen in mink that ate Great Lakes fish. Because OC pesticide levels have decreased in the environment since then, they would be even less significant today, which probably accounts for the lack of studies regarding them.

Dioxins and furans

The toxicity of dioxins and furans to mink is well-established (Basu *et al.* 2006, Hochstein *et al.* 1998, 2001, Render *et al.* 2000, 2001), and LOAELs (Lowest Observable Adverse Effect Levels) are frequently reported as 2,3,7,8-TCDD equivalents (TEQs) in ppb (dietary) or pg/g (tissue residue) wet weight.

Many studies evaluated toxic end points in terms of dietary concentrations but not as tissue residues in the mink. Render *et al.* (2000) reported that a dietary concentration of 5 ppb 2,3,7,8-TCDD fed to adult females for six months caused proliferation of squamous epithelial cells in bone adjacent to teeth, and Render *et al.* (2001) found the same effect in 6- and 12-week-old kits fed 2.4 ppb 2,3,7,8-TCDD for as little as 14 days. Hochstein *et al.* (1998) reported that 1 ppb caused 62.5% mortality in adult female mink fed 2,3,7,8-TCDD for 125 days. Hochstein *et al.* (2001) found that when mink dams were fed 0.053 ppb 2,3,7,8-TCDD, kit survival was reduced to 47% vs. 83% in the control group (0.0006 ppb TCDD). Unfortunately, these studies do not provide data that is directly comparable to our tissue analyses (Pagano and Haynes, in preparation).

Tissue residues as well as dietary concentrations were reported by several researchers feeding wild-caught fish to mink. Since the fish used as the source of BCCs contained PCBs as well as dioxins and furans, their total TEQ values also included contributions from the coplanar PCBs in the fish (Table 1 and Appendix A). Heaton *et al.* (1995) and Tillit *et al.* (1996) reported reduced 3- and 6-week-old kit survival at a maternal dietary concentration of 22.4 pg TEQ/g (0.72 µg/g TPCBs), which resulted in maternal liver residues of 208.3 pg TEQ/g (2.19 µg/g TPCBs). Bursian *et al.* (2006a, b) reported increased mortality in 3- to 6-week-old mink kits whose dams had been fed 68.5 pg TEQ/g (3.7 µg/g TPCBs), resulting in maternal liver residue levels of 218.4 pgTEQ/g (3.133 µg/g TPCBs). Bursian *et al.* (2006a, b, c) also found jaw lesions in 27- and 31-week-old juveniles fed 47 pg TEQ/g (1.1 µg/g TPCBs) and 9.2 pg TEQ/g (0.96 µg/g TPCBs), respectively, with corresponding juvenile liver residues of 75 pg TEQ/g (16 µg/g TPCBs) and 40.2 pg TEQ/g (1.7 µg/g TPCBs).

Polychlorinated biphenyls

Bursian *et al.* (2006a) found that the dietary LC₁₀ and LC₂₀ for total PCBs were 0.231 and 0.984 µg/g TPCBs, and estimated a threshold dietary concentration of 33.2 pg/g TPCBs. Restum *et al.* (1998) reported reduced whelping in dams fed 0.25 ppm PCBs, resulting in a liver concentration of 860 ng/g. Halbrosk *et al.* (1999) found a trend (P = 0.069) for reduced

litter size in dams fed 1360 ppb Aroclor 1260 equivalent (EQ) PCBs in the diet, resulting in a maternal liver concentration of 7.25 ppm Aroclor 1260 EQ and a maternal adipose concentration of 129 ppm Aroclor 1260 EQ. However, these results may have been confounded by the presence of mercury in the fish used in the diet, resulting in 0.22 ppm Hg in the diet and 3.67 ppm in the maternal livers.

Mercury

In 1974, Aulerich *et al.* reported that all mink fed a diet including 5 ppm mercury died after 29 days of treatment, with brain residue levels of approximately 20 ppm. Wobeser *et al.* (1976) reported that 1.1 ppm dietary MeHg caused “classic mercury intoxication,” including neurotoxicity; the corresponding brain concentration was 8.2 ppm. Wren *et al.* (1987a, b) reported that 1.0 µg/g dietary MeHg caused adult mortality as well as a reduction in litter size, with a brain residue of 15.3 µg/g. However, this dietary level is misleading because the 1 ppm chow was used only every other day after unexpected mortalities within less than three months.

Dansereau *et al.* (1999) reported a reduction in the proportion of females whelping at dietary levels of 0.5 ppm Hg, resulting in liver residues of 80.4 µg/g. Halbhook *et al.* (1997) found reduced litter size at 0.22 ppm dietary Hg (in fish), resulting in liver residues of 3.67 ppm. Using the average of brain:liver residue ratios from Evans *et al.* (2000), Wobeser *et al.* (1976) and Wren *et al.* (1987a, b), we estimated the brain residues for the reduced whelping (Dansereau *et al.* 1999) as 23.2 µg/g, and for reduced litter size (Halbrook *et al.* 1997) as 1.06 µg/g. While these estimations are not strictly accurate, because the brain:liver ratio varies within and among studies, they should be close enough to allow comparison with our brain residue results (Pagano and Haynes, in preparation).

Overlaps of PCB and dioxin/furan toxicities

Unfortunately, miscommunication between PIs Pagano and Haynes and with Columbia Analytical Services resulted in no analyses for co-planar PCBs in our study. However, several studies show distinct relationships between total TEQ, PCB TEQ and dioxin-furan TEQ. Analyses of fish from the Housatonic River used in mink diets by Bursian *et al.* (2006a) showed that approximately 91% of the dietary TEQ value was contributed by PCBs, and only about one-tenth of the TEQ value by dioxins and furans. According to Bursian (2006c), Heaton *et al.* (1995) and Tillitt *et al.* (1996) reported that PCBs accounted for 73% of the TEQ in fish from Saginaw Bay, Michigan. In contrast, PCBs made up less than 44% of the TEQ in the Saginaw Bay fish examined by Bursian *et al.* (2006c), with about half the TEQ value contributed by dioxins and furans. Thus, an estimate of the total environmental TEQ exposure, based on analysis of only dioxins and furans, would have to multiply the dioxin/furan TEQ by a correction factor between two and ten, and this is what we did.

Conclusion

In our literature review, jaw lesions had the lowest LOAELs (Table 1). Table 2 shows total TEQ values (low, average, high) for lakeshore and inland mink in the AOC, based on dioxin/furan analyses, and calculated estimates including co-planar PCBs. The highest measured TEQ value for AOC lakeshore mink in our study was 47.62 pg TEQ/g wet weight (Pagano and Haynes, in preparation), which is higher than the LOAEL of 40 pg TEQ/g liver at which jaw lesions were seen (Bursian *et al.* 2006a, b). The lowest measured TEQ value in lakeshore mink, 0.22 pg TEQ/g, even when multiplied by ten is still an order of magnitude smaller than the LOAEL, indicating no risk. However, the average (excluding high and low) of 7.8 pg TEQ/g for our lakeshore mink (Pagano and Haynes, in preparation), if multiplied by a correction factor of five, approaches the LOAEL for jaw lesions. This would indicate that some

lakeshore mink are at risk of developing jaw lesions, which have been shown to lead to jaw deformities, osteolysis, and tooth loss (Render *et al.* 2001). Histological examinations of our minks' jaws for this lesion could help to determine whether to delist the RELO AOC for deformities. Further comparisons of the levels of BCCs in our mink tissues to LOAELs will be presented in our final report (Haynes *et al.* in preparation).

The highest measured TEQ value for inland mink was 4.16 pg TEQ/g. When multiplied by the correction factor of ten, the result is approximately equal to the LOAEL of 40 pg TEQ/g, indicating that the most exposed of the inland mink may be at risk for developing jaw lesions. However, the lowest value of 0.00 pg TEQ/g and the average TEQ value of 0.25 pg TEQ/g, even when multiplied by a factor of ten, indicate that the majority of inland mink are not at risk.

BIOACCUMULATION MODEL

Method

For modeling the bioaccumulation of chemicals in mink, we started with Equation 28 from Sample *et al.* (1996), adding the units for clarity:

$$C_w \left(\frac{mg}{L} \right) = \frac{NOAEL \left(\frac{mg}{kg \cdot d} \right) \times bw(kg)}{W \left(\frac{L}{d} \right) + \left[F \left(\frac{kg}{d} \right) \times BAF \right]}, \quad (1)$$

where C_w is the concentration of the BCC in the water, $NOAEL$ is the No Observed Adverse Effects Level; W and F are the daily water and food consumption rates in L/day and kg/day, respectively; BAF is the bioaccumulation factor for the chemical of concern (based on the trophic level of the animal and the octanol-water partition coefficient, k_{ow} , a measure of hydrophobicity of the compound); and bw is the body weight of the animal in kilograms (Sample *et al.* 1996).

We solved for $NOAEL$, and taking into account the aquatic portion of the animal's diet (P_{aq}), got an equation to predict the exposure level of an animal to a BCC in water:

$$NOAEL \left(\frac{mg}{kg (bw) \cdot d} \right) = \frac{C_w \left(\frac{mg}{kg} \right) [W(kg) + (F(kg) \times P_{aq} \times BAF)]}{bw(kg)}. \quad (2)$$

According to Sample *et al.* (1996), the dietary concentration C_f (mg/kg) equivalent to the $NOAEL$ is

$$C_f \left(\frac{mg}{kg} \right) = \frac{NOAEL \left(\frac{mg}{kg \cdot d} \right) \times bw(kg)}{F \left(\frac{kg}{d} \right)}. \quad (3)$$

Substituting equation (2) for $NOAEL$ into equation (3) for C_f , the bw terms in the numerator and denominator cancel out and give a dietary concentration equivalent to the exposure level based on the BCC concentration in water, the food and water consumption rates of mink (177 g and 0.1 L as reported by Wellman and Haynes 2006), the percent aquatic diet

(between 50 and 90% as in USEPA 1995), and the bioaccumulation factor (Sample *et al.* 1996):

$$C_f \left(\frac{mg}{kg} \right) = \frac{C_w \left(\frac{mg}{kg} \right) \left[W \left(\frac{kg}{d} \right) + F \left(\frac{kg}{d} \right) \times P_{aq} \times BAF \right]}{F \left(\frac{kg}{d} \right)}. \quad (4)$$

This dietary concentration equivalent can be directly compared to dietary concentrations of BCCs known to cause adverse effects in mink.

Using the highest and lowest values for diet-to-tissue biomagnification factors (BMF_t) calculated from the literature (see Appendix A), we predicted levels of selected BCCs in mink tissue with the equation

$$C_t \left(\frac{\mu g}{g} \right) = C_f \left(\frac{\mu g}{g} \right) \times BMF_t. \quad (5)$$

Results and Discussion

Table 3 compares the predicted low and high values of total PCBs and TEQs from dioxins and furans to the lowest and highest tissue levels found in the livers, and the values of MeHg in the brains, of lakeshore mink. The low predicted values were calculated using the lowest C_w found in either Luckey and Litton (2005) or in Environment Canada's 2004 survey of Lake Ontario (J. Vincent, personal communication), and assuming 50% aquatic diet and the lowest diet-to-tissue BMF calculated from the literature (Hg: Wobeser *et al.* 1976; TEQs: Heaton *et al.* 1995, Tillitt *et al.* 1996; PCBs: Bursian *et al.* 2006a, b). The high predicted values were calculated using the highest C_w , 90% aquatic diet, and highest BMF (Hg: Wobeser *et al.* 1976; TEQs: Heaton *et al.* 1995, Tillitt *et al.* 1996; PCBs: Halbrook *et al.* 1999). The measured values were provided by J. Pagano (personal communication) and will be detailed in a forthcoming report (Pagano and Haynes, in preparation).

The model worked well for dioxin/furan TEQs and for PCBs. In both cases, the predicted low and high values bounded our measured values, except for the low estimate for PCBs, which was very close to the lowest measured value in lakeshore mink. This is to be expected, as the AOC is neither the most polluted nor the cleanest portion of Lake Ontario (Luckey and Litton 2005; J. Vincent, personal communication). The model did not predict tissue levels of mercury well; the measured values were up to three orders of magnitude higher than predicted values. The reason for this discrepancy is not known. One possibility is the fact that the model is based on the octanol-water coefficient, a concept which applies only to lipophilic compounds, which mercury is not. However, Sample *et al.* (1996) apparently intended the model to be used with mercury, as they provided BAF factors for it (as well as several other heavy metals). Another possibility is that the model predicts mercury concentrations in tissue based only on aquatic exposures, while the mink in our study might have had exposure to mercury through terrestrial sources unaccounted for by the model. Further investigation and development of the model will be required if it is deemed necessary to predict mercury levels in mink of the Rochester Embayment.

SUMMARY

The first question addressed by this study was: Which BCCs, and at what levels, are known to cause adverse effects on populations or reproduction, or to cause deformities, in mink? We found that the most sensitive endpoint for PCBs and TEQs was a squamous epithelial cell lesion in mink jawbones, corresponding to 40.2 pg TEQ/g wet weight in mink livers (Bursian *et al.* 2006a, b). We compared this level to tissue residues in Lake Ontario shoreline mink, and concluded that lakeshore mink are at risk for developing jaw lesions. We have the jaws of mink used in this study. A small amount of additional funding would permit analysis of lesions and final determination of whether mink in the Rochester Embayment AOC are adversely impacted by BCCs.

The second question addressed by this study was: How do predicted levels of BCCs in mink tissues (based on concentrations in Lake Ontario water) compare with measured tissue residues in our lakeshore mink specimens? We found that for PCBs and dioxins/furans, the model worked well, but it was less successful in predicting levels of mercury in lakeshore mink.

ACKNOWLEDGMENTS

We are grateful to Steve Bursian, Bob Gilliam, Kurunthachalam Kannan, Fred Luckey, Gary Neuderfer, Don Tillit, Jennifer Vincent, and John Waud for providing articles or data required for this study.

LITERATURE CITED

- Aulerich, R.J., R.K. Ringer, and S. Iwamoto. 1974. Effects of dietary mercury on mink. *Archives of Environmental Contamination and Toxicology* 2(1): 43-51
- Basu, N., A.M. Scheuhammer, S.J. Bursian, J. Elliot, K. Rouvinen-Watt, and H. M. Chan. 2006. Mink as a sentinel species in environmental health. *Environmental Research* 103: 130-144.
- Bursian, S.J., C. Sharma, R.J. Aulerich, B. Yamini, R.R. Mitchell, C.E. Orazio, D.R.J. Moore, S. Svirsky, and D.E. Tillit. 2006a. Dietary exposure of mink (*Mustela vison*) to fish from the Housatonic River, Berkshire County, Massachusetts, USA: Effects on reproduction, kit growth, and survival. *Environmental Toxicology and Chemistry* 25(6): 1533-1540.
- Bursian, S.J., C. Sharma, R.J. Aulerich, B. Yamini, R.R. Mitchell, K.J. Beckett, C.E. Orazio, D.R.J. Moore, S. Svirsky, and D.E. Tillit. 2006b. Dietary exposure of mink (*Mustela vison*) to fish from the Housatonic River, Berkshire County, Massachusetts, USA: Effects on organ weights and histology and hepatic concentrations of polychlorinated biphenyls and 2,3,7,8-tetrachlorodibenzo-*P*-dioxin toxic equivalence. *Environmental Toxicology and Chemistry* 25(6): 1541-1550.
- Bursian, S.J., K.J. Beckett, B. Yamini, P.A. Martin, K. Kannan, K.L. Shields, F.C. Mohr. 2006c. Assessment of effects in mink caused by consumption of carp collected from the Saginaw River, Michigan, USA. *Archives of Environmental Contamination and Toxicology* 50: 614-623.

- Dansereau, M., N. Lariviere, D. Du Tremblay, and D. Belanger. 1999. Reproductive performance of two generations of female semidomesticated mink fed diets containing organic mercury contaminated freshwater fish. *Archives of Environmental Contamination and Toxicology* 36: 221-226.
- Evans, R.D., E.M. Addison, J.Y. Villeneuve, K.S. MacDonald, and D.G. Joachim. 2000. Distribution of inorganic and methylmercury among tissues in Mink (*Mustela vison*) and Otter (*Lutra canadensis*). *Environmental Research Section A* 84: 133-139.
- Giesy, J.P., D.A. Verbrugge, R.A. Othout, W.W. Bowerman, M.A. Mora, P.D. Jones, J.L. Newsted, C. Vandervoot, S.N. Heaton, R.J. Aulerich, S.J. Bursian, J.P. Ludwig, G.A. Dawson, T.J. Kubiak, D.A. Best, and D.E. Tillitt. 1994. Contaminants in fishes from Great Lakes-influenced sections and above dams of three Michigan rivers. II: Implications for health of mink. *Archives of Environmental Contamination and Toxicology* 27: 213-223.
- Halbrook, R.S., L.A. Lewis, R.I. Aulerich, and S.J. Bursian. 1997. Mercury accumulation in mink fed fish collected from streams on the Oak Ridge Reservation. *Archives of Environmental Contamination and Toxicology* 33: 312-316.
- Haynes, J.M., Pagano, J.J., and Wellman, S.T. 2002. New York State Great Lakes Protection Fund Application for State Assistance—RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species.
- Heaton, S.N., S.J. Bursian, J.P. Giesy, D.E. Tillitt, J.A. Render, P.D. Jones, D.A. Verbrugge, T.J. Kubiak, and R.J. Aulerich. 1995. Dietary exposure of mink to carp from Saginaw Bay, Michigan. 1. Effects on reproduction and survival, and the potential risks to wild mink populations. *Archives of Environmental Contamination and Toxicology* 28:334-343
- Hochstein, J.R., S.J. Bursian, and R.J. Aulerich. Effects of dietary exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin in adult female mink (*Mustela vison*). *Archives of Environmental Contamination and Toxicology* 35: 348-353.
- Hochstein, J.R., J.A. Render, S.J. Bursian, R.J. Aulerich. 2001. Chronic toxicity of dietary 2,3,7,8-tetrachlorodibenzo-P-dioxin to mink. *Veterinary and Human Toxicology* 43(3): 134-139.
- Kannan, K., A.L. Blankenship, P.D. Jones, and J.P. Giesy. 2000. Toxicity reference values for the toxic effects of polychlorinated biphenyls to aquatic mammals. *Human and Ecological Risk Assessment* 6(1): 181-201.
- Leonards, P.E.G., T.H. de Vries, W. Minnaard, and S. Stuijzand. 1995. Assessment of experimental data on PCB –induced reproduction inhibition in mink, based on an isomer- and congener-specific approach using 2,3,7,8-tetrachlorodibenzo-p-dioxin toxic equivalency. *Environmental Toxicology and Chemistry* 14(4): 639-652
- Luckey, F., and S. Litten. 2005. Bioaccumulative Contaminants in Lake Ontario Surface Water, 1999. USEPA. New York, NY.

- RAP. 1993. Stage I Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- RAP. 1997. Stage II Remedial Action Plan for the Rochester Embayment of Lake Ontario. Monroe County Department of Health. Rochester, NY.
- Render, J.A., J.R. Hochstein, R.J. Aulerich, and S.J. Bursian. 2000. Proliferation of periodontal squamous epithelium in mink fed 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *Veterinary and Human Toxicology* 42(2): 85-86
- Render, J.A., S.J. Bursian, D.S. Rosenstein, and R.J. Aulerich. 2001. Squamous epithelium proliferation in the jaws of mink fed diets containing 3,3',4,4',5-pentachlorobiphenyl (PCB 126) or 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *Veterinary and Human Toxicology* 43(1): 22-26
- Restum, J.C., S.J. Bursian, J.P. Giesy, J.A. Render, W.G. Helferich, E.B. Shipp, D.A. Verbrugge, and R.J. Aulerich. 1998. Multigenerational study of the effects of consumption of PCB-contaminated carp from Saginaw Bay, Lake Huron, on Mink. 1. Effects on mink reproduction, kit growth and survival, and selected biological parameters. *Journal of Toxicology and Environmental Health, Part A*, 54: 343-375.
- Rush, G.F., J.H. Smith, K. Maita, M. Bleavins, R.J. Aulerich, R.K. Ringer, and J.B. Hook. 1983. Perinatal hexachlorobenzene toxicity in the mink. *Env. Res.* 31:116-124.
- Sample, B.E., D.M. Opresko, and G.W. Suter II. 1996. Toxicological Benchmarks for Wildlife: 1996 Revision. Oak Ridge National Laboratory, Oak Ridge, TN.
- Tillitt, D.E., R.W. Gale, J.C. Meadows, J.L. Zajicek, P.H. Peterman, P.D. Jones, S.J. Bursian, T.J. Kubiak, J.P. Giesy, and R.J. Aulerich. 1996. Dietary exposure of mink to carp from Saginaw Bay, Michigan. 3. Characterization of dietary exposure to planar halogenated hydrocarbons, dioxin equivalents, and biomagnification. *Environmental Science and Technology* 30: 283-291.
- Wellman, S.T. and J.H. Haynes. 2006. Age, size, and stable isotope data of mink populations, and a predictive model of bioaccumulation of chemicals of concern in the Rochester embayment of Lake Ontario. Prepared for the NY Great Lakes Protection Fund, NY Department of Environmental Conservation. Buffalo, NY.
- Wobeser, G., Nielsen, N.O., Schiefer, B., 1976a. Mercury and mink. II. Experimental methyl mercury intoxication. *Canadian Journal of Comparative Medicine* 40: 34-45.
- Wren, C.D., Hunter, D.B., Leatherland, J.F., Stokes, P.M., 1987a. The effects of polychlorinated biphenyls and methylmercury, singly and in combination, on mink. I. Uptake and toxic responses. *Archiv. Env. Contamination Toxicol.* 16: 441-447.
- Wren, C.D., Hunter, D.B., Leatherland, J.F., Stokes, P.M., 1987b. The effects of polychlorinated biphenyls and methylmercury, singly and in combination on mink. II. Reproduction and kit development. *Archiv. Env. Contamination Toxicol.* 16: 449-454.

TABLES

Table 1. Selected endpoints and effects levels reported for mercury, PCBs, and TEQs in mink diets and tissues. (Values in italics were estimated by the authors of this report, using the average brain:liver ratios from Evans *et al.* 2000, Wobeser *et al.* 1976, and Wren *et al.* 1987a, b.) CDD = chlorinated dibenzo dioxins, CDF = chlorinated dibenzo furans, HCB = hexachlorobenzene.

Impairment	Endpoint	Toxin	Effect Level	Conc. (ppm or ug/g)		Reference
				Diet	Tissue	
Brain						
Population	Adult mortality	Hg	LC100	5 ppm	19.9 ppm	Aulerich et al. 1974
Reproduction	Whelping reduced	Hg in fish	LOAEL	0.5 ppm	23.2 <i>ug/g</i>	Dansereau et al. 1999
Reproduction	Litter size reduced	Hg in fish	LOAEL	0.22 ppm	1.06 <i>ppm</i>	Halbrook et al. 1997
Population	Hg intoxication	MeHg	LOAEL	1.1 ppm	8.2 ppm	Wobeser et al. 1976
Reproduction	Litter size reduced	MeHg	LOAEL	1.0 ug/g	2.0 ug/g	Wren et al. 1987a,b
Liver						
Reproduction	Kit survival 3 & 6 wks	PCBs	LOAEL	720 pg/g	2190 pg/g	Heaton et al. 1995, Tillit et al. 1996
		CDDs	LOAEL	60 pg/g	2626 pg/g	
		CDFs	LOAEL	13 pg/g	335 pg/g	
		TEQs	LOAEL	22.4 pg/g	208.3 pg/g	
Deformities	Jaw lesion in 31-wk kits	PCBs	LOAEL	0.96 ug/g	1.698 ug/g	Bursian et al. 2006a, b
		TEQs	LOAEL	9.2 pg/g	40.2 pg/g	
Deformities	Jaw lesion in 27-wk kits	PCBs	LOAEL	1.1 ug/g	16 ug/g	Bursian et al. 2006c
		TEQs	LOAEL	47 pg/g	75 pg/g	
Reproduction	Litter size	PCBs	LOAEL	1360 ppb	7250 ppb	Halbrook et al. 1999
Reproduction	P-1 Whelping reduced	PCBs	LOAEL	0.25 ppm	860 ng/g	Restum et al. 1998
	F-2 Kit mortality	PCBs	LOAEL	0.5 ppm	464 ng/g	
<u>Adipose</u>						
Reproduction	Kit mortality	HCB	LOAEL	1 ppm	95 ppb	Rush et al. 1983

Table 2. TEQ values from dioxins and furans for Lakeshore and Inland mink livers, showing high, low and average (excluding high and low) values for each category.

Location	Value	TEQ	TEQ*2	TEQ*10
Lakeshore	Low	0.22	0.44	2.2
	Average (8)	7.75	15.50	77.5
	High	47.62	95.24	476.2
Inland	Low	0.00	0.00	0.00
	Average (8)	0.25	0.50	2.50
	High	4.16	8.32	41.6

Table 3. Predicted versus measured values for tissue residues of dioxin/furans (TEQs), methylmercury, and PCBs, based on water concentrations in Lake Ontario as reported by J. Vincent (2006, personal communication) and Luckey and Litton (2005).

		Water Conc.	Tissue Level	
			Predicted	Measured
BCC	Value	pg/kg	ng/g	ng/g
TEQs (liver)	Low	6.00E-05	5.52E-05	2.20E-04
	High	2.40E-02	6.21E-02	2.13E-02
MeHg (brain)	Low	0.00E+00	0.00E+00	9.00E+01
	High	1.80E+01	4.70E+00	1.55E+03
PCBs (liver)	Low	2.60E+01	1.92E+01	1.36E+01
	High	9.15E+02	1.60E+05	5.87E+03

Appendix A: Adverse Effects Levels for Selected BCCs

Notes

NOAEL = No Observed Adverse Effects Level

LOAEL = Lowest Observed Adverse Effects Level

ETD = Estimated Threshold Dose; geometric mean of NOAEL and LOAEL $=(\text{SqRt}(\text{NOAEL} \times \text{LOAEL}))$

LC10 = Concentration Lethal to 10% of population

TEQs = 2,3,7,8-Tetrachloro-dibenzi-P-dioxin equivalent units

Tissue/Diet BMF is diet-to-tissue Biomagnification Factor

In rare cases, we have made an estimation based on reported data. Our estimations are shown in italics.

Unless noted as "estimated by STW", all calculations are done by the authors.

NA = Not Available or Not Addressed in report

LOAELs for each report are highlighted in yellow

Estimated threshold values are highlighted in gold

A-1. Adverse Effects Levels of Dioxin							
Endpoint	Toxin	Effect	Concentration		References	Notes	
		Level	Diet	Tissue			
Adult Mortality	TCDD	NOAEL	0.1 ppb		Hochstein et al. 1998		
		LOAEL	1.0 ppb				
Adult Mortality	TCDD	NOAEL	0.18 ppb	N/A	Hochstein et al. 2001		
		LOAEL	1.40 ppb				
Kit survival @ 3 wks		NOAEL	0.0006 ppb				
		LOAEL	0.053 ppb				
Toenail deformities		NOAEL	0.18 ppb				
		LOAEL	1.40 ppb				
Jaw lesions	TCDD	NOAEL	0 ppb	N/A	Render et al. 2000		
		LOAEL	5 ppb				
Jaw lesions	TCDD	NOAEL	0 ppb	N/A	Render et al. 2001	Jaw lesions caused deformities and displacement of teeth	
		LOAEL	2.4 ppb				
Kit mortality		NOAEL	0 ppb	N/A			
		LOAEL	24 ppb				

A-2. Adverse Effects Levels of Polychlorinated Biphenyls									
Impairment	Endpoint	Toxin	Effect	Concentration		References	Notes	Tissue/Diet	
			Level	Diet	Tissue			BMF	
Reproduction	Kit survival reduced	PCBs	NOAEL	1.6 ug/g	3.083 ug/g	Bursian et al. 2006i, 2006ii			
		PCBs	LOAEL	3.7 ug/g	3.133 ug/g			1.9	
		PCBs	ETD	2.4 ug/g				0.8	
		PCBs	LC10	0.231 ug/g			calc. by STW from report data		
		PCBs	LC20	0.984 ug/g			estimated by regression analysis		
		TEQs	NOAEL	16.1 pg/g	55.9 pg/g		estimated by regression analysis		
		TEQs	LOAEL	68.5 pg/g	218.4 pg/g			3.5	
		TEQs	ETD	33.2 pg/g				3.2	
Deformities	Jaw lesion in 31-wk kits	PCBs	LOAEL	0.96 ug/g	1.698 ug/g			1.8	
		TEQs	LOAEL	9.2 pg/g	40.2 pg/g			4.4	
Reproduction	Reproduction and kit survivability				Liver	Bursian et al. 2006iii	Reproduction effects measured included breeding success, whelping success, gestation length, and litter size.		
		PCBs	NOAEL	1.7 ug/g	NA				
Deformities	Jaw lesion in 27-wk kits	TEQs	NOAEL	73 pg/g	NA				
		PCBs	NOAEL	0.83 ug/g	8.1 ug/g	Jaw lesions can lead to displaced and loose teeth (see Render et al 2000, 2001)	9.8		
		PCBs	LOAEL	1.1 ug/g	16 ug/g		14.5		
		TEQs	NOAEL	28 pg/g	28 pg/g		1.0		
TEQs	LOAEL	47 pg/g	75 pg/g						
Reproduction	Litter size				Liver	Halbrook et al. 1999	PCBs = Aroclor 1260 equivalents, in fish		
		PCBs	NOAEL	1009 ppb	<5 ppb				
		PCBs	LOAEL	1360 ppb	7250 ppb			Results may have been confounded by presence of Hg in fish (Halbrook et al 1997)	5.3
					Adipose				
		PCBs	NOAEL	1009 ppb	105860 ppb			105.0	
		PCBs	LOAEL	1360 ppb	128630 ppb			95.0	

					Liver	Martin et al. 2006				
Metabolic	Plasma T4 increase in 6-wk kits	TPCBs	NOAEL	0.03 mg/kg	0.19 ug/kg				6.3	
			LOAEL	0.83 mg/kg	4.38 ug/kg				5.3	
			TEQs	NOAEL	3.4 ng/kg		3.1 pg/kg			0.9
				LOAEL	27.9 ng/kg		72 pg/kg			2.6
	Plasma T3 decrease in 27- wk juveniles	TPCBs	NOAEL	1.05 mg/kg	16.25 ug/kg			Many of our mink were only a month or two older than these juveniles	15.5	
			LOAEL	1.69 mg/kg	17.79 ug/kg				10.5	
			TEQs	NOAEL	47.6 ng/kg		74.6 pg/kg			1.6
				LOAEL	73.2 ng/kg		105.6 pg/kg			1.4
	Plasma retinol and retinyl esters decreased in 6- wk kits and 27- wk juveniles	TPCBs	NOAEL	1.05 mg/kg	Kits 10.54 ug/kg juvs 16.25 ug/kg					
			LOAEL	1.69 mg/kg	kits 18.8 ug/kg juvs 17.79 ug/kg					
			TEQs	NOAEL	47.6 ng/kg		Kits 152.4 pg/kg juvs 74.6 pg/kg			
				LOAEL	73.2 ng/kg		Kits 306.6 pg/kg juvs 105.6 pg/kg			
	Kidney retinyl esters reduced in 6-wk kits and 27-wk juveniles	TPCBs	NOAEL	0.03 mg/kg	kits 0.19 ug/kg juvs 2.57 ug/kg					
				LOAEL	0.83 mg/kg	kits 4.38 ug/kg juvs 8.14 ug/kg				

		TEQs	NOAEL	3.4 ng/kg	kits 3.1 pg/kg juvs 7.6 pg/kg			
			LOAEL	27.9 ng/kg	kits 71.2 pg/kg juvs 26.9 pg/kg			
		PCBs			Liver	Restum et al. 1998	Liver concentrations here are weighted averages of male and female values given in report	
Reproduction	Delayed estrus P-1 and F-1		NOAEL	0.0 ppm	P-1 70.7 ng/g F-1 83.2 ng/g			
			LOAEL	0.25 ppm	P-1 860 ng/g F-1 635 ng/g			P-1 =3.4 F-1 = 2.5
	Reduced mating P-1 females		NOAEL	0.5 ppm	923 ng/g			1.8
			LOAEL	1.0 ppm	1580 ng/g			1.6
	Reduced whelping P-1		NOAEL	0.0 ppm	70.7 ng/g			
			LOAEL	0.25 ppm	860 ng/g			3.4
	F-1 Kit mortality		NOAEL	0.25 ppm	635 ng/g			2.5
			LOAEL	0.5 ppm	967 ng/g			1.9
	F-1 Kit body weight		NOAEL	0.0 ppm	83.2 ng/g			
			LOAEL	0.25 ppm	635 ng/g			2.5
	F-2 Kit mortality		NOAEL	0.25 ppm	275 ng/g		No F-2 males at 0.5 ppb or above, or females at 1.0 ppb, survived to 3 weeks	1.1
			LOAEL	0.5 ppm	464 ng/g			0.9
	F-2 Kit body weight		NOAEL	0.25 ppm	275 ng/g			
			LOAEL	0.5 ppm	464 ng/g			
Deformity	Jaw lesions	PCBs	NOAEL	0 ppb	N/A	Render et al. 2001	Jaw lesions caused deformities and displacement of teeth	
			LOAEL	24 ppb				
Population	Kit mortality		NOAEL	0 ppb	N/A			
			LOAEL	24 ppb				

A-3. Adverse Effects Levels of Mercury									
Impairment	Endpoint	Toxin	Effect	Concentration (ppm or ug/g)		Reference s	Notes	Tissue/Diet	
			Level	Diet	Tissue			BMF	
Population		Hg			Brain	Aulerich et al. 1974		Brain	
	Adult mortality		LC100	5 ppm	19.9 ± 4.55 ppm			All mink died after 30-37 days after treatment started; treatment ended on 29th day.	4.0
Population	Adult mortality	tHg in fish			Liver	Dansereau et al. 1999		Liver	
			LC60	1.0 ppm	96.6 ppm			In 1st generation females	96.6
Reproduction	Whelping reduced		NOAEL	0.1	28.2 ppm			In 1st and 2nd generation females	282.0
			LOAEL	0.5 ppm	80.4 ppm				168.0
					Brain				Brain
			LC60	1.0 ppm	26.0 ug/g			<i>Estimated by STW using average of Liver:Brain levels in other studies = 3.7</i>	
			NOAEL	0.1	7.6 ug/g				
		LOAEL	0.5 ppm	21.6 ug/g					
		tHg in env.		N/A	Brain	Evans et al. 2000			
					0.34 ± 0.24 ug/g			<i>Liver:Brain Ratio =</i>	4.5
					Liver				
					1.53 ± 1.24 ug/g				
					Fur				
					17.26 ug/g				
					Brain				
		MeHg in env.		N/A	0.26 ± 0.19 ug/g		<i>Liver:Brain Ratio =</i>	4.7	
					Liver				
					1.21 ± 0.85 ug/g				
					Fur				
					11.25 ug/g				

Reproduction	Reduced litter Size	Hg in fish			Liver	Halbrook et al. 1997		Liver		
				0.02 ppm			0.41 ppm		20.5	
				0.05 ppm			0.61 ppm		12.2	
				0.09 ppm			1.06 ppm		11.8	
				NOAEL 0.15 ppm			1.93 ppm		12.9	
				LOAEL 0.22 ppm			3.67 ppm		16.7	
							Fur			
				0.02 ppm			3.79 ppm			
				0.05 ppm			7.43 ppm			
				0.09 ppm			7.71 ppm			
				NOAEL 0.15 ppm			13.44 ppm			
				LOAEL 0.22 ppm			19.03 ppm			
							<i>Brain</i>			
				0.02 ppm			0.11 ppm	<i>Estimated by STW using average of Liver:Brain levels in other studies = 3.4</i>		
				0.05 ppm			0.18 ppm			
				0.09 ppm			0.31 ppm			
		NOAEL 0.15 ppm		0.56 ppm						
		LOAEL 0.22 ppm		1.06 ppm						
Population	Adult mortality	MeHg			Liver	Wobeser et al. 1976	Report does not specify whether total or methyl Hg in tissues, but Hg in feed was MeHg.		Liver	
				NOAEL 1.1 ppm			25.4 ppm			25.4
				LOAEL 1.8 ppm			21.3ppm			21.3
							Brain			
				NOAEL 1.1 ppm			8.2 ppm		7.5	
				LOAEL 1.8 ppm			8.2 ppm		10.4	
				LC40 1.8 ppm			8.2 ppm			
				Est. Threshold			10 ppm			
	Sugg. Diag. Crit.			5ppm		Suggested diagnostic criterion				

	Classic Hg intoxication (neuro-toxicity, etc.)				Liver		Liver
			NOAEL	0.1 ppm	0.45 ppm		4.5
			LOAEL	1.1 ppm	25.4 ppm		23.1
						Brain	Brain
			NOAEL	0.1 ppm	0.1 ppm		1.0
			LOAEL	1.1 ppm	8.2 ppm		7.5
Population	Adult mortality	MeHg				Brain	Brain
			NOAEL	0.5 ug/g	N/A		
			LOAEL	1.0 ug/g	15.3 ug/g		15.3
						Liver	Liver
			NOAEL	0.5 ug/g	N/A		
			LOAEL	1.0 ug/g	44.1 ug/g		44.1
			LC56	1.0 ug/g			
			LC19	1.0 ug/g			
Reproduction			Kits/female mated	NOAEL	1.0 ug/g	0.84 ug/g	
		LOAEL	1.0 ug/g	2.0 ug/g			
	Kit growth	NOAEL	1.0 ug/g	2.0 ug/g			
		NOAEL	0.5 ug/g	1.32 ug/g			
		LOAEL	1.0 ug/g	0.84 ug/g			
						<i>Liver:Brain Ratio (above)</i> =	2.9
					Wren et al. 1987i, ii		
					Dietary levels are somewhat misleading because after unexpected mortalities within less than 3 months, the 1.0 ppm chow was used only every other day.		
					9 of 16 in treatment group died		
					3 of 16 in treatment group died		
					Addition of PCBs decreased effect on #kits/female mated		
					Hg without PCBs had no effect		

A-4. Adverse Effects Levels of Organochlorine Pesticides							
Impairment	Endpoint	Toxin	Effect Level	Concentration		References	Notes
				Diet	Tissue		
Population	Adult Mortality	Heptachlor	NOAEL	50 mg/kg		Aulerich et al. 1990	
			LOAEL	100 mg/kg			
Reproduction	Reduced number of second matings	Lindane	LOAEL	1 mg/kg bw/d		Beard et al. 1997, Beard and Rawlings 1998	NOAEL for all is zero, as only one concentration was tested.
	Reduced whelping (incl. embryo loss)		LOAEL	1 mg/kg bw/d			
	Increase duration of pregnancy		LOAEL	1 mg/kg bw/d			
	Reduced litter size (2nd generation)		LOAEL	1 mg/kg bw/d			Effect "far greater" in 2nd gen. than in 1st gen.
	Reduced testis size		LOAEL	1 mg/kg bw/d			
Reproduction	Reduced number of second matings		PCP	LOAEL	1 mg/kg bw/d		
	Reduced whelping (incl. embryo loss)	LOAEL		1 mg/kg bw/d			

	Increase duration of pregnancy		LOAEL	1 mg/kg bw/d		
	Increased prostate hyperplasia		LOAEL	1 mg/kg bw/d		
	Increased serum progesterone	Carbofuran	LOAEL	0.05 mg/kg bw/d		NOAEL for all is zero, as only one concentration was tested.
Population	Adult Mortality	HCB	NOAEL	31 ppm	Bleavins et al. 1984	
			LOAEL	106 ppm		
Reproduction	Litter Size (total)		NOAEL	2 ppm,		
			LOAEL	31 ppm		
	Litter Size (live)		NOAEL	2 ppm,		
			LOAEL	31 ppm		
	Kit weight (birth)		NOAEL	0 ppm		
			LOAEL	1 ppm		
	Kit weight (3 & 6 wks)		NOAEL	1 ppm		
			LOAEL	2 ppm,		
	Kit mortality (3 & 6 wks)		NOAEL	0 ppm		
			LOAEL	1 ppm		
	Adult Mortality Female		Heptachlor	NOAEL		0 ppm
		LOAEL		5 ppm		
		LC50	10.5 ppm			
	Adult Mortality Male		NOAEL	5 ppm		
		LOAEL	12 ppm			
	% Kits stillborn		NOAEL	5 ppm		
		LOAEL	12 ppm			
	Kit weight (birth)		NOAEL	5 ppm		
		LOAEL	12 ppm			
	Kit weight (3 & 6 wks)		NOAEL	0 ppm		
		LOAEL	5 ppm			
			NOAEL	5 ppm		

	Kit mortality (3 wks)		LOAEL	12 ppm			
	Kit mortality (17 wks)	HCB			Adipose	Rush et al. 1983	Tissue levels not given here were all ND (not detected)
			NOAEL	0 ppm	36 ppb		
			LOAEL	1 ppm	95 ppb		
				5 ppm	626 ppb		
					Brain		
				5 ppm	36 ppb		
					Liver		
				1 ppm	5.6 ppb		
				5 ppm	37 ppb		
					Kidney		
				1 ppm	4 ppb		
				5 ppm	15 ppb		
					Muscle		
				1 ppm	1 ppb		
		5 ppm	8 ppb				
				<i>Estimated by STW, based on Liver:Adipose at 5 ppb dietary level.</i>			

A-5. Adverse Effects Levels of Multiple Polyhalogenated Hydrocarbons										
Impairment	Endpoint	Toxin	Effect Level	Concentration		References	Notes	Tissue/Diet		
				Diet	Tissue			BMF		
					Liver					
Reproduction	Gestation length	PCBs	NOAEL	0.015 ug/g	90 pg/g	Heaton et al. 1995, Tillit et al. 1996	Mink were fed fish from Saginaw Bay which contained PCBs, dioxins (CCD), and furans (CDF) (and other BCCs).	6		
		CDDs		42 pg/g	617 pg/g			14.7		
		CDFs		1 pg/g	6 pg/g			6		
		PCBs	LOAEL	.72 ug/g	2190 pg/g			3		
		CDDs		60 pg/g	2626 pg/g			43.8		
		CDFs		13 pg/g	335 pg/g			25.8		
		TEQs	NOAEL	0.9 pg/g	17.5 pg/g			10.7		
		TEQs	LOAEL	22.4 pg/g	208.3 pg/g			16.7		
		Ave. litter size	PCBs	NOAEL	1.53 ug/g			3050 pg/g	Values for TEQs calculated by STW from Table 8 of Tillit et al (1996) using WHO TEQs from van den Berg et al (1998), per D. Tillitt's suggestion (2006, pers. comm.).	2
			CDDs		46 pg/g			2329 pg/g		50.6
	CDFs			20 pg/g	551 pg/g	27.6				
	PCBs		LOAEL	2.56 ug/g	6270 pg/g	2.4				
	CDDs			84 pg/g	3002 pg/g	35.7				
	CDFs			43 pg/g	914 pg/g	21.2				
	TEQs		NOAEL	43.4 pg/g	321.6 pg/g	14.5				
	TEQs		LOAEL	86.8 pg/g	560.1 pg/g	11.1				
	No. live kits	PCBs	NOAEL	1.53 ug/g	3050 pg/g					
		CDDs		46 pg/g	2329 pg/g					
		CDFs		20 pg/g	551 pg/g					
		PCBs	LOAEL	2.56 ug/g	6270 pg/g					
		CDDs		84 pg/g	3002 pg/g					

		CDFs		43 pg/g	914 pg/g			
		TEQs	NOAEL	43.4pg/g	321.6 pg/g			
		TEQs	LOAEL	86.8pg/g	89560 pg/g			
	Kit body wt birth	PCBs	NOAEL	.72 ug/g	2190 pg/g			
		CDDs		60 pg/g	2626 pg/g			
		CDFs		13 pg/g	335 pg/g			
		PCBs	LOAEL	1.53 ug/g	3050 pg/g			
		CDDs		46 pg/g	2329 pg/g			
		CDFs		20 pg/g	551 pg/g			
		TEQs	NOAEL	22.4 pg/g	208.3 pg/g			
		TEQs	LOAEL	43.4 pg/g	321.6 pg/g			
		Kit body wt 3 & 6 wks	PCBs	NOAEL	.015 ug/g	90 pg/g		
			CDDs		42 pg/g	617 pg/g		
	CDFs			1 pg/g	6 pg/g			
	PCBs		LOAEL	.72 ug/g	2190 pg/g			
	CDDs			60 pg/g	2626 pg/g			
	CDFs			13 pg/g	335 pg/g			
	TEQs		NOAEL	0.9 pg/g	17.5 pg/g			
	TEQs		LOAEL	22.4 pg/g	208.3 pg/g			
	Kit survival 3 & 6 wks	PCBs	NOAEL	.015 ug/g	90 pg/g			
		CDDs		42 pg/g	617 pg/g			
		CDFs		1 pg/g	6 pg/g			
		PCBs	LOAEL	.72 ug/g	2190 pg/g			
		CDDs		60 pg/g	2626 pg/g			
		CDFs		13 pg/g	335 pg/g			
		TEQs	NOAEL	0.9 pg/g	17.5 pg/g			
		TEQs	LOAEL	22.4 pg/g	208.3 pg/g			

**References
Used in
Appendix A.**

To avoid confusion, we have used i, ii, iii, here when referring to multiple papers in the same year, as the report may have referred to them in a different order as a, b, c.

- Aulerich et al. 1974 = Aulerich, R.J., R.K. Ringer, and S. Iwamoto. 1974. Effects of dietary mercury on mink. Archives of Environmental Contamination and Toxicology 2(1): 43-51
- Aulerich et al. 1990 Aulerich, R.J., S.J. Bursian, and A.C. Napolitano. 1990. Subacute Toxicity of dietary heptachlor to min (*Mustela vison*). Archives of Environmental Contamination and Toxicology. 19: 913-916
- Beard et al. 1997 = Beard, A.P., A.C. McRae, and N.C. Rawlings. 1997. Reproductive efficiency in mink (*Mustela vison*) treated with the pesticides lindane, carbofuran, and pentachlorophenol. Journal of Reproduction and Fertility 111: 21-28.
- Beard and Rawlings 1998 = Beard, A.P., and N.C. Rawlings. 1998. Reproductive effects in mink (*Mustela vison*) exposed to the pesticides lindane, carbofuran, and pentachlorophenol in a multigeneration study. Journal of Reproduction and Fertility 113: 95-104
- Bleavins et al. 1984 = Bleavins, M.R., S.J. Bursian, J.S Brewster, and R.J. Aulerich. 1984. Effects of dietary hexachlorobenzene exposure on the reproductive performance and survivability of mink and European ferrets. Arch Environ. Contam. Toxicol. 13: 357-365.
- Bursian et al. 2006i = Bursian, S.J., C. Sharma, R.J. Aulerich, B. Yamini, R.R. Mitchell, C.E. Orazio, D.R.J. Moore, S. Svirsky, and D.E. Tillit. 2006. Dietary Exposure of mink (*Mustela vison*) to fish from the Housatonic River, Berkshire County, Massachusetts, USA: Effects on reproduction, kit growth, and survival. Environmental Toxicology and Chemistry 25(6): 1533-1540.
- Bursian et al. 2006ii = Bursian, S.J., C. Sharma, R.J. Aulerich, B. Yamini, R.R. Mitchell, K.J. Beckett, C.E. Orazio, D.R.J. Moore, S. Svirsky, and D.E. Tillit. 2006. Dietary Exposure of mink (*Mustela vison*) to fish from the Housatonic River, Berkshire County, Massachusetts, USA: Effects on organ weights and histology and hepatic concentrations of polychlorinated biphenyls and 2,3,7,8-tetrachlorodibenzo-*P*-dioxin toxic equivalence. Environmental Toxicology and Chemistry 25(6): 1541-1550.
- Bursian et al. 2006iii = Bursian, S.J., K.J. Beckett, B. Yamini, P.A. Martin, K. Kannan, K.L. Shields, F.C. Mohr. 2006. Assessment of effects in mink caused by consumption of carp collected from the Saginaw River, Michigan, USA. Arch. Environ. Contam. Toxicol. 50:614-623.

- Crum et al/ 1993 = Crum, J.A. S.J. Bursian, R.J. Aulerich, D. Polin, and W.E. Braselton. 1993. The reproductive effects of dietary heptachlor in mink (*Mustela vison*). Arch. Environ. Contam. Toxicol. 24:156-164.
- Dansereau et al. 1999 = Dansereau, M., N. Lariviere, D. Du Tremblay, and D. Belanger. 1999. Reproductive performance of two generations of female semidomesticated min fed diets containing organic mercury contaminated freshwater fish. Arch. Environ. Contam. Toxicol. 36:221-226.
- Evans et al. 2000 = Evans, R.D., E.M. Addison, J.Y. Villeneuve, K.S. MacDonald, and D.G. Joachim. 2000. Distribution of inorganic and methylmercury among tissues in Mink (*Mustela vison*) and Otter (*Lutra canadensis*). Environmental Research Section A 84: 133-139.
- Halbrook et al. 1997 = Halbrook, R.S., L.A. Lewis, R.I. Aulerich, and S.J. Bursian. 1997. Mercury accumulation in mink fed fish collected from streams on the Oak Ridge Reservation. Arch. Environ. Contam. Toxicol. 33:312-316.
- Halbrook et al. 1999 = Halbrook, R.S., R.I. Aulerich, S.J. Bursian, and L.A. Lewis. 1999. Ecological risk assessment in a large river-reservoir: 8. Experimental study of the effects of polychlorinated biphenyls on reproductive success in mink. Env. Toxicol. Chem. 18(4): 649-654.
- Heaton et al. 1995 = Heaton, S.N., S.J. Bursian, J.P. Giesy, D.E. Tillitt, J.A. Render, P.D. Jones, D.A. Verbrugge, T.J. Kubiak, and R.J. Aulerich. 1995. Dietary exposure of mink to carp from Saginaw Bay, Michigan. 1. Effects on reproduction and survival, and the potential risks to wild mink populations. Arch. Environ. Contam. Toxicol. 28:334-343
- Hochstein et al. 1998 = Hochstein, J.R., S.J. Bursian, and R.J. Aulerich. Effects of dietary exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin in adult female mink (*Mustela vison*). Arch. Environ. Cont. Toxicol. 35: 348-353.
- Hochstein et al. 2001 = Hochstein, J.R., J.A. Render, S.J. Bursian, R.J. Aulerich. 2001. Chronic toxicity of dietary 2,3,7,8-tetrachlorodibenzo-P-dioxin to mink. Vet. Human Toxicol. 43(3): 134-139.
- Martin et al 2006 = Martin, P.A., G.J. Mayne, S. Bursian, V. Palace, and K. Kannan. Changes in thyroid and vitamin A status in mink fed polyhalogenated-aromatic-hydrocarbon-contaminated carp from Saginaw River, Michigan, USA. Environmental Research 101:53-67.
- Rush et al. 1983 = Rush, G.F., J.H. Smith, K. Maita, M. Bleavins, R.J. Aulerich, R.K. Ringer, and J.B. Hook. 1983. Perinatal hexachlorobenzene toxicity in the mink. Environmental Research 31:116-124.

- Render et al.
2000 = Render, J.A., J.R. Hochstein, R.J. Aulerich, and S.J. Bursian. 2000. Proliferation of Periodontal Squamous Epithelium in mink fed 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *Veterinary and Human Toxicology* 42(2): 85-86
- Render et al.
2001 = Render, J.A., S.J. Bursian, D.S. Rosenstein, and R.J. Aulerich. 2001. of Squamous epithelium proliferation in the jaws of mink fed diets containing 3,3',4,4',5-pentachlorobiphenyl (PCB 126) or 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *Veterinary and Human Toxicology* 43(1): 22-26
- Restum et al.
1998 = Restum, J.C., S.J. Bursian, J.P. Giesy, J.A. Render, W.G. Helferich, E.B. Shipp, D.A. Verbrugge and R.J. Aulerich. 1998. *Journal of Toxicology and Environmental Health, Part A*, 54: 343-375.
- Tillitt et al.
1996 Tillitt, D.E., R.W. Gale, J.C. Meadows, J.L. Zajicek, P.H. Peterman, P.D. Jones, S.J. Bursian, T.J. Kubiak, J.P. Giesy, and R.J. Aulerich. 1996. Dietary exposure of mink to carp from Saginaw Bay, Michigan. 3. Characterization of dietary exposure to planar halogenated hydrocarbons, dioxin equivalents, and biomagnification. *Environ. Sci. Technol.* 30: 283-291.
- Wren 1987i = Wren, C.D., Hunter, D.B., Leatherland, J.F., Stokes, P.M., 1987. The effects of polychlorinated biphenyls and methylmercury, singly and in combination, on mink. I. Uptake and toxic responses. *Arch. Environ. Contam. Toxicol.* 16, 441-447.
- Wren 1987ii = Wren, C.D., Hunter, D.B., Leatherland, J.F., Stokes, P.M., 1987. The effects of polychlorinated biphenyls and methylmercury, singly and in combination on mink. II. Reproduction and kit development. *Arch. Environ. Contam. Toxicol.* 16, 449-454.
- Wobeser et al.
1976 = Wobeser, G., Nielsen, N.O., Schiefer, B., 1976. Mercury and mink. II. Experimental methyl mercury intoxication. *Can. J. Comp. Med.* 40, 34-45.

Appendix 5. Mink jaw deformities in relation to total PCBs and TEQs in their tissues. Mink 17 had multiple proliferative squamous cysts from incisor to molar on the mandible and both maxillae. na = not analyzed, nd = not detected, us = unsuitable sample.

Mink	AOC-Lakeshore			Deformities		
	Total PCB Liver ng/g ww	Adipose TEQ pg/g ww	Liver TEQ pg/g ww	Mandible	Left Maxilla	Right Maxilla
17	5870.8	338.98	21.26	Yes	Yes	Yes
20	86.4	na	Na			
21	682.0	nd	3.50	us	No	No
22	2388.8	22.38	47.62			
38	213.6	10.79	0.30			
39	35.3	1.24	1.17			
41	32.2	3.59	Nd			
56	14.7	7.66	nd	us	No	No
57	153.4	10.23	nd			
58	96.4	16.07	nd			
AOC-Inland						
1	8.5	0.03	0.10	No	No	No
23	18.1	na	na			
24	27.3	3.99	0.64			
43	29.7	1.63	nd			
44	13.4	0.31	nd			
45	28.8	3.23	nd			
59	10.6	3.38	nd			
60	12.8	3.99	nd			
61	64.2	8.96	nd	No	No	Us
62	250.5	12.57	4.16			
63	554.4	9.13	nd	No	No	No
Out of AOC-Lakeshore						
46	229.8	19.63	2.09	No	No	No
47	43.4	9.35	nd			
48	184.8	na	na			
49	755.0	38.29	38.31	No	No	No
50	171.2	9.22	nd			
51	411.3	30.14	nd			
52	69.1	5.43	nd			
53	13.6	5.30	0.92	No	No	No
54	66.7	3.54	nd			
55	360.0	18.03	nd			

Out of AOC-Inland

3	11.7	4.78	0.00			
5	10.0	na	na			
10	8.0	0.35	0.01			
11	27.8	na	na			
14	7.0	nd	0.03	us	No	No
30	45.0	na	na	No	No	No
31	31.4	na	na			
32	14.5	na	na			
33	19.3	na	na	us	No	No
34	31.7	0.79	nd			

DRAFT

Final Report: BUI Delisting Studies in the Rochester Embayment AOC, 2013-2014

James M. Haynes and Sara T. Wellman
The College at Brockport, SUNY
350 New Campus Drive
Brockport, NY 14420-2973

December 2015

Executive Summary

1. Substantial evidence of live mink was observed along the shoreline of the Genesee River portion of the RE AOC, which supports delisting the “mink are present and are reproducing” criterion of the Degradation of Fish and Wildlife Populations and the Loss of Fish and Wildlife Habitat BUIs.
2. According to the USFWS Habitat Suitability Index Model, habitat appears to be highly suitable (85%) for mink along the Genesee River shoreline of the RE AOC, which supports delisting of the Loss of Fish and Wildlife Habitat BUI.
3. For total mercury chemical analysis:
 - a. No amphibian, crayfish and lower trophic level fish samples exceeded the published dietary lowest observed adverse effect level (LOAEL) for mink.
 - b. All upper trophic level fish samples exceeded the published dietary LOAEL for mink (500 ng/g), by 13% on average.
4. For PAH, PCB and dioxin (CDD)/furan (CDF) chemical analyses:
 - a. None of the 12 composited mink prey samples exceeded dietary LOAELs for total PCBs (960,000 pg/g) and TEQ for CDD/CDF (9.2 pg/g).
 - b. Ten of the 12 samples did not exceed the dietary LOAEL for PAHs, co-planar PCBs, and CDD/CDF combined (9.2 pg/g).
 - c. One upper trophic level fish sample exceeded the dietary LOAEL for PAHs by 147% because it contained ~100 times more PAHs (which accounted for 95% of total TEQ in that sample) than the other two samples.
 - d. One lower trophic level fish sample exceeded the dietary LOAEL for PCB TEQ by 4% because it contained ~90 times more PCB 126 (which accounted for 93% of total TEQ in that sample) than the other two samples.
5. Mink hazard assessment:
 - a. Using the “highest exposure” mink diet found in published literature (92% from aquatic sources), and using mean concentrations of BUI contaminants found in

potential mink prey in the Genesee River portion of the RE AOC, the maximum dietary exposure of mink would be 81% of the LOAEL for total mercury, 23% of the LOAEL for total PCBs, and 69% of the LOAEL for total TEQ (PAHs + CDD/CDF + co-planar PCBs). This is the “worst case” diet scenario.

- b. Using the average of six mink diets reported in published literature (65% from aquatic sources) comparable to what mink would eat in the Genesee River portion of the RE AOC, and using mean concentrations of BUI contaminants found in potential mink prey in the study area, the dietary exposure of mink would be 48% of the LOAEL for total mercury, 13% of the LOAEL for total PCBs, and 40% of the LOAEL for total TEQ. This is the “likely” diet scenario.
6. It would be reasonable to delist the Bird or Animal Deformities or Reproductive Problems BUI in the RE AOC because:
- a. Except for total mercury (13% above) and total TEQ (3.4% below; CDD/CDF, PAH and co-planar PCB TEQ combined) in upper trophic level fish, mean concentrations of BUI contaminants in the other three mink prey groups (crayfish, amphibians, lower trophic level fish) were far below dietary LOAELs for mink.
 - b. Using a worst case diet (92% aquatic) for mink, and the analytically-determined mean concentrations of BUI contaminants in potential prey, a hazard assessment showed that the dietary LOAELs for total mercury, total PCBs, and total TEQ would not be exceeded for mink in the Genesee River portion of the RE AOC.

Introduction

The Rochester Embayment Area of Concern (RE AOC) is located north of the City of Rochester, New York, and includes the 35-mi² portion of Lake Ontario south of a line between Bogus Point in the Town of Parma and Nine Mile Point in the Town of Webster (both in Monroe County, NY), adjacent wetlands and bays, and the 6-mile reach of the Genesee River from the river’s mouth at Lake Ontario to the Lower Falls in Rochester (Figure 1). Most of the river corridor in the AOC is urban, commercial or residential but the steep gorge makes access by foot challenging. The river has high boat traffic in summer (recreation) and fall (salmon fishing). Our study focused on mink prey in the Genesee River portion of the AOC, while an earlier study (Haynes et al. 2007) focused on mink in the Braddock Bay Fish and Wildlife Management Area in the western portion of the AOC. Both studies will be used to support delisting of the “Loss of Fish and Wildlife Habitat,” “Degradation of Fish and Wildlife Populations” and “Bird or Animal Deformities or Reproductive Problems” Beneficial Use Impairments (BUI) in the RE AOC.

The known or suspected chemicals thought to cause the “Degradation of Fish and Wildlife Populations” and “Bird or Animal Deformities or Reproductive Problems” BUIs are mercury, polychlorinated biphenyls (PCBs), dioxins (CDD)/furans (CDF) and the pesticide mirex (Ecology and Environment 2009; Table 1). Polycyclic aromatic hydrocarbons (PAHs) are

also present in the RE AOC. Although they are not listed as a known or suspected BUI, their concentrations and potential for adverse effects were evaluated in this study. The approach for addressing the BUI delisting criteria for the Genesee River portion of the RE AOC was two-fold:

1. Conduct mink habitat and population assessments.
2. Determine whether “Levels of [BUI contaminants] measured in the tissue of resident prey are below those known to be associated with mink reproductive failure.” Based on current knowledge, the BUI contaminants that might impair mink reproduction in the RE AOC are total PCBs, CDDs/CDFs and mercury. Pesticide residues (e.g., mirex) are not a concern for mink reproduction (Giesey et al. 1994), while the status of PAHs was established during this study by literature review and prey tissue analysis.

The approach of sampling mink prey, but not mink, was adopted for two reasons:

1. It was uncertain whether enough mink could be trapped in the study area to obtain a statistically defensible sample size for chemical analyses of their tissues.
2. Access by boat to trapping areas along the river during the icy winter trapping season would have been dangerous for the field crew.

Research questions

1. Are mink or their signs observed in the Genesee River portion of the RE AOC?
2. What is the extent and quality of mink habitat in the Genesee River portion of the RE AOC?
3. Are concentrations of PCBs, CDDs/CDFs, PAHs and mercury measured in the tissue of resident prey below those known to be associated with mink reproductive failure?

Methods

Mink and their signs in the lower Genesee River portion of the RE AOC

Twenty “black trakka” traps purchased from a supplier in New Zealand and marked with mink scent by the field crew were set out in likely mink microhabitats from August 7 through October 2, 2013 (Figure 2). These non-lethal traps are designed for animals to walk through a tunnel and leave foot prints on clean paper after stepping on inked paper. Traps and the muddy areas around them were checked once or twice weekly for mink prints.

Extent and quality of mink habitat in the lower Genesee River portion of the RE AOC

Google Earth and Pictometry.org images from the southern extent of continuous boat docks ~1.5 km upstream from Lake Ontario to the rapids ~0.5 km downstream from the Lower Falls of the Genesee River were examined to evaluate potential mink habitat in the study area. In August and September 2013 an experienced mink trapper and the project field crew leader made detailed habitat observations, by boat and on foot, within ~100m of the shoreline along the river (Figure 3). The U.S. Fish and Wildlife Service’s (USFWS) Habitat Suitability Index (HSI) model

for mink (Allen 1986) was used to estimate habitat suitability at 41 sites. In addition, the trapper gave an experience-based HSI for mink (i.e., likelihood of successfully trapping mink) for each site. Both indices were on scales of 0-1. During habitat suitability surveys each site also was checked for signs of mink (e.g., foot prints, scats, dens).

Stable isotope analysis to determine mink prey trophic levels

Stable isotopes of nitrogen are used to evaluate trophic webs of ecosystems to give lifetime, integrated estimates of trophic level for organisms (DeNiro and Epstein 1978, Cabana and Rasmussen 1994). ^{14}N has a stable, heavier isotope (^{15}N) which occurs naturally, and the heavier and lighter isotopes are differentially absorbed and metabolized by organisms (Fry 1991). Usually the lighter isotope is excreted preferentially, leading to a relative enrichment of the heavier isotope in organisms relative to their environment or diet. This enrichment is measurable through mass spectrometry, and is reported in parts per thousand ($\delta\text{‰}$) relative to a standard: $\delta X = [(R_{\text{sample}} - R_{\text{standard}}) - 1] \times 10^3$, where X is ^{15}N and R is the corresponding ratio of $^{15}\text{N}/^{14}\text{N}$. The standard for nitrogen is atmospheric nitrogen (Fry 1991).

Selective excretion of ^{14}N over ^{15}N by animals results in an increase of approximately 3.4‰ in the $\delta^{15}\text{N}$ at each trophic level; thus, ^{15}N analysis can determine the average trophic level at which an animal feeds (Peterson and Fry 1987, Cabana and Rasmussen 1994).

Trophic levels vary from 1 (herbivores) to 6 (apex predators.) Mink in riparian areas often eat amphibians, crayfish and fish (USEPA 1993). Three samples each of amphibians, crayfish, lower trophic level fish and upper trophic level fish were collected in the study area from 7 August 2013 through 2 August 2014. Frozen, composited, 10g samples of muscle tissue from amphibians (7-16 animals/sample), crayfish (52-73/sample), lower trophic level fish (10/sample) and upper trophic level fish (5/sample) were analyzed by the Cornell Stable Isotope Laboratory (COIL) for isotopic ratios of $^{15}\text{N}/^{14}\text{N}$ (δN) to determine average prey trophic levels.

PCB, CDD/CDF, PAH and mercury concentrations in the tissues of potential mink prey

Frozen, composited, >70g samples of each prey group sample (N=12: 4 prey types*3 samples each) were sent to ALS Global Environmental. Each of the 12 prey samples was homogenized, and separate aliquots were analyzed for total mercury (USEPA Method 1631app) and PAH (USEPA Method 8270D), PCB (USEPA Method 1668A) and CDD/CDF (USEPA Method 1613B) congeners in Kelso, WA or Houston, TX. Data were reported for 18 PAH congeners, seven of which had RFP ≥ 0.0000019 ; total PCBs, including 15 congeners with TEQ ≥ 0.01 ; and total CDD/CDF, including 15 congeners with TEQ ≥ 0.01 (Appendix C).

Mink hazard assessment

Concentrations of total mercury, total PCBs and total toxic equivalents (TEQ for PAHs, CDDs/CDFs and co-planar PCBs combined) found in mink prey were used to estimate the maximum potential dietary exposure of mink in the Genesee River portion of the RE AOC. TEQ (where 2,3,7,8-TCDD = 1) for CDD/CDF and PCB congeners were calculated using values from

Van den Berg *et al.* (2006). TEQ for PAH congeners was calculated using relative potency (REP) values from Villeneuve *et al.* (2002). TEQ was summed separately for CDD/CDF, PCBs and PAHs then all categories were summed to yield total TEQ for each prey group sample.

USEPA (1993) reported the results of 17 studies of mink diet at 25 different locations where the portion of the diet from aquatic sources ranged from 13.4% to 92%. The maximum potential exposure of mink to BUI contaminants in RE AOC water would be represented by the study on a river in lower Michigan (Alexander 1977 cited by USEPA 1993), consisting of 57.5% upper trophic level fish, 27.5% lower trophic level fish, 4% crustaceans and 3% amphibians (total 92% aquatic), and 8% “other” (birds, mammals, vegetation, and unidentified). We used these dietary percentages to represent a realistic “worst-case” dietary exposure to total mercury, total PCBs and total TEQ for mink in the RE AOC. We then averaged the results from the six most relevant diet studies (for mink living along rivers and streams) cited by USEPA (1993; studies averaged were Hamilton 1940, Korschgen 1958, Cowan and Reilly 1973, Alexander 1977a, b, and Burgess and Bider 1980). For each prey category, we averaged the proportion of that category from all six studies to get a “typical” proportion of the diet for that category. A “typical” riparian mink’s diet consists of 35.8% upper trophic level fish, 14.6% lower trophic level fish, 10.9% crustaceans and 8.7% amphibians, with a total of 65% from aquatic sources.

Dietary exposures of mink in the RE AOC were estimated by multiplying the average concentration of each BUI contaminant in each of the four prey groups by the corresponding portion of mink diet, and summing the results. We did these calculations twice: 1) for the worst-case diet Alexander (1977, in USEPA 1993), and 2) for the typical diet represented by the average of the six studies. Maximum estimated dietary exposures were then compared to published lowest observed adverse effect levels (LOAEL) reported by Haynes *et al.* (2007). The trophic levels calculated for each prey group were multiplied by that prey group’s proportion in the diet (the non-aquatic portion of each diet was assumed to be trophic level 1), and the results were summed to estimate the trophic levels of diets 1 and 2 above. The estimated dietary trophic levels were then used in a hazard estimate by comparison with known trophic levels of mink (hence diet) determined in the western RE AOC by Haynes *et al.* (2007).

Results

Mink and their signs in the lower Genesee River portion of the RE AOC

Although no mink walked through the traps and left inked tracks behind, definite evidence of mink was found 15 times (9.4% of 160 trap-checking days) on (muddy tracks) or around (foot prints) 10 of the 20 traps set throughout the study area (Figure 2). Three other sets of potential mink tracks near traps could not be identified definitively. One live mink, swimming across the river, was observed by the field crew (Appendix A).

Extent and quality of mink habitat in the lower Genesee River portion of the RE AOC

Much of the area within a minimum of 100m of the shoreline, on both the east and west banks of the river, appeared to be suitable mink habitat (Appendix B, Figure 3). USFWS HSI

model values averaged 0.85 ± 0.29 (standard deviation), with 1 as optimum habitat. According to the three criteria used in the USFWS HSI model (percent of surface water, percent vegetation cover within 30m of the shoreline, and percent shoreline cover within 1m of surface water) nearly all habitat in the study area was suitable for mink. Based on long experience the professional trapper rated average mink habitat suitability at 0.43 ± 0.24 . The trapper's lower scores ($P < 0.0001$, Paired T-Test; Statistix 2013) were based on steep, rocky slopes and evidence of much human disturbance (e.g., trails, fire pits, trash, fishing paraphernalia) along many sections of the shoreline.

Species composition and trophic levels of samples of potential mink prey

Three species of amphibians were collected: green frog, *Lithobates* (formerly *Rana*) *clamitans*, leopard frog, (*L. pipiens*) and American toad (*Anaxyrus americana*). Three species of crayfish were sampled: >96% were northern clearwater crayfish (*Orconectes propinguus*) and the rest were six white river crayfish (*Procambarus acutus acutus*) and one big river crayfish (*Cambarus robustus*). Lower trophic level fish species included in each composited sample were bluegill (*Lepomis macrochirus*), pumpkinseed (*L. gibbosus*) and yellow perch (*Perca flavescens*), while upper trophic level fish in each composited sample were northern pike (*Esox lucius*), largemouth bass (*Micropterus salmoides*) and smallmouth bass (*M. dolomieu*) (Table 2).

Mean trophic level ($\delta N \pm SD$) was 2.06 ± 0.08 for amphibians (AM), 3.72 ± 0.11 for crayfish (CR), 4.45 ± 0.06 for lower trophic level fish (LF), and 4.88 ± 0.05 for upper trophic level (UF) fish (Table 3; Appendix C). The trophic levels of the four groups were significantly different from each other ($P < 0.0001$, One-way ANOVA, Tukey's HSD; Statistix 2013). Separate aliquots of the tissue samples used to determine trophic level also were analyzed by ALS Global Environmental for total mercury and PAH, PCB and CDD/CDF congeners.

BUI contaminant concentrations (not lipid adjusted) in the tissue of potential mink prey

Total Mercury

Concentrations of mercury in the nine samples of amphibians, crayfish and lower trophic level fish (range: 65-302 ng/g) were below (13-61%) the dietary LOAEL for mercury (500 ng/g; Dansereau *et al.* 1999). Concentrations of mercury in the three upper trophic level fish samples (range: 517-600 ng/g); all exceeded (by $\leq 20\%$) the dietary LOAEL for total mercury (Table 3, Appendix C).

PAH Relative Potencies (REP=TEQ)

Concentrations of TEQ from PAHs (REPs from Villeneuve *et al.* 2002) in 11 of the 12 samples of amphibians, crayfish, lower trophic level fish and upper trophic level fish (range: nd-0.57 pg/g) were below (<6%) the dietary LOAEL for PAH TEQ (9.2 pg/g; Bursian *et al.* 2006). In one of the three upper trophic level fish samples (UF1) PAH TEQ (21.1 pg/g) exceeded (by 129%) the dietary LOAEL for REP/TEQ (Table 3, Appendix C).

Total PCB

Concentrations of total PCB in the 12 samples of amphibians, crayfish, lower trophic level fish and upper trophic level fish (range: 3,950-354,000 pg/g) were below (<37%) the dietary LOAEL for total PCB (960,000 pg/g; Bursian *et al.* 2006) (Table 3, Appendix C).

PCB TEQ

Concentrations of TEQ from PCBs in 11 of the 12 samples of amphibians, crayfish, lower trophic level fish and upper trophic level fish (range: 0.02-1.06 pg/g) were below (<12%) the dietary LOAEL for TEQ (9.2 pg/g; Bursian *et al.* 2006). In one of the three lower trophic level fish samples (LF2) PCB TEQ (9.51 pg/g) barely exceeded (3.4%) the dietary LOAEL for TEQ. One PCB congener (#126 with a toxic equivalency factor, TEF, of 0.1) was responsible for 96% (9.17 pg/g) of the PCB TEQ in sample LF2 (Table 3, Appendix C).

CDD/CDF TEQ

Concentrations of TEQ (calculated using World Health Organization TEFs from Van den Berg *et al.* 2006) from CDD/CDF in the 12 amphibian, crayfish, lower trophic level fish and upper trophic level fish samples (range: 0.02-1.17 pg/g) were below (<13%) the dietary LOAEL for TEQ (9.2 pg/g, Bursian *et al.* 2006) (Table 3, Appendix C).

Total TEQ

Total TEQ for 10 of the 12 samples of amphibians, crayfish, lower trophic level fish and upper trophic level fish (range: 0.21-2.16 pg/g) were below (<24%) the dietary LOAEL for TEQ (9.2 pg/g). Two samples, LF2 (10.01 pg/g) and UF1 (22.76 pg/g), exceeded dietary LOAEL for total TEQ by 9% and 147%, respectively. Samples LF2 and UF1 exceeded the 9.2 pg/g LOAEL for TEQ because of PCB 126 (95% of total TEQ) and PAHs (93% of total TEQ), respectively. On average across the three samples for each trophic level, neither lower trophic level fish (3.68 ± 5.48 pg/g) nor upper trophic level fish (8.95 ± 11.96 pg/g) exceeded the dietary LOAEL for total TEQ (Table 3, Appendix C).

Multivariate statistical analysis

Cluster Analysis: Concentrations of all BUI contaminants found in all samples were entered into the analysis. The dendrogram (Figure 4) shows the relative distances in Euclidean space among the 12 samples. Amphibian and crayfish samples grouped together closely, although AM3 and CR3 (collected in 2014 vs. 2013, and analyzed separately from the other two samples in each prey species) were slightly separated from samples AM1&2 and CR1&2, respectively. Samples UF1 (high PAH concentrations) and LF2 (high PCB 126 concentration) were clearly separated from samples UF2&3 and LF1&3, respectively.

Principal Component Analysis: Principal component axes PC1 and PC2 explained 60.9% and 23.5% of the variation in BUI contaminant composition among the 12 composited prey species samples, respectively, or 84.4% of all variation in the data set, a very robust result (Table 4a). Trophic level (-0.467), total mercury (-0.551 and CDD/CDF TEQ (-0.537) had strong

associations with PC axis 1, while PAH REP (-0.644) and PCB TEQ (0.714) had strong associations with PC axis 2 (Table 4b, Figure 5). Amphibian and crayfish samples occupied the same area of multivariate space because they had very similar principal component scores, and these taxa were not strongly associated with any of the eigenvectors (important variables contributing to the distribution of samples in multivariate space) shown in Figure 5. Lower trophic level fish samples LF1&3 and upper trophic level fish samples UF2&3 each occupied their own areas in multivariate space. While samples LF1&3 were not associated with any eigenvectors, samples UF2&3 were associated with the PCB TEQ eigenvector. As in the cluster analysis, samples LF2 and UF1 each occupied a separate area of multivariate space. Sample LF2 was pulled toward the PCB TEQ eigenvector by its high PCB 126 TEQ, and sample UF1 was pulled far toward the PAH REP eigenvector. All UF samples were associated with the trophic level, total mercury and CDD/CDF TEQ eigenvectors but high PAH REP pulled sample UF1 away from samples UF2&3. In sum, the five eigenvectors all were negatively associated with PC axis 1, four of the five were negatively associated with PC axis 2, and one was positively associated with PC axis 2. Samples LF2 (PCB TEQ) and UF1, UF2 and UF3 (total mercury) had particularly strong associations with one or both of PC1 and PC2 (Table 4c, Figure 5).

Mink hazard assessment

Assuming the “worst case” mink diet: The maximum estimated dietary exposure of mink in the Genesee River portion of the RE AOC would be 407 ng/g (82% of the dietary LOAEL—500 ng/g) for total mercury, 6.2 pg/g (69% of the dietary LOAEL—9.2 pg/g) for total TEQ, and 216,071 pg/g (23% of the dietary LOAEL—960,000 pg/g) for total PCBs. The trophic level of the “worst case” diet (using average trophic levels for each prey group) would be 4.3 (Table 5).

Assuming the “typical” mink diet: The estimated actual dietary exposure in the Genesee River portion of the RE AOC would be 243 ng/g (49% of the dietary LOAEL) for total mercury, 3.6 pg/g (39% of the dietary LOAEL) for total TEQ, and 125,184 pg/g (14% of the dietary LOAEL) for total PCBs. The trophic level of the typical mink diet (using average trophic levels for each prey group) would be 3.1 (Table 5).

Discussion

Trophic levels of potential mink prey and estimated mink diets

The mean trophic levels of our samples can be compared to previously measured trophic levels of Lake Ontario fish, although direct comparisons cannot be made as our samples were multiple species taken from the lower Genesee River. In Lake Ontario, the trophic levels of salmon (N= 23) averaged 5.04 ± 0.29 ($\delta N \pm SD$) and trophic levels of alewives (N=34) averaged 3.75 ± 0.18 , respectively (Elizabeth Damaske, former SUNY Brockport M.S. student, personal communication, 2005). In the Genesee River our upper trophic level fish (UF) samples measured 4.88 ± 0.05 , while lower trophic level fish (LF) measured 4.45 ± 0.06 .

The trophic level of the worst-case diet, using the weighted mean of the trophic levels of mink prey taken from the Genesee River, would be 4.3. The trophic level of mink in our previous study in the western portion of the RE AOC (Haynes *et al.* 2007) ranged from 2.71 to 4.97 with an average of 3.5, corresponding to dietary trophic levels between 1.71 and 3.97 with an average of 2.5. This indicates it is unlikely that mink actually consume the worst-case diet in the AOC.

The trophic level of the literature-based typical diet, using the trophic levels of Genesee River prey, is 3.1. This agrees well with estimates found in USEPA (1995) which reported estimates for mink prey ranging from 2.5 to 2.9. Furthermore, this estimate also agrees with the results of our previous study (Haynes *et al.* 2007), falling slightly above the average for RE AOC lakeshore mink, which had a dietary trophic level of 2.8. This indicates that our “typical” diet is a good and conservative estimate of what mink are actually consuming in the RE AOC.

BUIs: Loss of Fish and Wildlife Habitat; Degradation of Fish and Wildlife Populations

Habitat appears to be suitable for mink along the lower Genesee River shoreline in the RE AOC, which supports delisting of the Loss of Fish and Wildlife Habitat BUI. Substantial evidence of live mink was observed along the shoreline of the study area, which supports delisting of the Degradation of Fish and Wildlife Populations.

BUIs: Bird or Animal Deformities or Reproductive Problems, and Degradation of Fish and Wildlife Populations

Mercury

Amphibian, crayfish and lower trophic level fish samples were all well below the dietary LOAEL for total mercury (500 ng/g), while the mean concentration of total mercury in upper trophic level fish in the Genesee River portion of the RE AOC (567 ng/g) exceeded the dietary LOAEL by 13%. Assuming that mink consume the “worst-case” diet, the maximum potential dietary exposure to total mercury would be 407 ng/g, or 81% of the dietary LOAEL. Assuming that mink consume the “typical” diet, the estimated dietary exposure to total mercury would be 48% of the dietary LOAEL.

Total PCBs and Total TEQ

Total PCB concentrations in all mink prey samples were far below the dietary LOAEL, and total TEQ was far below the dietary LOAEL in 10/12 samples. One upper trophic level fish sample (N=5 fish) exceeded the dietary TEQ (REP) LOAEL for PAHs by 129% and one lower trophic level fish sample (N=10 fish) exceeded the dietary TEQ LOAEL for PCB TEQ by 3.4%. The average total TEQ for the three upper trophic level fish samples was 97% of the dietary LOAEL (9.2 pg/g), and the average total TEQ for the three lower trophic level fish samples was 40% of the dietary LOAEL.

Hazard Analysis

Even the worst-case estimated dietary exposure did not exceed dietary LOAELs for total mercury and total TEQ. While it is possible to construct a mink diet that would exceed the dietary LOAELs for these parameters, that diet would have a trophic level above 4.4 (all lower and upper trophic level fish in the Genesee River), which is highly unlikely (see above re: measured mink and prey trophic levels and our best estimate of mink diet in the RE AOC). Also, Haynes *et al.* (2007) found that the average concentrations of total mercury and CDD/CDF TEQ in mink caught in the Braddock Bay portion of the RE AOC, as well as in nearby sites in the Genesee River watershed and along Lake Ontario, were below the LOAEL for mink liver. These data, together with the mink prey data collected in this study, support delisting the Bird or Animal Deformities or Reproductive Problems BUI and the Degradation of Fish and Wildlife Populations BUI for mercury and total TEQ in the REAOC.

Evidence for point and non-point sources of BUI contaminants in the RE AOC

According to Albers (2003) and Rice *et al.* (2003), both PAHs and CDDs/CDFs are by-products of combustion, such as in internal combustion engines, home heating systems and waste incinerators. The output of CDD/CDF from municipal waste incinerators is usually dominated by octachlorodibenzo-p-dioxin (OCDD; Rice *et al.* 2003, Gilpin *et al.* 2003). This was true for all prey samples collected in the RE AOC, in which OCDD ranged from 49-78% of the total CDD/CDF. This result strongly suggests that the majority of CDD and CDF compounds are coming from waste incinerators dispersed across the regional landscape, which amount to a non-point source for the RE AOC that cannot be remediated within the RAP context.

Because fish in the six composited samples (3 LF, 3 UF) were collected throughout the lower Genesee River study area in different months, the high concentrations of all 12 PAHs analyzed and the associated high REP in only one upper trophic level sample (UF1, 5 fish) suggests that a single fish contained the high concentrations. Two explanations may account for this finding: At least one fish in sample UF1 either 1) was exposed to a point source of PAHs, or 2) had a poorly functioning liver with regard to PAH biotransformation and excretion. PAHs do not bioaccumulate like other BUI contaminants, because higher trophic level organisms (i.e., vertebrates) are generally efficient at metabolizing PAHs (Nakata *et al.* 2003). PAH concentrations in fish are usually low (Eisler 1987), and our data show no correlation between PAH concentrations and trophic level (Figure 6). Conversely, mercury, PCBs and CDD/CDF are bioaccumulated and positively correlated with trophic level (Figure 6).

Similarly, the abundance of PCB congener 126 in sample LF2 was probably due to one fish in that composite of 10 fish after exposure to a point source or inability of the fish's liver to metabolize or excrete the congener.

Recommendations

1. The Loss of Fish and Wildlife BUI delisting criterion relating to mink states that this aspect of the BUI can be considered unimpaired when it is demonstrated that:
 - Mink inhabit and reproduce within areas contiguous to the Genesee River and streams within the defined area; OR
 - Physical and biological habitat is suitable for mink.

This study found substantial evidence of live mink along the Genesee River portion of the RE AOC, and an evaluation of mink habitat using the USFWS mink Habitat Suitability Index Model found highly suitable habitat (85%) along the Genesee River shoreline of the AOC. These findings support the delisting of the Loss of Fish and Wildlife BUI. It is recommended that the Loss of Fish and Wildlife BUI be delisted if achievement of the other six delisting criteria for this BUI can also be demonstrated.

2. The Degradation of Fish and Wildlife Populations BUI delisting criterion relating to mink states that this aspect of the BUI can be considered unimpaired when it is demonstrated that:
 - Mink are present and are reproducing; OR
 - Levels of PCBs, dioxins/furans, mirex, and mercury measured in the tissue of resident mink prey are below those known to be associated with mink reproductive failure.

This study used mink diet composition assessments in literature to design a “worst-case diet” (trophic level 4.3) and a “typical diet” (trophic level 3.1) for mink in the Rochester Embayment AOC, and conducted a hazard assessment for each diet using the analytically-determined mean concentrations of BUI contaminants in the potential mink prey sampled in the AOC. Even the worst-case estimated dietary exposure did not exceed dietary LOAELs for total mercury and total TEQ. While it is possible to construct a mink diet that would exceed the dietary LOAELs for these parameters, that diet would have a trophic level above 4.4, which is highly unlikely because results of our previous study (Haynes *et al.* 2007) found that RE AOC lakeshore mink had a dietary trophic level of 2.8.

The findings of this study demonstrate the achievement of this BUI, and it is recommended that the Degradation of Fish and Wildlife Populations BUI be delisted.

3. The Bird and Animal Deformities and Reproductive Problems BUI delisting criterion is identical to the one stated above for Degradation of Fish and Wildlife Populations. It is the only delisting criterion for this BUI.

For the reasons stated above in #2, the findings of this study demonstrate the achievement of this BUI, and it is recommended that the Bird and Animal Deformities and Reproductive Problems BUI be delisted.

4. Gather information on the possible existence and determine the locations of sites near the study area with point sources of PAHs and PCB congener 126. If found, consider the feasibility of remediating the sites.

Acknowledgments

We thank Anthony Marsocci who led all field activities, tabulated data collected for the RE AOC project and created Figures 2 & 3; expert mink trapper Randall Baase who evaluated mink habitat quality and found signs of live mink in the study area; Katherine Bailey who performed the multivariate statistical analyses and created Figures 4-6; and the many people who assisted field collections: Kingdon Barrett, Jennifer Curry, Kimberly Engels, Andrea Graham, Steven Hart, Christopher Hayes, Justin Hulbert, Chelsea Lipp, Nicholas Marsocci, Kelly Owens, Matthew Pavilaitis, Brendan Ryan, David Sanderson-Kilchenstein, Noelle Sofee, Tanner Squires, Anthony Tornatore, Alexander Silva, Alyssa Vogel and Cassandra Wolfanger.

Literature Cited

- Albers, P. H. 2003. Petroleum and individual polycyclic aromatic hydrocarbons. Pages 341-371, *in* Hoffman, D.J., B.A Barnett, G.A.Burton, Jr., and J. Cairns, Jr. (Eds.). *Handbook of Ecotoxicology* (2nd Ed.). Boca Raton, FL: CRC Press LLC.
- Allen, A. W. 1986. Habitat Suitability Index Models: Mink, revised. U. S. Fish and Wildlife Service, Biological Report 82 (10.127). 23 pp.
- Bursian, S. J., C. Sharma, R. J. Aulerich, B. Yamini, R. R. Mitchell, C. E. Orazio, D. R. J. Moore, S. Svirsky, and D. E. Tillit. 2006. Dietary exposure of mink (*Mustela vison*) to fish from the Housatonic River, Berkshire County, Massachusetts, USA: Effects on reproduction, kit growth, and survival. *Environmental Toxicology and Chemistry* 25(6): 1533-1540.
- Cabana, G., and J. B. Rasmussen. 1994. Modeling food chain structure and contaminant bioaccumulation using stable nitrogen isotopes. *Nature* 372: 255-257.
- Dansereau, M., N. Lariviere, D. Du Tremblay, and D. Belanger. 1999. Reproductive performance of two generations of female semidomesticated mink fed diets containing organic mercury contaminated freshwater fish. *Archives of Environmental Contamination and Toxicology* 36: 221-226.
- DeNiro, M. J., and S. Epstein. 1978. Influence of diet on the distribution of carbon isotopes in animals. *Geochemica et Cosmochemica Acta* 42: 495-506.
- Ecology & Environment. 2009. Rochester Embayment Area of Concern Beneficial Use Impairment Delisting Criteria Report. Ecology & Environment, 368 Pleasant View Drive, Lancaster NY 14086.

- Eisler, R. 1987. Polycyclic aromatic hydrocarbon hazards to fish, wildlife, and invertebrates: a synoptic review. U. S. Fish and Wildlife Service, Patuxent Wildlife Research Center, Laurel, MD.
- Fry, B. 1991. Stable isotope diagrams of freshwater food webs. *Ecology* 72(6): 2293-2297.
- Giesy, J. P., D. A. Verbrugge, R. A. Othout, W. W. Bowerman, M. A. Mora, P. D. Jones, J. L. Newstead, C. Vandervoot, S. N. Heaton, R. J. Aulerich, S. J. Bursian, J. P. Ludwig, G. A. Dawson, T. J. Kubiak, D. A. Best, and D. E. Tillitt. 1994. Contaminants in fishes from Great Lakes-influenced section and above dams of three Michigan rivers. II: Implications for health of mink. *Archives of Environmental Contamination and Toxicology* 27:213-223.
- Gilpin, R.K., D.J. Wagel, and J.G. Solch. 2003. Production, distribution, and fate of polychlorinated dibenzo-*p*-dioxins, dibenzofurans, and related organohalogenes in the environment. Pages 55-87, in Schechter, A. and T.A. Gasiewicz. (Eds.), *Dioxins and Health*. Hoboken, NJ: Wiley-Interscience.
- Haynes, J. M., S. T. Wellman, and J. J. Pagano. 2007. RAP Progress in the Rochester Embayment of Lake Ontario: Population Monitoring, Trophic Relationships, and Levels of Bioaccumulative Chemicals of Concern in Mink, a Sentinel Species. Final report to the NYS Great Lakes Protection Fund. NYS Department of Environmental Conservation: Buffalo, NY. 146 pp.
- Nakata, H., Y. Sakai, T. Miyawaki, and A. Takemura. 2003. Bioaccumulation and toxic potencies of polychlorinated biphenyls and polycyclic aromatic hydrocarbons in tidal flat and coastal ecosystems of the Ariake Sea, Japan. *Environmental Science and Technology* 37: 3515-3521.
- Peterson, B. J., and B. Fry. 1987. Stable isotopes in ecosystem studies. *Annual Review of Ecology and Systematics* 18: 293-320.
- Rice, C. P., P. W. O'Keefe, and T. J. Kubiak. 2003. Sources, pathways, and effects of PCBs, dioxins, and dibenzofurans. Pages 501-573 in Hoffman, D. J., B. A. Barnett, G. A. Burton Jr., and J. Cairns Jr. (Eds.). 2003. *Handbook of Ecotoxicology* (2nd Ed.). Boca Raton, FL: CRC Press LLC.
- Statistix 10. 2013. Analytical Software. Tallahassee, FL.
- USEPA. 1993. Wildlife exposure factors handbook: Appendix: Literature review database. U. S. Environmental Protection Agency, Office of Research and Development. Washington, D.C. EPA/600/R-93/187b.
- USEPA. 1995. Great Lakes water quality criteria documents for the protection of wildlife. U. S. Environmental Protection Agency, Office of Research and Development. Washington, D.C. EPA/820/B-95/008.

- Van den Berg, M., L. S. Birnbaum, M. Denison, M. De Vito, W. Farland, M. Feeley, H. Fiedler, H. Hakansson, A. Hanberg, L. Haws, M. Rose, S. Safe, D. Schrenk, C. Tohyama, A. Tritscher, J. Tuomisto, M. Tysklind, N. Walker, and R. E. Peterson. 2006. The 2005 World Health Organization reevaluation of human and mammalian toxic equivalency factors for dioxins and dioxin-like compounds. *Toxicological Sciences* 93(2): 223-241.
- Villeneuve, D. L., J. S. Khim, K. Kannan and J. P. Giesy. 2002. Relative potencies of individual polycyclic aromatic hydrocarbons to induce dioxin-like and estrogenic responses in three cell lines. *Environmental Toxicology* 17: 128-137.

Table 1: Rochester Embayment AOC BUI Delisting Criteria related to mink (Ecology & Environment 2009).

BUI	BUI Status	Delisting Criteria
Bird or Animal Deformities or Reproductive Problems	Impaired	Mink are present and are reproducing, OR Levels of PCBs, dioxins/furans, mirex, and mercury measured in the tissue of resident prey are below those known to be associated with mink reproductive failure.
Degradation of Fish and Wildlife Populations	Impaired	SAME as above
Loss of Fish and Wildlife Habitat	Impaired	Mink inhabit and reproduce within areas contiguous to the Genesee River and streams within a defined area, OR Physical and biological habitat is suitable for mink.

Table 2: Sampling periods and species caught in the lower Genesee River portion of the RE AOC.

		7 Aug 2013	13-20 Aug 2013	23-28 Aug 2013	8-11 Sep 2013	15 Sep 2013	29 Sep- 5 Oct 2013	13-30 Jun 2014	26 Apr- 2 Aug 2014
Amphibians									
Green frog	<i>Lithobates clamitans</i>				8	9		2	
Leopard frog	<i>Lithobates pipiens</i>				4	7		3	
American toad	<i>Anaxyrus americana</i>				-	-		2	
Crayfish									
Northern clearwater	<i>Orconectes propinguus</i>		72				54		48
White river	<i>Procambarus a. acutus</i>		-				-		6
Big river	<i>Cambarus robustus</i>		1				-		-
Lower Trophic Level Fish									
Bluegill	<i>Lepomis macrochirus</i>	5		5		4			
Pumpkinseed	<i>Lepomis gibbosus</i>	4		3		5			
Yellow perch	<i>Perca flavescens</i>	1		2		1			
Upper Trophic Level Fish									
Northern pike	<i>Esox lucius</i>	1		1		1			
Largemouth bass	<i>Micropterus salmoides</i>	2		2		2			
Smallmouth bass	<i>Micropterus dolomieu</i>	2		2		2			

Table 3. Summary results [mean (standard deviation) of three samples] of mink prey chemical analysis. Item in bold exceeds LOAEL (lowest observed adverse effect level) from Villeneuve *et al.* 2002 and Van den Berg *et al.* 2006.

	Dietary LOAEL	Amphibian (SD)	Crayfish (SD)	Lower TL Fish (SD)	Upper TL Fish (SD)
Trophic Level (δN)		2.06 (0.08)	3.72 (0.11)	4.45 (0.06)	4.88 (0.05)
Total Mercury (ng/g)	500	114.67 (16.86)	74.13 (10.53)	272.00 (26.00)	567.33 (44.23)
Total PAH REP (pg/g)	9.2	0.32 (0.22)	0.18 (0.05)	0.04 (0.07)	7.06 (12.16)
Total Dioxin/Furan TEQ (pg/g)	9.2	0.15 (0.21)	0.08 (0.06)	0.30 (0.09)	1.09 (0.08)
Total PCB (pg/g)	960,000	4,810 (1,239)	23,900 (4,468)	88,233 (15,509)	331,667 (32,808)
Total PCB TEQ (pg/g)	9.2	0.20 (0.32)	0.21 (0.25)	3.34 (5.23)	0.80 (0.29)
Total REP/TEQ (pg/g)	9.2	0.67 (0.42)	0.47 (0.30)	3.68 (5.48)	8.95 (11.96)

Table 4a: Eigenvalues for principal component axes 1 and 2 of the Principal Component Analysis of chemicals of concern in mink prey sampled in the RE AOC.

PC Axis	Eigenvalue	Percent Variation	Cumulative % Variation
1	3.05	60.9	60.9
2	1.17	23.5	84.4

Table 4b: Eigenvectors (coefficients in the linear combinations of variations making up the variables contributing to principal component axes 1 and 2) for the Principal Component Analysis of chemicals of concern in mink prey sampled in the RE AOC.

Variable	PC1	PC2
Trophic Level	-0.467	0.240
Total Mercury	-0.551	-0.059
REP for PAHs	-0.334	-0.644
TEQ for Dioxins/Furans	-0.537	-0.120
TEQ for PCBs	-0.280	0.714

Table 4c: Principal component scores for 12 mink prey samples collected in the RE AOC. Large magnitude scores strongly influencing the analysis are in bold.

Sample	PCA Score 1	PCA Score 2
Amphibian 1	1.84	-0.496
Amphibian 2	1.82	-0.535
Amphibian 3	1.30	-0.495
Crayfish 1	1.25	-0.008
Crayfish 2	1.16	-0.007
Crayfish 3	1.06	-0.002
Lower Fish 1	0.188	0.186
Lower Fish 2	-0.923	2.32
Lower Fish 3	-0.010	0.115
Upper Fish 1	-3.30	-2.37
Upper Fish 2	-2.29	0.581
Upper Fish 3	-2.01	0.836

Table 5: Trophic levels and estimated dietary exposures based on average BUI contaminant concentrations in each prey group compared to the dietary LOAEL for each BUI contaminant.

Chemical	Worst-case diet	Typical diet	LOAEL
Mercury, total, ng/g	407	242	500
TEQ, total, pg/g	6.2	3.6	9.2
TEQ, PAHs, pg/g	4.1	2.4	
TEQ, dioxins/furans, pg/g	0.7	0.4	
TEQ, PCBs, pg/g	1.4	0.8	
PCBs, total, pg/g	216,072	125,184	960,000
Trophic level	4.3	3.1	

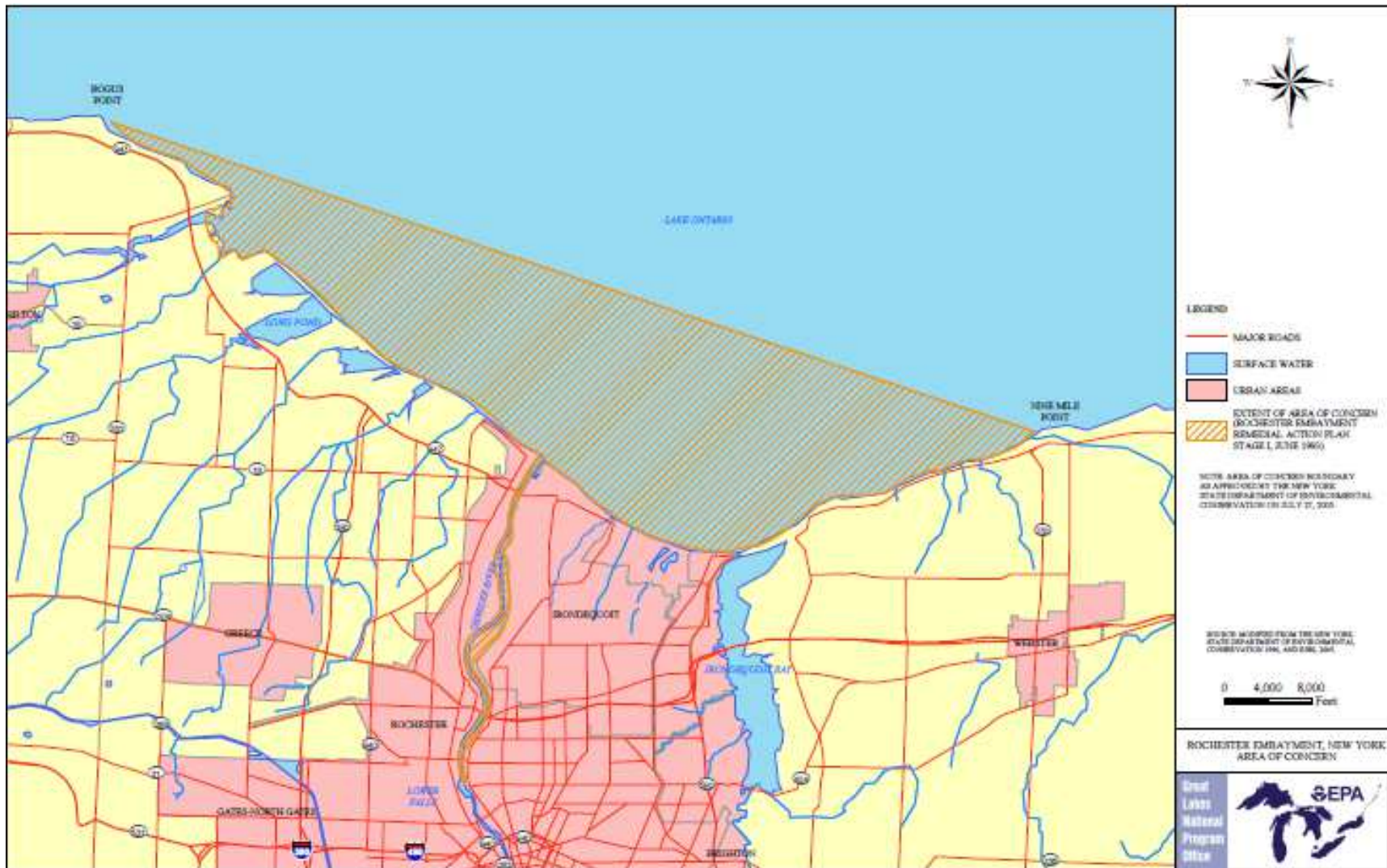


Figure 1: Rochester Embayment Area of Concern (tan). The lower Genesee River study area extends south from Lake Ontario into the City of Rochester (<http://www.epa.gov/glnpo/aoc/rochester/index.html>).

Number of Mink Signs 2013

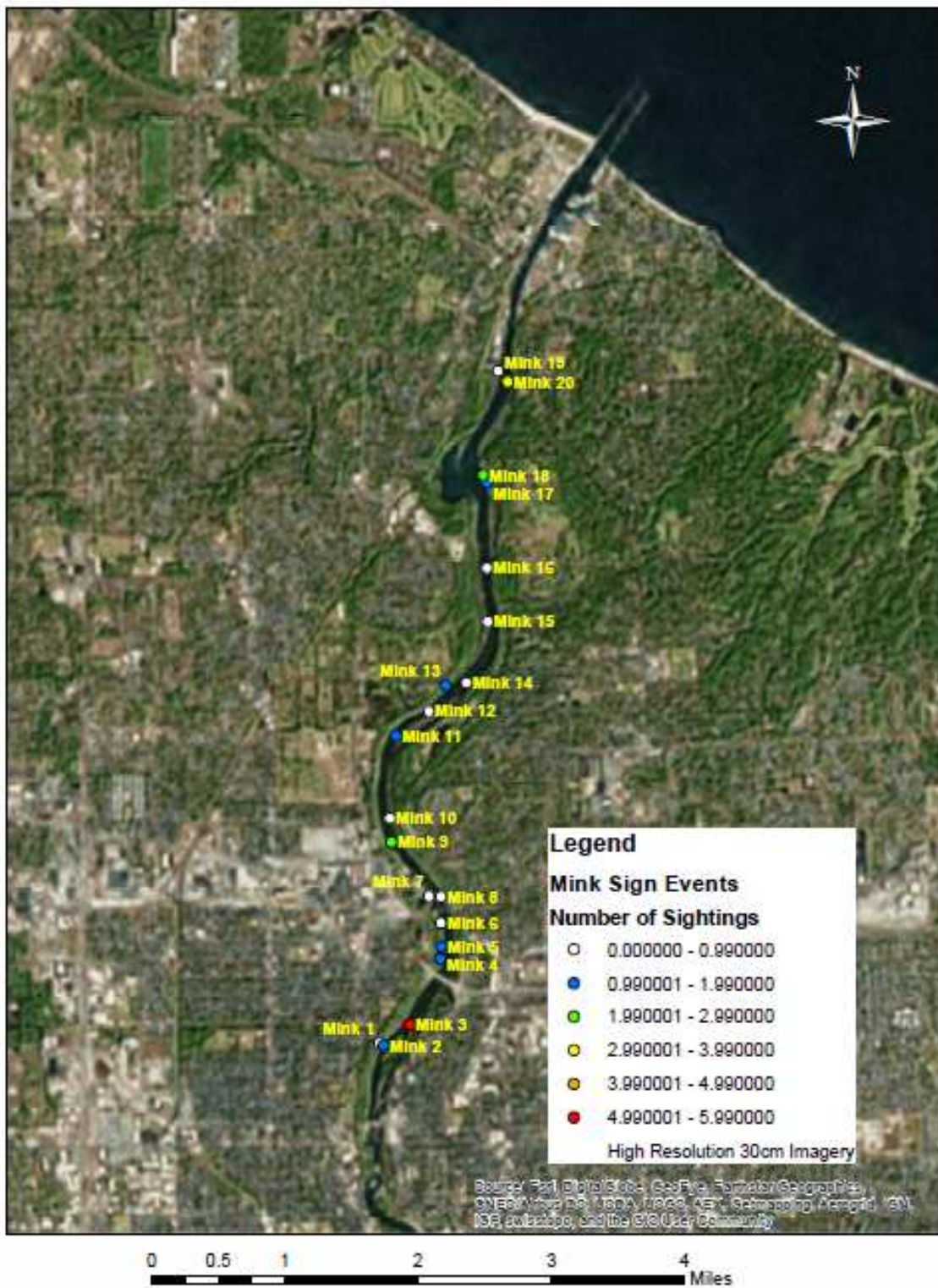


Figure 2: Sightings of mink and their signs in the lower Genesee River (see Appendix A).

Mink HSI 2013 Full Extent



Figure 3a: Mink habitat assessment locations in the lower Genesee River (see Appendix B).

Mink HSI 2013 Northern AOC

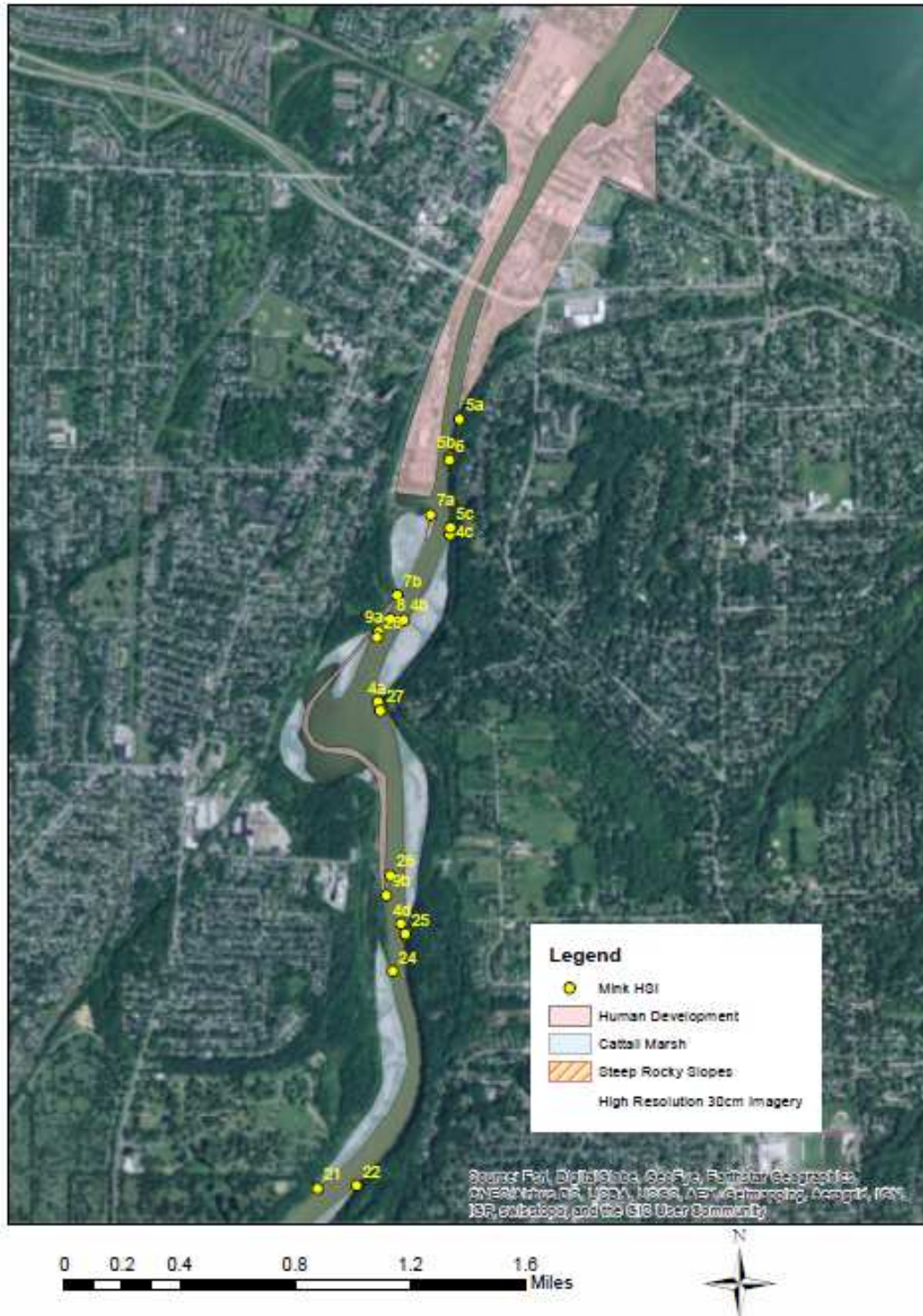


Figure 3b: Mink habitat assessment locations in the northern part of the lower Genesee River portion of the Rochester Embayment Area of Concern (see Appendix B).

Mink HSI 2013 Southern AOC

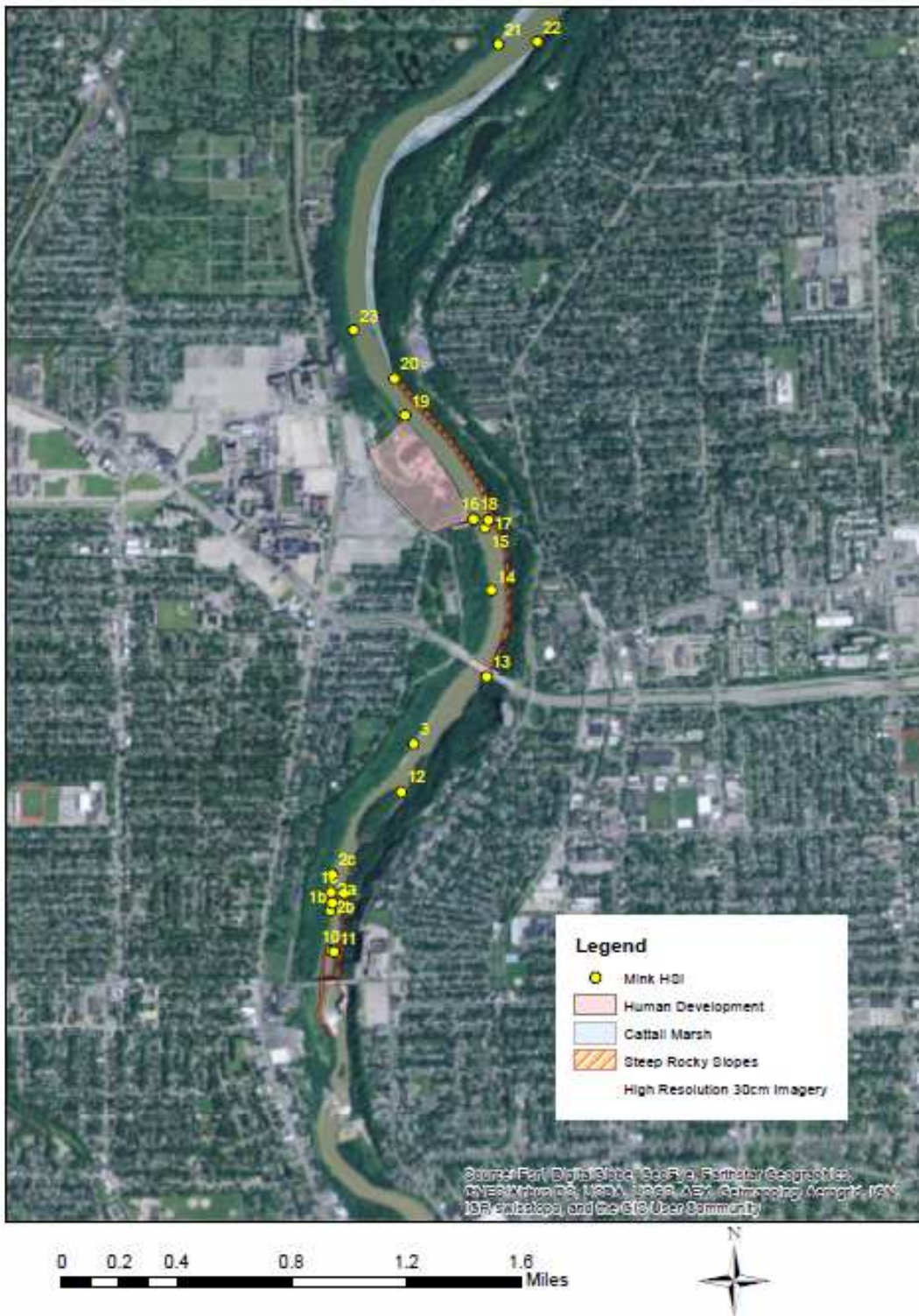


Figure 3c: Mink habitat assessment locations in the southern part of the lower Genesee River portion of the Rochester Embayment Area of Concern (see Appendix B).

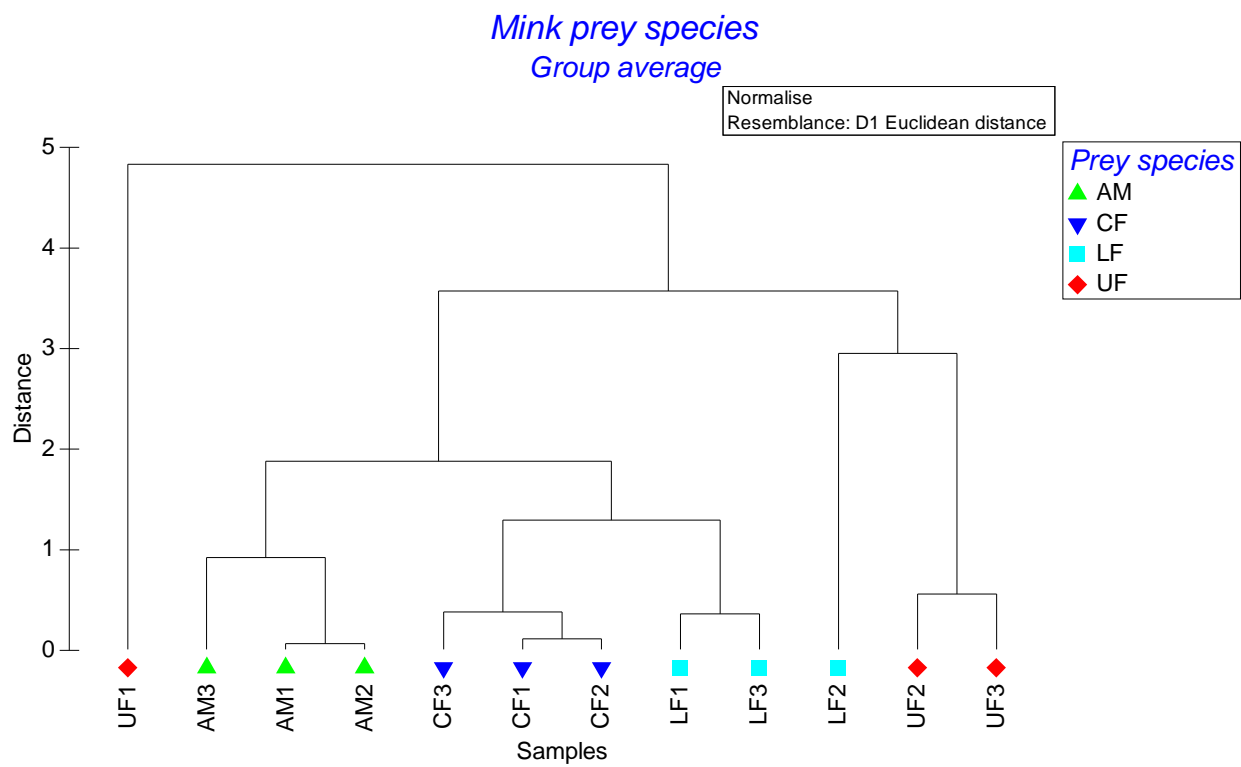


Figure 4: Cluster diagram of chemical relationships among 12 composited mink prey samples collected in 2013 and 2014.

Mink prey trophic guilds

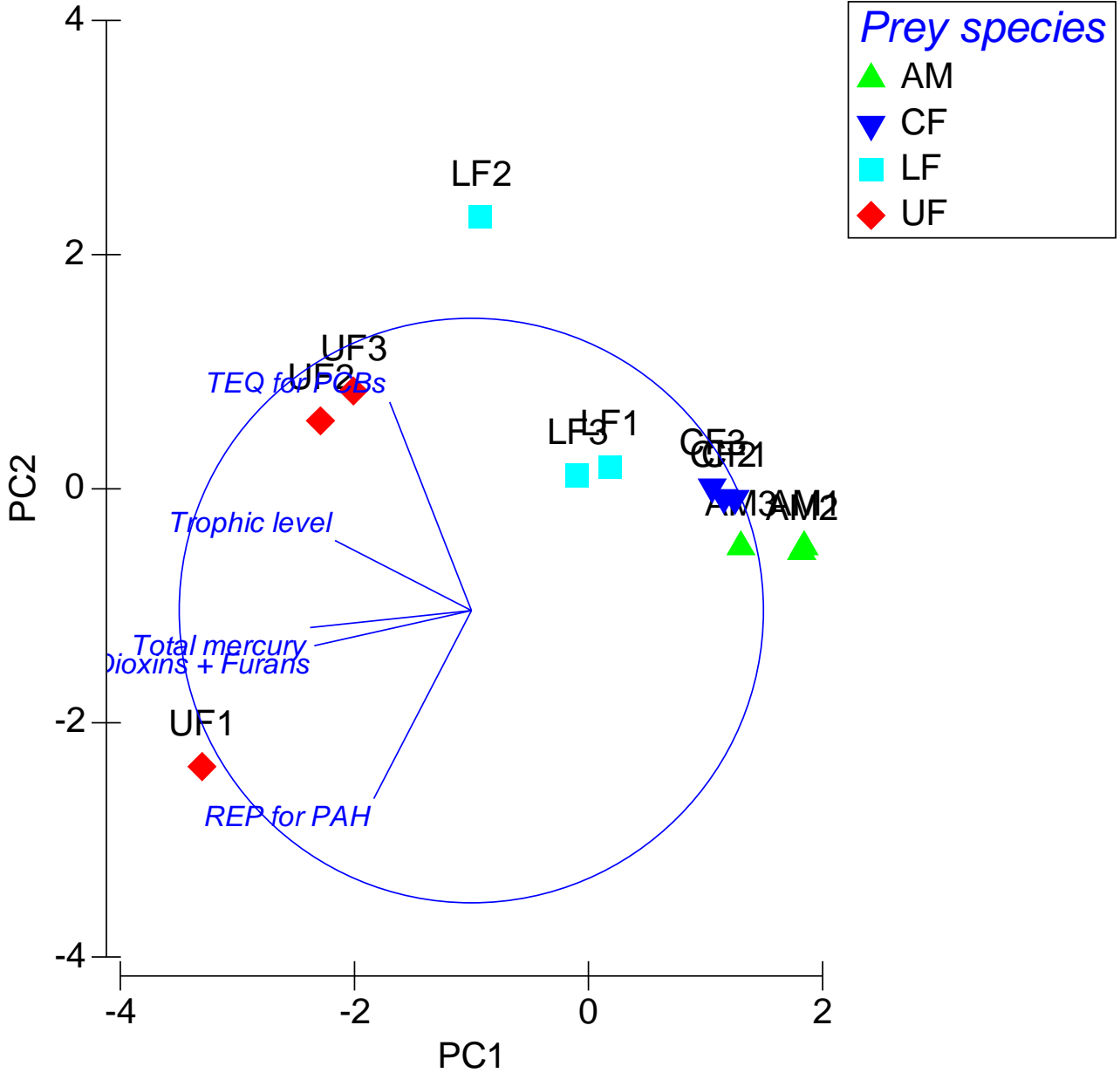


Figure 5: Principal component analysis diagram of chemical relationships in multivariate space among 12 composited mink prey samples collected in 2013 and 2014, and the variables (vectors) influencing the relationships.

Mink prey species

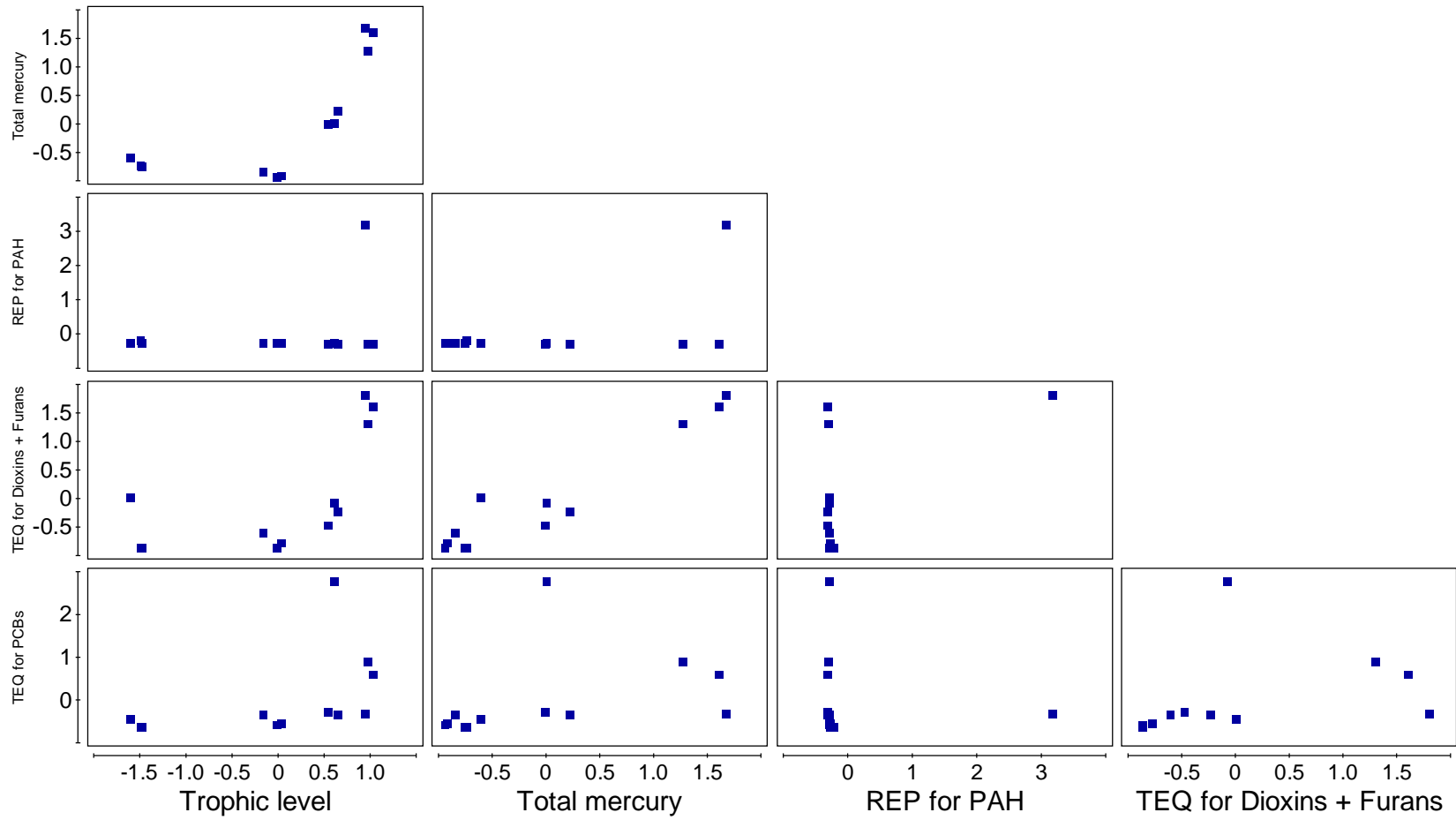


Figure 6: Relationships between pairs of variables in the multivariate analysis. Note the relatively strong relationships between PCB and CDD/CDF TEQs, and total mercury, but not for PAH REP, and trophic level.

Appendix A: Mink and their signs observed in the lower Genesee River portion of the RE AOC.

Mink Sign Observations	Date	Side of River	Site Description	Signs Observed
1	8/7/2013	East	Muddy Beach just before river branches around Seth Green Island	1. Trap surrounded by mink tracks
2	8/7/2013	West	Flat wooded area just before 104 bridge	2. Mink tracks on top of trap
3	8/7/2013	West	Last clump of trees upstream from Kodak WTP	3. Possible mink track on outside of trap
4	8/20/2013	East	Muddy Beach just before river branches around Seth Green Island	4. Multiple mink, raccoon, deer tracks in mud
5	8/20/2013	West	Last clump of trees upstream from Kodak WTP	5. Single mink track in mud 5m from trap
6	9/4/2013	East	Muddy Beach just before river branches around Seth Green Island	6. Multiple mink tracks in mud 5m from trap
7	9/4/2013	West	Small sandbar with scrubby vegetation next to wetland boat docks.	7. Possible mink track on trap ink card
8	9/8/2013	West	Small hardwood tree stand surrounded by cattail marsh	8. Multiple mink tracks in mud
9	9/11/2013	East	Muddy Beach just before river branches around Seth Green Island	9. Probable mink track in mud
10	9/25/2013	East	Muddy Beach just before river branches around Seth Green Island	10. Single mink track in mud
11	9/25/2013	West	Flat wooded area centered between Kodak bridge and 104 bridge	11. Multiple mink tracks in mud
12	9/25/2013	West	Small sandbar flat next to wetland boat docks. Scrub brush vegetation.	12. Multiple mink tracks in sand; two sizes
13	9/29/2013	East	Center of Seth Green Island. Flat wooded area with rocky beach	13. Multiple mink tracks on muddy beach
14	9/29/2013	East	Small hardwood tree stand surrounded by cattail marsh	14. Single mink track in mud
15	9/29/2013	East	Lots of woody debris and garbage accumulated in water on east side of turnaround basin. Trap on wooded shoreline	15. Multiple mink tracks in mud
16	10/2/2013	East	Small stream inlet entering east side of turnaround basin. Trap located ~100m inland along a creek bed.	16. Mink swimming across river
17	10/2/2013	East	Lots of woody debris and garbage accumulated in water on east side of turnaround basin. Trap on wooded shoreline	17. Multiple mink tracks in mud
18	10/2/2013	West	Small sandbar flat next to wetland boat docks. Scrub brush vegetation	18. Multiple mink tracks in sand

Appendix B: Extent and quality of mink habitat at 41 sites along the lower Genesee River portion of the RE AOC. USFWS HSI calculated as shown by Allen (1986). Trapper HSI estimated from 40 years of experience.

GPS Latitude	GPS Longitude	% Surface Water	% Vegetation Cover (30m)	% Shoreline Cover (1m)	USFWS HSI	Trapper HSI	Site # / Notes / Observations
4311041	7737687	100	100	10	0.32	0.30	1. ~200m downstream from Ave E bridge. No wetlands here. Sandy beach with rock rubble and woody debris.
4311058	7737650	100	95	50	0.69	0.30	2. ~150m downstream from Ave E Bridge. Gravelly beach with emergent wetland vegetation.
4311060	7737689	100	100	30	0.55	0.10	3. ~250m downstream from Ave E bridge. Rocky beach, logs, steep ~5 m embankment ~8 m from shore. Lots of trash.
4310138	7737440	100	100	30	0.55	0.20	4. ~3m cliff right at shoreline. Heavily shaded by trees, fallen logs and sparse vegetation.
4311036	7737687	100	100	80	0.89	0.30	5. Muddy beach, <i>Phragmites</i> , shrubs and tree cover.
4311099	7737686	100	100	100	1.00	0.60	6. 100% <i>Phragmites</i> , full overhead cover. ~100m cliff about ~50 m inland.
4311387	7737440	100	100	95	0.97	0.50	7. Shoreline is solid cattail with small vegetated islands of trees and shrubs. Solid ground ~10m wide. Cattail marsh ~30m wide and backs to a steep forested slope.
4313508	7736932	100	100	100	1.00	0.60	8. End of east side cattail.
4313995	7737002	100	100	100	1.00	0.50	9. 100% cattail with occasional canals about 8 feet wide. Backed up by steep forested slope after ~100m.
4314175	7736923	100	100	100	1.00	0.50	10. 100% cattail with occasional canals about 8 feet wide. Backed up by steep forested slope after ~100m.
4314363	7736786	100	100	100	1.00	0.50	11. 100% cattail with occasional canals about 8 feet wide. Backed up by steep forested slope after ~100m.
4314377	7736785	100	100	100	1.00	0.60	12. South end of habitat with dense vegetation. Steep rocky banks with plenty of fallen tree branches and other cover.
4314531	7736786	100	100	100	1.00	0.60	13. Center of habitat, rocky shelves along shoreline.
4314615	7736756	100	100	100	1.00	0.60	14. North end of eastern wooded habitat. Steep, rocky banks.

GPS Latitude	GPS Longitude	% Surface Water	% Vegetation Cover (30m)	% Shoreline Cover (1m)	USFWS HSI	Trapper HSI	Site # / Notes / Observations
4314525	7736787	100	90	100	0.95	0.60	15. Small wetland bay at the end of boat docks. Mostly shrub with human development.
4314405	7736843	100	100	100	1.00	0.60	16. Beginning of cattail marsh. Rocky shoreline about 10m behind cattails
4314229	7736944	100	100	100	1.00	0.60	17. End of cattail marsh.
4314177	7736964	100	90	50	0.67	0.40	18. Walkway bridge near turning basin. Rocky bare soil around shoreline. About 50m long. Lots of garbage
4314149	7737001	100	100	100	1.00	0.60	19. Start of cattail marsh surrounding the turning basin. Cattail marsh for about 80m then cliff with houses above.
4313570	7736977	100	100	100	1.00	0.60	20. End of the west side cattail marsh at the Stephen F. Roman concrete unloading dock.
4310931	7737689	100	100	100	1.00	0.20	21. ~30m cliffs at base of the Lower Falls. Lots of fallen trees and huge boulders.
4370928	7737681	100	100	100	1.00	0.20	22. End of rocky section between Lower Falls and Seth Green Island. Too many humans to be good mink habitat.
4311281	7737477	100	100	100	1.00	0.30	23. Seth Green Island. Muddy/rocky shore with fallen logs, overhead cover and lots of poison ivy.
4311534	7737220	100	0	0	0.00	0.00	24. 104 Bridge (Landmark Location)
4311725	7737205	100	100	100	1.00	0.80	25. POI: mink "Point of Interest" between Kodak railroad bridge and 104 bridge.
4311863	7737224	100	100	100	1.00	0.60	26. Kodak railroad bridge.
4311882	7737259	100	100	95	0.97	0.80	27. ~30 m of flat wooded area along shore with dense vegetation, plenty of cover and wet solid ground. See POI #14
4311880	7737214	100	100	100	1.00	0.80	28. Heavily wooded area with branches overhanging water's edge. Relatively steep slope with plenty of cover and possible mink denning sites. Ground dry and brushy. Rocky near water's edge.
4311882	7737259	100	100	0	0.00	0.00	29. POI: Kodak bridge and storm sewer outlet. North of #16. (For photos see #15)
4312110	7737466	100	0	0	0.00	0.00	30. Industrial infrastructure From #18 to Kodak King's Landing Wastewater Treatment Facility

GPS Latitude	GPS Longitude	% Surface Water	% Vegetation Cover (30m)	% Shoreline Cover (1m)	USFWS HSI	Trapper HSI	Site # / Notes / Observations
4312191	7737498	100	90	100	0.95	0.50	31. Heavily wooded area with branches overhanging water's edge. Relatively steep slope with plenty of cover and possible mink denning sites. Ground dry and brushy. Rocky near water's edge.
4312927	7737184	100	100	95	0.97	0.50	32. Steep heavily wooded slope.
4312933	7737066	100	100	100	1.00	0.60	33. Cattail marsh for ~30m with occasional small wooded islands, then steep wooded slope.
4312298	7737621	100	100	60	0.77	0.00	35. POI: waterfall between #19 and #21. Steep ~50m concrete storm sewer pipeline.
4313404	7736958	100	100	100	1.00	0.60	36. Cattail marsh ~50m wide backing up to steep wooded slope.
4313486	7736919	100	90	95	0.92	0.60	37. Steep wooded slope. Potential mink runways on flat rocks on shoreline
4313613	7736964	100	80	80	0.80	0.00	39. POI: Stephen F. Roman concrete delivery dock.
4313976	7736995	100	100	100	1.00	0.60	40. Cattail marsh ~30m wide backing up to steep wooded slope.
4314136	7737004	100	100	100	1.00	0.40	41. Turnaround basin with cattail marsh 10-40m wide, backing up to steep wooded slope with large docks and walking trail.
					Avg. HSI	0.85	0.44
					SD	0.29	0.24

Appendix C: Total mercury, PAH relative potencies (REP, Villeneuve *et al.* 2002), and CCD, CDF and PCB toxic equivalents (TEQ, Van den Berg *et al.* 1998) for twelve composite tissue samples of potential mink prey from the Genesee River portion of the RE AOC.

LOAELs taken from Dansereau <i>et al.</i> (1999)-Hg and Bursian <i>et al.</i> (2006)	Sample ID	AM1	AM2	AM3	CR1	CR2	CR3	LF1	LF2	LF3	UF1	UF2	UF3
	ALS #	-003	-004	-19.00	-001	-002	-20.00	-005	-006	-007	-008	-009	-010
	δN	7.19	7.12	6.70	12.80	12.95	12.22	14.89	15.18	15.28	16.42	16.76	16.57
	Trophic level	2.12	2.10	1.97	3.76	3.81	3.59	4.38	4.46	4.50	4.83	4.93	4.87
	% Lipid/100	0.013	0.015	0.056	0.011	0.012	0.081	0.027	0.027	0.033	0.020	0.020	0.024
Compounds													
Mercury, total (ng/g)		103.0	107.0	134.0	65.6	70.9	85.9	256	258	302	600	585	517
Dietary Hg LOAEL = 500 ng/g													
Total TEQ from PAH, CDD&F, PCB		0.29	0.61	1.12	0.21	0.38	0.80	0.47	10.01	0.55	22.76	1.93	2.16
Dietary (Prey) TEQ LOAEL = 9.2 pg/g													
PAHs (ug/kg=ng/g)	REPs												
Naphthalene		1.10	0.76	0.72	1.60	0.85	0.60	2.20	2.20	2.90	13.00	1.80	2.70
2-Methylnaphthalene		0.90	0.74	1.00	1.30	0.86	0.94	2.20	2.80	2.60	6.40	2.10	2.10
Acenaphthylene		ND	0.92	0.28	ND	ND	0.74	0.38	0.34	0.39	23.00	0.64	0.75
Acenaphthene		0.34	0.60	0.38	1.40	1.30	0.98	2.30	3.00	3.40	52.00	2.80	3.00
Dibenzofuran		0.45	0.43	0.42	ND	0.71	0.66	1.10	1.60	1.70	34.00	1.50	1.30
Fluorene		0.37	0.48	0.63	0.89	1.20	0.68	2.10	2.90	3.40	63.00	2.70	3.10
Phenanthrene		1.30	1.70	1.60	4.80	8.00	4.90	5.50	7.00	8.20	400.00	5.50	6.60
Anthracene		ND	0.53	1.00	0.69	0.94	0.53	0.76	0.86	1.50	190.00	0.85	1.20
Fluoranthene		2.40	7.10	3.20	6.00	9.20	7.30	3.70	5.00	6.10	610.00	3.30	5.80
Pyrene		2.50	11.00	2.50	4.30	6.60	4.70	1.10	1.50	6.30	380.00	1.00	3.10
Benz(a)anthracene	1.90E-06	1.40	6.20	1.00	0.98	1.70	1.20	ND	0.39	1.00	290.00	ND	1.10
Chrysene	2.30E-06	2.70	7.40	2.30	1.40	2.90	2.80	ND	1.40	1.00	260.00	ND	1.00
Benzo(b)fluoranthene	5.10E-06	2.50	8.20	2.30	1.50	2.80	2.20	ND	0.54	0.50	270.00	0.28	1.50
Benzo(k)fluoranthene	1.40E-04	1.40	3.00	0.77	0.78	1.30	0.82	ND	0.75	ND	110.00	ND	0.44
Benzo(a)pyrene	1.60E-06	1.50	5.50	1.40	0.87	1.80	2.50	ND	ND	ND	260.00	ND	0.97

LOAELs taken from Dansereau et al. (1999)-Hg and Bursian et al. (2006)	Sample ID	AM1	AM2	AM3	CR1	CR2	CR3	LF1	LF2	LF3	UF1	UF2	UF3
	ALS #	-003	-004	-19.00	-001	-002	-20.00	-005	-006	-007	-008	-009	-010
	δN	7.19	7.12	6.70	12.80	12.95	12.22	14.89	15.18	15.28	16.42	16.76	16.57
	Trophic level	2.12	2.10	1.97	3.76	3.81	3.59	4.38	4.46	4.50	4.83	4.93	4.87
	% Lipid/100	0.013	0.015	0.056	0.011	0.012	0.081	0.027	0.027	0.033	0.020	0.020	0.024
Compounds													
Indeno(1,2,3-cd)pyrene	1.50E-05	1.70	4.30	1.20	0.91	1.90	1.40	ND	0.32	ND	170.00	ND	0.79
Dibenz(a,h)anthracene	4.60E-06	0.27	1.00	0.28	ND	0.60	0.30	ND	ND	ND	45.00	ND	ND
Benzo(g,h,i)perylene		1.60	3.60	0.87	0.70	1.70	1.50	ND	0.34	ND	120.00	ND	0.76
REPs from PAHs (Dietary [Prey] TEQ LOAEL = 9.2 pg/g)		0.25	0.57	0.15	0.14	0.24	0.16	0.00	0.12	0.01	21.10	0.00	0.09
CDDs and CDFs (ng/kg = pg/g)	WHO TEFs												
1,2,3,4,6,7,8-Heptachlorodibenzofuran (HpCDF)	0.01	0.44	0.35	0.21	0.41	0.49	0.22	0.25	0.26	0.28	0.458	0.48	0.29
1,2,3,4,6,7,8-Heptachlorodibenzo-p-dioxin (HpCDD)	0.01	1.61	1.72	0.43	1.28	1.65	0.42	1.22	1.32	1.48	2.27	1.80	2.54
1,2,3,4,7,8,9-Heptachlorodibenzofuran (HpCDF)	0.01	ND	ND	ND	ND	ND	0.06	ND	ND	ND	ND	ND	ND
1,2,3,4,7,8-Hexachlorodibenzofuran (HxCDF)	0.1	ND	ND	0.07	ND	ND	0.05	ND	ND	ND	ND	ND	ND
1,2,3,4,7,8-Hexachlorodibenzo-p-dioxin (HxCDD)	0.1	ND	ND	0.09	ND	ND	ND	ND	ND	ND	ND	ND	ND
1,2,3,6,7,8-Hexachlorodibenzofuran (HxCDF)	0.1	ND	ND	0.06	ND	ND	ND	ND	ND	ND	ND	ND	ND
1,2,3,6,7,8-Hexachlorodibenzo-p-dioxin (HxCDD)	0.1	ND	ND	0.11	ND	ND	0.08	ND	ND	ND	0.34	ND	0.35
1,2,3,7,8,9-Hexachlorodibenzofuran (HxCDF)	0.1	ND	ND	ND	ND	0.38	ND	1.85	3.32	2.36	8.24	8.01	8.28
1,2,3,7,8,9-Hexachlorodibenzo-p-dioxin (HxCDD)	0.1	ND	ND	0.06	ND	ND	ND	ND	ND	ND	ND	ND	ND
1,2,3,7,8-Pentachlorodibenzofuran (PeCDF)	0.03	ND	ND	0.04	ND	ND	ND	ND	ND	ND	ND	ND	ND
1,2,3,7,8-Pentachlorodibenzo-p-dioxin (PeCDD)	1	ND	ND	0.13	ND	ND	0.10	ND	ND	ND	ND	ND	ND
2,3,4,6,7,8-Hexachlorodibenzofuran (HxCDF)	0.1	ND	ND	0.06	ND	ND	0.05	ND	ND	ND	ND	ND	ND
2,3,4,7,8-Pentachlorodibenzofuran (PeCDF)	0.3	ND	ND	0.09	ND	ND	ND	ND	ND	ND	0.38	0.35	ND
2,3,7,8-Tetrachlorodibenzofuran (TCDF)	0.1	ND	ND	0.09	ND	ND	0.21	ND	0.32	0.59	1.62	1.47	1.16
2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD)	1	ND	ND	0.17	ND	ND	ND	ND	ND	ND	ND	ND	ND

LOAELs taken from Dansereau et al. (1999)-Hg and Bursian et al. (2006)	Sample ID	AM1	AM2	AM3	CR1	CR2	CR3	LF1	LF2	LF3	UF1	UF2	UF3
	ALS #	-003	-004	-19.00	-001	-002	-20.00	-005	-006	-007	-008	-009	-010
	δN	7.19	7.12	6.70	12.80	12.95	12.22	14.89	15.18	15.28	16.42	16.76	16.57
	Trophic level	2.12	2.10	1.97	3.76	3.81	3.59	4.38	4.46	4.50	4.83	4.93	4.87
	% Lipid/100	0.013	0.015	0.056	0.011	0.012	0.081	0.027	0.027	0.033	0.020	0.020	0.024
Compounds													
Octachlorodibenzofuran (OCDF)	0.0003	1.40	0.86	0.18	0.91	1.43	0.39	0.86	1.30	1.20	1.22	0.98	1.25
Octachlorodibenzo-p-dioxin (OCDD)	0.0003	13.20	8.74	2.49	12.00	17.80	5.74	7.72	11.00	13.00	27.50	15.00	19.60
Heptachlorodibenzo-p-dioxins (HpCDD), Total		11.10	0.71	0.33	1.06	1.35	1.06	0.71	2.37	2.48	4.02	1.80	2.54
Heptachlorodibenzofurans (HpCDF), Total		1.27	0.90	ND	0.41	0.88	ND	0.75	ND	0.88	1.12	1.40	0.99
Hexachlorodibenzo-p-dioxins (HxCDD), Total		ND	ND	0.24	ND	ND	0.18	ND	ND	ND	ND	ND	0.35
Hexachlorodibenzofurans (HxCDF), Total		ND	ND	0.06	ND	0.38	0.05	2.52	3.96	2.84	9.39	9.41	8.28
Pentachlorodibenzo-p-dioxin (PeCDD), Total		ND	ND	0.07	ND	ND	0.10	ND	ND	ND	ND	ND	ND
Pentachlorodibenzofurans (PeCDF), Total		ND	ND	ND	0.79	1.57	0.68	1.54	ND	ND	ND	0.66	ND
Tetrachlorodibenzo-p-dioxins (TCDD), Total		ND	ND	ND	ND	0.62	0.22	ND	ND	ND	ND	ND	ND
Tetrachlorodibenzofurans (TCDF), Total		ND	ND	ND	0.66	ND	0.46	1.06	0.32	ND	2.97	0.59	ND
Total Dioxins and Furans		26.97	11.21	3.37	15.83	24.03	8.87	15.16	18.95	20.40	46.22	29.83	33.01
TEQs from Dioxins and Furans		0.02	0.02	0.42	0.02	0.06	0.14	0.20	0.38	0.31	1.24	1.15	1.01
Dietary (Prey) TEQ LOAEL = 9.2 pg/g													
PCBs (ng/kg = pg/g)	WHO TEFs												
Monochlorobiphenyls, Total		ND	ND	ND	6.09	ND	ND	ND	ND	ND	ND	ND	2.55
Dichlorobiphenyls, Total		ND	ND	52.40	202.00	124.00	69.90	ND	38.60	ND	ND	167.00	185.00
Trichlorobiphenyls, Total		172	132	78	865	951	509	2230	2870	3130	5030	3700	3870
Tetrachlorobiphenyls, Total		778	720	310	3900	3680	2950	11400	14000	14900	42000	36600	43000
Pentachlorobiphenyls, Total		1030	923	1020	5410	6210	9530	22900	28800	22000	103000	94300	104000
Hexachlorobiphenyls, Total		1060	1020	2160	4560	7420	9630	25700	30600	18900	111000	91100	109000
Heptachlorobiphenyls, Total		837	805	1680	2830	5260	3650	17400	21100	11400	66000	50200	64200
Octachlorobiphenyls, Total		230	235	671	826	1320	910	4180	5800	3010	20800	13200	16600
Nonachlorobiphenyls, Total		98	84	193	208	234	218	1120	1300	804	4150	3280	4180
Total PCBs		4250	3950	6230	18900	25300	27500	85300	105000	74400	354000	294000	347000

LOAELs taken from Dansereau et al. (1999)-Hg and Bursian et al. (2006)	Sample ID	AM1	AM2	AM3	CR1	CR2	CR3	LF1	LF2	LF3	UF1	UF2	UF3
	ALS #	-003	-004	-19.00	-001	-002	-20.00	-005	-006	-007	-008	-009	-010
	δN	7.19	7.12	6.70	12.80	12.95	12.22	14.89	15.18	15.28	16.42	16.76	16.57
	Trophic level	2.12	2.10	1.97	3.76	3.81	3.59	4.38	4.46	4.50	4.83	4.93	4.87
	% Lipid/100	0.013	0.015	0.056	0.011	0.012	0.081	0.027	0.027	0.033	0.020	0.020	0.024
Compounds													
Dietary (Prey) TPCBs LOAEL = 960,000 pg/g													
PCB 105	0.00003	79.90	71.30	82.80	404.00	520.00	899.00	1940.00	2350.00	1580.00	ND	6030.00	6950.00
PCB 114	0.00003	8.84	7.23	15.20	ND	ND	73.80	91.40	116.00	58.50	37.90	378.00	451.00
PCB 118	0.00003	365.00	390.00	364.00	1110.00	1550.00	2520.00	5590.00	7230.00	4890.00	145.00	17200.00	22400.00
PCB 123	0.00003	6.37	7.93	11.40	ND	8.43	51.40	56.10	75.70	51.00	307.00	212.00	265.00
PCB 126	0.1	ND	ND	5.58	ND	ND	3.65	ND	91.70	ND	ND	ND	ND
PCB 167	0.00003	24.10	23.30	54.50	69.80	114.00	179.00	306.00	378.00	234.00	23.60	1040.00	1280.00
PCB 169	0.03	ND	ND	ND	ND	ND	ND	ND	ND	ND	13.40	ND	ND
PCB 189	0.00003	5.35	ND	14.00	12.70	23.90	26.90	68.80	80.00	38.80	ND	178.00	213.00
PCB 77	0.0001	4.49	ND	ND	33.50	27.10	23.50	53.60	68.60	73.40	227.00	160.00	142.00
PCB 81	0.0003	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	14.70	ND
PCBs 156 + 157	0.00003	52.60	49.90	110.00	191.00	247.00	501.00	794.00	970.00	589.00	1670.00	2680.00	3290.00
TEQs from PCBs (Dietary (Prey) TEQ LOAEL = 9.2 pg/g)		0.02	0.02	0.58	0.06	0.08	0.49	0.27	9.52	0.23	0.49	0.85	1.06